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1 Introduction

Faba bean (*Vicia faba* L.) is one of the most important legumes with high nutritional value for humans and animals, due to its richness in many nutrients such as proteins, starch, dietary fibers, fatty acids, vitamins and minerals.^{1,2} In addition, faba bean is considered a rich source of micronutrients called phytonutrients, which are secondary metabolites accumulated in different parts of plants.³ These phytonutrients are bioactive compounds that act as natural antioxidants due to their high content of many functional phenolic and flavonoid compounds such as tannins, proanthocyanidins, L-3,4-dihydrophenylalanine (L-dopa), flavonols and flavones.^{4,5} In humans, natural antioxidants can protect biological molecules such as

Faba beans with enhanced antioxidant activity ameliorate acetic acid-induced colitis in experimental rats

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Faba beans are among the legumes that are of the greatest importance due to their high nutritional value. In addition to the essential nutrients that faba beans contain, they also contain bioactive compounds such as phenolics and flavonoids that are considered as potent natural antioxidants. Ulcerative colitis (UC) is an inflammatory bowel disease in which oxidative stress plays an essential role in the pathophysiology. The aim of the current study was to evaluate the antioxidant activity of faba bean seeds harvested from plants grown from seeds pre-treated with selenium, garlic husk extract and/or lemon peel extract and to evaluate their in vivo effects in a rat model of UC. 54 female rats were divided randomly into nine groups (n = 9). All groups were given the different tested treatments 14 days prior to UC induction using acetic acid (intra-rectal injection of 2 ml, 4% v/v in saline). Our results revealed that the treatment of faba bean seeds with a mixture of selenium, garlic husk extract and lemon peel extract before planting led to a significant increase in selenium, nitrogen, potassium, total protein, phenolic and flavonoid content in the harvested faba bean seeds with a subsequent enhancement of their antioxidant capacity. Consumption of such faba beans showed potential protective and therapeutic effects during experimental colitis by reducing colonic oxidative stress and increasing colonic antioxidant defense mechanisms. Further research is required to understand the mechanisms by which faba beans influence colitis, their effects on various inflammatory biomarkers and their impact on the severity of colitis in humans.

lipids, proteins, and DNA against reactive oxygen and nitrogen species (ROS and RNS).⁶

Selenium (Se) is an essential micronutrient involved in many biological processes. An adequate daily intake of Se is important for the functional balance of many organs such as the thyroid, brain, muscles, prostate and testes.⁷ Se acts as an antioxidant protecting cells and tissues from oxidative stress, and therefore sustaining redox status in cells. The antioxidant activity of Se is due to its incorporation into the structure of selenoproteins such as thioredoxin reductases (TrxR), glutathione peroxidases, selenoprotein P (SelP), selenoprotein F (SelF), selenoprotein S (SelS) and selenoprotein M (SelM).⁸⁻¹⁰

Se organic compounds are much more bioavailable than Se inorganic compounds. Accordingly, it should be pointed out that obtaining the daily requirement of selenium from foods with a high content of Se is much better than obtaining Se from supplements.^{7,11}

Garlic (*Allium sativum* L.) is a common spice with many health benefits that are attributed to its various bioactive compounds, such as organic sulfides, saponins, phenolic compounds, and polysaccharides.^{12,13} Garlic and its active ingredients, mainly phenols and saponins, have a certain antioxidant activity.¹⁴ Moreover, the extract of garlic skins (peels or husks)



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has shown strong antioxidant activity due to its constituents such as phenyl propanoids. $^{15}\,$

Lemon (*Citrus limon* L.) is an important medicinal plant that belongs to the Rutaceae family. Lemon peels exhibit a wide range of biological activities due to the abundance of bioactive constituents. Lemon peels contain phenolic compounds (phenolic acids and flavanones) that are responsible for various bioactivities such as antimicrobial, antioxidant and anticancer.¹⁶

Ulcerative colitis (UC) is one of the inflammatory bowel diseases characterized by inflammation and ulceration of the lining of the colon and rectum. The exact cause of UC remains unknown. Previously, diet and stress were thought to be the cause, but now it is known that these factors may exaggerate but do not cause UC. One possible cause is an uncontrolled immune system response against intestinal microbial flora. This abnormal immune response causes the immune system to attack the cells in the digestive tract as well.^{17,18}

Oxidative stress plays a critical role in the pathogenesis of UC and its malignant progression to colorectal cancer (CRC).¹⁹ Oxidative stress occurs if the generation of ROS exceeds the defensive capability of the antioxidant system in cells. This excessive ROS production causes lipid peroxidation, intestinal mucosal barrier damage, bacterial translocation, and an inflammatory response.²⁰ Accordingly, increasing the buffering capability of the antioxidant defense in the host using a diet with enhanced antioxidant activity may neutralize ROS overproduction and ameliorate UC.

In a previous study,²¹ we successfully grew bean plants from grains pre-treated with selenium, garlic husk extract and lemon peel extract using a recent three-in-one approach. This new approach takes advantage of the nutritional value of the residue extracts in increasing the yield of the beans and enhancing their antioxidant activity as well as recycling these residues.

In the present study, we aim to evaluate the antioxidant activity of faba bean seeds harvested from plants grown from seeds pre-treated with selenium, garlic husk extract and/or lemon peel extract and to investigate the extent of their prophylactic effects in preventing acetic acid (AA)-induced colitis in experimental rats.

2 Materials and methods

2.1 Animals

The study was performed in accordance with the Guidelines for the Care and Use of Laboratory Animals,^{22,23} the basis of the animal ethics guidelines of the Institutional Animals Ethics Committee (protocol no. 2011/186) and the basis of the Canadian Council on Animal Care guidelines. In the current study, a total of 54 female albino rats aged 6–8 weeks old and weighing approximately 90–120 g were obtained from the animal house, Agriculture Research Center, Cairo, Egypt. The rats were housed in plastic cages, and were provided a standard pellet diet and water *ad libitum*. Animals were housed at normal temperatures under a normal dark/light cycle [temperature (25 \pm 2 °C) and 12 h light/dark cycle] and acclimatized for a period of 7 days to laboratory conditions.

2.2 Experimental design

This experiment included the following nine treatments using 6 experimental rats per each treatment: (1) the negative control group (NC) - in which rats were fed the basal diet; (2) the colitis positive control group (PC) - in which rats were fed the basal diet; (3) the bean group (B) - in which rats were fed the basal diet supplemented with untreated beans; (4) the selenium group (Se) - in which rats were fed the basal diet supplemented with beans pre-treated with selenium before planting; (5) the garlic group (G) – in which rats were fed the basal diet supplemented with beans pre-treated with garlic husk extract before planting; (6) the lemon group (L) - in which rats were fed the basal diet supplemented with beans pre-treated with lemon peel extract before planting; (7) the garlic + selenium group (G + Se) - in which rats were fed the basal diet supplemented with beans pre-treated with garlic husk extract and selenium before planting; (8) the lemon + selenium group (L + Se) - in which rats were fed the basal diet supplemented with beans pre-treated with lemon peel extract and selenium before planting; and (9) the garlic + lemon + selenium group (G + L + Se) - in which rats were fed the basal diet supplemented with beans pre-treated with lemon peel, garlic husk extracts and selenium before planting.

2.3 Induction of colitis

All groups were given the mentioned treatments for 14 days. Under light sedation with ether, colitis was induced on day 14 by intra-rectal injection of 2 ml of acetic acid (AA) (4% v/v in saline) using an elastic catheter (with an outer diameter of 2 mm).^{24,25} Negative control animals underwent the same procedure using an equal volume of normal saline instead of AA solution.

2.4 Sample preparation

The rats were subjected to the previously mentioned treatments for a period of 60 days. At the end of the experiment, blood was withdrawn from the retro-orbital plexus and the serum was separated and stored at -20 °C for biochemical analysis. Animals were sacrificed, and the colons were removed, washed with normal saline and observed for macroscopic colitis assessment, and then cut into small portions and kept at -80 °C for biochemical analysis and histopathological investigations.

2.5 Evaluation of the disease activity index (DAI)

The clinical severity of colitis was evaluated through the calculation of DAI according to a previously described method.²⁶ The DAI uses a scoring system for evaluating the percentage of weight loss, stool consistency, and rectal bleeding. The following parameters were recorded daily: body weight loss (0, none; 1, 1-5%; 2, 6-10%; 3, 11-20%; and 4, >20%), diarrhea (0, normal; 1, soft stool but still formed; 2, very soft stool; 3, mild

diarrhea; and 4, severe diarrhea), and rectal bleeding (0, normal; 1, positive hemoccult; 2, visible blood traces in stool; 3, mild bleeding; and 4, severe bleeding). The DAI values were calculated according to this equation:

DAI=

 $\frac{\text{body weight loss score} + \text{diarrhea score} + \text{rectal bleeding score}}{3}$

2.6 Macroscopic scoring

For about half of the rats, the entire excised colon was weighed and used for macroscopic scoring. The assessment of macroscopic injury was carried out by analyzing the presence or absence of colonic thickening, hyperemia, ulcers, necrosis, and adherence to nearby organs, according to previously described criteria.²⁷

2.7 Histopathological investigations

Histopathological assessments were performed according to the method described by Bancroft and Gamble (2008).²⁸ Colon biopsies were fixed with 10% neutral buffer formalin and embedded in paraffin. The specimens were washed, dehydrated using alcohol, cleared in xylene and embedded in paraffin wax blocks. For histopathological assessments, 3 μ m thickness sections were cut and stained with hematoxylin and eosin (H&E) and were mounted and observed microscopically for histopathological changes by an experienced pathologist in a blinded fashion.

2.8 Assessment of oxidative stress and the antioxidant indices in colon tissues

2.8.1 Colonic malondialdehyde (MDA). The colonic malondialdehyde (MDA) content was determined based on the method described by Ohkawa *et al.* (1979).²⁹ 2 ml of colon homogenate (20% w/v) supernatant was mixed with an equal amount of thiobarbituric acid TCA (10% w/v), frozen for 15 min, and then centrifuged at 20 000 rpm. Afterwards, 2 ml of TCA was mixed again with 2 ml of the supernatant. The solution was heated for 10 min at 100 °C and rapidly cooled at 0 °C for 5 min. The concentration was measured by spectrophotometry at 535 nm.

2.8.2 Colonic superoxide dismutase (SOD) activity. SOD activity in the colonic tissue homogenate was measured according the method described by Nishikimi *et al.* (1972).³⁰ SOD assay relies on the ability of SOD to inhibit the phenazine methosulphate-mediated reduction of the nitro-blue tetrazolium dye.

2.8.3 Colonic catalase (CAT) activity. CAT activity in the colonic tissue homogenate was assessed based on the method described by Aebi (1984).³¹ CAT assay is a colorimetric method that depends on the measurement of the hydrogen peroxide (H_2O_2) substrate using a redox dye. The change in the color intensity at 570 nm is directly proportional to the CAT activity.

2.8.4 Colonic glutathione reductase (GR) activity. GR activity in the colonic tissue homogenate was assayed colorimetrically as described by Goldberg and Spooner (1983).³² The

method is based on the capacity of GR to catalyze the reduction of GSSG to GSH in the presence of NADPH, which is oxidized to NADPH⁺. The decrease in absorbance at 340 nm was measured.

2.8.5 Colon reduced glutathione (GSH) level. The GSH level in the colon tissue homogenate was measured based on the method described by Beutler *et al.* (1963).³³ The method produces a yellow color by reducing 5,5'-dithiobis(2-nitrobenzoic acid) (DTNB) with glutathione (GSH). The reduced chromogen was directly proportional to the GSH concentration and its absorbance could be measured at 405 nm.

2.9 Determination of the total phenolic content (TPC) of the dry faba bean seed extract

The TPC was determined according to the Folin–Ciocâlteu colorimetric method referred to Gargouri *et al.* (2019).³⁴ Briefly, 1 mL of the ethanolic extract was mixed with 0.5 mL of the Folin–Ciocâlteu reagent in a test tube and thoroughly shaken. After 3 min, 1 mL of saturated Na₂CO₃ (20%) was added to the mixture and then the volume was made up to 10 mL with distilled water. The reaction was allowed to proceed for 1 h. A blank was prepared with 1 mL of distilled water instead of the sample. After 1 h, the absorbance was recorded at 725 nm using a spectrophotometer (UV-vis spectrophotometer UV 9100 B, LabTech). The concentration of the total soluble phenols was calculated using the standard curve of gallic acid. The total phenol concentration was expressed as μ g equivalents of gallic acid per g DW of the sample.

2.10 Determination of the total nitrogen, total potassium, total selenium and total soluble protein content of the dry faba bean seed extract

Faba bean seeds were oven-dried at 65 °C, and then wet digested using a mixture of H_2SO_4 and H_2O_2 according to the method outlined by Cottenie *et al.* (1982).³⁵ The total nitrogen content in the dried faba bean seeds was determined by the micro-Kjeldahl method using 5% boric acid and 40% NaOH as described by Black (1965).³⁶ The total potassium content was determined using a flame photometer.³⁷ The total selenium content was determined using ICP mass spectrometry.³⁸ The total soluble protein concentration was quantified by the method of Bradford³⁹ using bovine serum albumin (BSA) as a standard.

2.11 Determination of the total flavonoid content (TFC) of the dry faba bean seed extract

The TFC was determined by the aluminum chloride colorimetric assay as described previously.⁴⁰ An aliquot of 1 mL of the ethanolic extract was added to 4 mL of distilled water in a 10 mL volumetric flask. Then, 0.3 mL of 5% NaNO₂ was added. After 5 min, 0.3 mL of 10% AlCl₃ was added. At the 6th min, 2 mL of 1 M NaOH was added and the total volume was made up to 10 mL with distilled water. The solution was mixed well and the absorbance was measured against the blank at 510 nm. The concentration of total flavonoids was calculated using the standard curve of quercetin and was expressed as μg of quercetin equivalent per g DW of the sample.

2.12 Determination of the antioxidant capacity of the dry faba bean seed extract

The antioxidant activity of the dry faba bean seed extract was determined based on 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical-scavenging activity as described by Alexandra *et al.* (2019).⁴¹ In the DPPH assay, 1 mL of the sample was mixed with 0.5 mL of 50 μ M DPPH in ethanol and kept in the dark for 30 min. The absorbance of the mixture was measured at 517 nm. A vitamin C standard (5–30 μ g mL⁻¹) was used as the positive control. The radical scavenging activity was determined based on the percentage inhibition of absorbance, which was calculated using the following formula:

% inhibition = $[(A_{\text{control}} - A_{\text{sample}})/A_{\text{control}}] \times 100$,

where A_{control} is the absorbance of the reaction without the sample and A_{sample} is the absorbance of the sample.

Lower absorbance indicates higher DPPH radical scavenging activity. The IC_{50} value is defined as the concentration of the sample required to scavenge 50% of DPPH. Therefore, lower IC_{50} indicates higher antioxidant activity.

2.13 Statistical analysis

The SPSS statistical package 21.0 (SPSS Inc., Chicago, Illinois, USA) was used for all statistical analyses. All data were presented as means \pm standard deviation. One-way ANOVA was used to analyze differences between means for normally distributed values, followed by *post hoc* analysis. Correlations were evaluated using the Pearson correlation coefficient. All *p*-values were two-sided and the probability value, *p* < 0.05, was considered statistically significant.

3 Results

3.1 Effect of feeding with different types of beans on AAinduced alterations in disease severity and colonic macroscopic damage

The clinical severity of colitis was evaluated for each experimental group. The DAI uses a scoring system for evaluating the percentage of weight loss, stool consistency, and rectal bleeding. The DAI scores of the positive control (PC) rats were significantly higher than those of the negative control (NC) group (p < 0.001; Fig. 1A). However, compared with the PC group, rats fed with the different types of beans showed significant reductions in the DAI scores, especially those of the Se and G + L + Se groups (p < 0.001; Fig. 1A). Furthermore, macroscopic injury was evaluated with scores ranging from 0 to 10 points. The NC group scored below 1, while the PC group showed major damage such as increased colonic thickening, necrosis, ulcers, and adherence to adjacent organs. Feeding with different types of beans significantly decreased the intensity of all of the macroscopic damage especially in the Se, L + Se and G + L + Se groups (p < 0.001; Fig. 1A and C).

3.2 Effect of feeding with different types of beans on AAinduced histopathological changes in rat colon tissues

Fig. 2 shows the results of hematoxylin and eosin staining. The analysis of the Swiss-roll sections of the NC group revealed the normal histology of the colon wall; it consisted of tunica mucosa (mucosal crypts) that was formed of simple columnar epithelium and lamina propria that contained glands with basally situated nuclei and numerous goblet cells. Tunica mucosa rested on muscularis mucosa, and then it was submucosa and finally the muscular wall of the colon. The PC group showed severe damage with many ulcerated areas of the mucosa with minimal remnants of the crypts, which indicated hemorrhage and inflammatory cell infiltration. Also, the submucosa showed large areas of hemorrhage and inflammatory cell infiltration. The B group showed ulceration and destruction of the colon mucosa with necrosis in the glands and extensive inflammatory cell infiltration. The best protective action of a single agent was observed in the Se group. Few sections showed mild mononuclear inflammatory cell infiltration in the colonic mucosa, and some sections were apparently normal. The colon sections from the G group showed the least improvement. Some sections exhibited exaggerated mucus secretion with cystically dilated glands. The submucosa showed inflammatory edema. Some other sections showed extensive glandular necrosis with heavy inflammatory cell infiltration mainly by neutrophils. Both mucosa and submucosa were infiltrated by dense inflammatory cells. The L group showed mildly affected mucosa that appeared infiltrated by a few inflammatory cells, while the submucosa showed extensive inflammatory reaction. An extensive inflammatory reaction was clearly observed in the colonic wall of animals from this group. Some severely affected sections exhibited marked mucosal damage and ulceration. Some sections showed cystically dilated crypts. Regarding the groups treated using more than one protective agent, they exhibited a synergistic action in alleviating the induced colon damage. The G + Se group showed an apparently normal colon wall in most examined sections. Only mild inflammatory cell infiltration was observed around the glands. Concerning the colon sections from the L + Se group, all examined sections appeared apparently normal, except one case exhibiting mucosal damage and inflammatory cell infiltration of both mucosa and submucosa. Cystically dilated glands with exaggerated mucus production was also a quite common finding. The best protective action was achieved in the G + L + Se group. Apparently normal mucosa was observed in almost all examined sections. Mild mucosal and submucosal inflammatory reactions were also detected. A few sections showed cystically dilated glands with increased mucus secretion.

3.3 Effect of feeding with different types of beans on the colonic oxidative and antioxidant profiles in acetic acid (AA)-induced ulcerative colitis

We evaluated the effect of feeding with different types of beans on oxidative stress and antioxidant defense in colon tissues.



Fig. 1 Effect of feeding with different types of beans on acetic acid (AA)-induced alterations in disease severity and colonic macroscopic damage. (A) Disease activity index (DAI) scores of rat colons in different groups. (B) Quantification of the colon damage macroscopic scoring in different groups. (C) Photographic evaluation of the colon. Negative control (NC) group: colon without morphological alterations; positive control (PC) group: colon shows severe shortening and thickening with severe hyperemia and extensive tissue necrosis; bean (B) group: colon shows moderate shortening and thickening and thickening and thickening and thickening with mild hyperemia; garlic (G) group: colon shows moderate shortening and thickening with mild hyperaemia; garlic + selenium (G + Se) group: colon shows moderate shortening and thickening with mild hyperaemia; garlic + selenium (G + L + Se) group: colon shows only mild hyperaemia. Each value represents the mean \pm SD (n = 6). Significant vs. the Se group. b: Significant vs. the PC group. c: Significant vs. the G group. d: Significant vs. the L group. g: Significant vs. the G + Se group. h: Significant vs. the L + Se group.

The results, presented in Table 1 and Fig. 3, show that in the positive control (PC) group, colonic MDA was increased by 2.7-fold compared to the negative control (NC) group (p < 0.001), indicative of increased oxidative stress in AA-induced colitis. On the other hand, rats fed different types of beans, especially those in the selenium (Se) and G + L + Se groups showed a significant decrease in the levels of colonic MDA by 31.4% and 29.6%, respectively, when compared to the PC group (p < 0.001).

In the PC group, the GRD and GSH activity levels in the colon tissues were lower than those in the NC group by 41.5% and 59.3%, respectively (p < 0.001), while the activities of SOD and CAT were increased by 1.37- and 1.6-fold, respectively (p < 0.001). Feeding with different types of beans had

protective effects against these changes. Rats fed beans in the Se and G + L + Se groups showed increased SOD activity by 1.16- and 1.12-fold, respectively, with significant differences compared to the PC group. With regard to the colonic GSH level and GRD activity, rats fed beans in the G + L + Se, Se, L + Se, and G + Se groups showed significantly increased GSH levels by 3.23-, 2.18-, 1.8-, and 1.4-fold, respectively, compared to the PC group (p < 0.001). Moreover, colonic GRD activity significantly increased by 2.27-, 2.37-, 1.63- and 1.45-fold, respectively, compared to the PC group (p < 0.001). Regarding colonic CAT activity, rats fed beans in the Se, G and G + L + Se groups showed significantly increased CAT activity by 2-, 1.16- and 1.1-fold, respectively, compared to the PC group (p < 0.001).



Fig. 2 Effect of feeding with different types of beans on acetic acid (AA)-induced histopathological changes in rat colonic tissues. Photomicrographs of the Swiss-roll sections of the colons of the negative control (NC) group: showing a normal structure of the colon wall. Inset shows a higher magnification of the crypts, showing many goblet cells and the absorptive simple columnar cells (arrow). The positive control (PC) group: showing distorted mucosa with remnants of crypts (black arrow) and mononuclear inflammatory cell infiltration of the submucosa (arrow). The inset shows a higher magnification of the submucosa with mononuclear inflammatory cells. The bean (B) group: showing complete destruction and necrosis of the colonic mucosa with submucosal edema and mononuclear inflammatory cell infiltration. Inset shows a higher magnification of the submucosal edema and mononuclear inflammatory cell infiltration. Inset shows a higher magnification of the gland with an extensive inflammatory reaction to the serosa. The garlic (G) group: showing exaggerated mucus secretion with cystic dilatation of the gland with an extensive inflammatory cell infiltration. The lemon (L) group: showing inflammatory cell infiltration in the mucosa with cystic dilatation. The garlic + selenium (G + Se) group: showing an apparently normal colon wall. The inset shows a higher magnification, showing mild inflammatory cell infiltration in the colonic mucosa. The inset shows a higher magnification, showing mild inflammatory cell infiltration in the colonic mucosa. The inset shows a higher magnification, showing many gole cell infiltration, showing mild inflammatory cell infiltration in the colonic mucosa. The inset shows a higher magnification, showing many gole cells infiltration in the colonic mucosa. The inset shows a higher magnification, showing many gole cells infiltration in the colonic mucosa. The inset shows a higher magnification, showing many gole cells infiltration in the colonic mucosa. The inset shows a higher magnification,

3.4 Effect of different treatments on the total phenolic content (TPC), total flavonoid content (TFC), total nitrogen (N) content, total potassium (K) content, total selenium content (Se), total soluble protein content and the antioxidant capacity of the dry faba bean seed extract

The levels of total phenolic and total flavonoid content in the ethanolic extracts of dry faba bean seeds are shown in Fig. 4. Faba bean seeds from the G + L + Se and L + Se treatments showed the highest level of phenolic compounds compared to

the control bean group (B) and the other treatments with high significant difference (p < 0.001) (Fig. 4A). On the other hand, G + L + Se, L + Se and Se treatments significantly increased the total flavonoid content of faba bean seeds compared to the control bean group (B) and the other treatments with high significant difference (p < 0.001) (Fig. 4B).

All treatments increased the total N and K contents of dry faba bean seeds compared to the control bean group (B) with high significant difference (p < 0.001) (Fig. 4C). Also, the Se content of dry faba bean seeds was increased in all treatments,

$ \begin{array}{llllllllllllllllllllllllllllllllllll$		NC	PC	В	Se	C	L	G + Se	L + Se	G + L + Se	Statistics
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	-MDA (nmol	8.922 ±	$24.07^{(a)} \pm$	$18.91^{(a,b)} \pm$	$16.5^{(a,b,c)} \pm$	$19.2^{(a,b,d)} \pm$	$21.1^{(a,b,c,d,e)} \pm$	$19.64^{(a,b,d,f)} \pm$	$19.31^{\rm (a,b,d,f)}\pm$	$16.94^{(a,b,c,e,f,g,h)} \pm$	F: 13.21, P:
$ \begin{array}{ccccc} \text{SOD} \left(\text{U}\text{mg}^{-1} & 1.964 \pm & 2.70^{(a)} \pm & 1.961^{(b)} \pm & 3.142^{(a,b,c,i)} \pm & 2.40^{(a,b,c,i)} \pm & 2.578^{(a,b,c,d,c)} \pm & 2.094^{(a,b,c,d,c,1)} \pm & 2.738^{(a,c,d,c,1)} \pm & 2.788^{(a,b,c,d,c,1)} \pm & 2.788^{(a,b,c,1)} \pm & 2.788^{(a,b,$	s^{-1} tissue)	0.434	1.80	0.319_{1}	0.987	1.616	4.43	2.38	1.619	0.377	0.000
$ \begin{array}{ccccc} \text{protein} & 0.023 & 0.010 & 0.049 & 0.156 & 0.049 & 0.0179 & 0.0171 & 0.0167 & 0.0036^{(a_1,b_1c_1,b_1,c_2,d_2)} \pm 0.0034^{(a_1,b_1,c_1,c_1,b_1)} \pm 0.0036^{(a_1,d_2,c_1,b_1)} \pm 0.00356^{(a_1,d_2,c_1,b_1)} \pm 0.00356^{(a_1,d_2,c_1,b_1)} \pm 0.00356^{(a_1,d_2,c_1,b_1)} \pm 0.00754^{(a_1,b_2,c_1,c_1,b_2,c_1,c_1,b_1)} \pm 0.00756^{(a_1,b_2,c_1,c_1,b_2,c_1,c_1,b_2,c_1,c_1,b_2,c_1,c_1,b_2,c_1,c_1,b_2,c_1,c_1,b_2,c_1,c_1,b_2,c_1,c_1,c_1,c_1,c_1,c_1,c_1,c_1,c_1,c_1$	SOD (U mg ⁻¹	$1.964 \pm$	$2.707^{(a)} \pm$	$1.961^{(b)} \pm$	$3.142^{(a,b,c)} \pm$	$2.40^{(a,b,c,d)} \pm$	$2.578^{(a,b,c,d,e)} \pm$	$2.094^{(a,b,c,d,e,f)} \pm$	$2.738^{(a,c,d,e,f,g)}\pm$	$3.025^{(a,b,c,d,e,f,g,h)} \pm$	F: 124.4, P:
$ \begin{array}{ccccc} 2 \mathrm{AT} \left(\mathrm{U}\mathrm{mg}^{-1} & 0.0023 \pm & 0.0037^{(a)} \pm & 0.0036^{(a)} \pm & 0.0074^{(a,b,c)} \pm & 0.0043^{(a,b,c,d,c)} \pm & 0.0034^{(a,b,c,d,c)} \pm & 0.0036^{(a,d,c,f)} \pm 3.8 & (3.10^{-5} & 4.5 \times 10^{-5} & 1.4 \times 10^{-4} & 3.7 \times 10^{-4} & 9.1 \times 10^{-5} & 5.7 \times 10^{-5} & 5 \times 10^{-5} & \times 10^{-5} & 1.4 \times 10^{-5} & 3.7 \times 10^{-4} & 9.1 \times 10^{-5} & 5.7 \times 10^{-5} & 5 \times 10^{-5} & \times 10^{-5} & 1.4 \times 10^{-5} & 1.4 \times 10^{-4} & 0.1406^{(a,b,d)} \pm & 0.406^{(a,b,d)} \pm & 0.406^{(a,b,d)} \pm & 0.449^{(a,b,c,d,c,f)} \pm & 0.574^{(a,b,c,d,c,f)} \pm & 0.5056^{(a,b,d)} \pm & 0.5056^{(a,b,d)} \pm & 0.574^{(a,b,c,d,c,f)} \pm & 0.574^{(a,b,c,d,c,f)} \pm & 0.574^{(a,b,c,d,c,f)} \pm & 0.574^{(a,b,c,d,c,f)} \pm & 0.5056^{(a,b,d)} \pm & 0.5056^$	orotein)	0.023	0.010	0.049	0.156	0.049	0.0179	0.0171	0.0167	0.1052	0.000
$ \begin{array}{c cccc} \text{ orderin)} & 8 \times 10^{-5} & 4.5 \times 10^{-5} & 1.4 \times 10^{-4} & 3.7 \times 10^{-5} & 5.1 \times 10^{-5} & 5.7 \times 10^{-5} & 5.7 \times 10^{-5} & 8 \times 10^{-5} & 1355555555555555555555555555555555555$	CAT (U mg ⁻¹	$0.0023 \pm$	$0.0037^{(a)} \pm$	$0.0036^{(a)} \pm$	$0.0074^{(a,b,c)} \pm$	$0.0043^{(a,b,c,d)} \pm$	$0.004^{(a,b,c,d,e)} \pm$	$0.0034^{(a,b,d,e,f)} \pm$	$0.0036^{(a,d,e,f)} \pm 3.8$	$0.0041^{(a,b,c,d,g,h)} \pm$	F: 248.5, P:
$ \begin{array}{ccccccc} 3SH(\text{mmol} & 0.784 \pm & 0.319^{(a)} \pm & 0.376^{(a,b,b)} \pm & 0.696^{(a,b,c)} \pm & 0.406^{(a,b,d)} \pm & 0.406^{(a,b,d)} \pm & 0.406^{(a,b,d)} \pm & 0.406^{(a,b,d)} \pm & 0.0075 \pm & 0.0075 \\ \text{ng}^{-1} & 0.0081 & 0.001 & 0.0075 & 0.0354 & 0.011 & 0.0034 & 0.0056 & 0.0075 & 0 \\ \text{notein} & & & & & & & & & & & & & & & & & & &$	protein)	$8 imes 10^{-5}$	$4.5 imes 10^{-5}$	$1.4 imes 10^{-4}$	$3.7 imes 10^{-4}$	$9.1 imes 10^{-5}$	$5.7 imes 10^{-5}$	$5 imes 10^{-5}$	$ imes 10^{-5}$	$1.3 imes 10^{-4}$	0.000
mg^{-1} 0.0081 0.001 0.005 0.0354 0.011 0.0034 0.0056 0.0075 (protein) $2.2.2$ (and $2.2.2$ (abed a constant) $2.2.2$ (abed constant) $2.2.2$ (bred of constant) $2.2.2$ (bred constant) $2.2.2$ (bred constant) $2.2.2$ (bred constant) $2.2.2$ (bred constant) $2.2.2$ (constant)	BSH (mmol	$0.784 \pm$	$0.319^{(a)} \pm$	$0.376^{(a,b)} \pm$	$0.696^{(a,b,c)} \pm$	$0.406^{(a,b,d)} \pm$	$0.406^{(a,b,d)} \pm$	$0.449^{(a,b,c,d,e,f)} \pm$	$0.574^{(a,b,c,d,e,f,g)} \pm$	$1.033^{(a,b,c,d,e,f,g,h)} \pm$	F: 504.5, P:
brotein) 	ng ⁻¹	0.0081	0.001	0.0095	0.0354	0.011	0.0034	0.0056	0.0075	0.0365	0.000
and first a start a star	orotein)										
$\frac{1}{3} \frac{1}{3} \frac{1}$	JRD (U	$2.767 \pm$	$1.619^{(a)} \pm$	$2.096^{(a,b)} \pm$	$3.843^{(a,b,c)} \pm$	$2.185^{(a,b,d)} \pm$	$1.895^{(a,b,c,d,e)} \pm$	$2.351^{(a,b,c,d,e,f)} \pm$	$2.641^{(b,c,d,e,f,g)} \pm$	$3.689^{(a,b,c,d,e,f,g,h)} \pm$	F: 317.7, P:
ng ⁻¹ 0.0283 0.0029 0.0496 0.1779 0.0287 0.0079 0.0261 0.0082 (morein)	ng ⁻¹ protein)	0.0283	0.0029	0.0496	0.1779	0.0287	0.0079	0.0261	0.0082	0.1154	0.000

group. d: Significant versus the selenium (Se) group. e: Significant versus the garlic (G) group. f: Significant versus the lemon (L) group. g: Significant versus the garlic + selenium (G + Se group. As selenium (L + Se) group. SOD: super oxide dismutase, GSH: reduced glutathione, GRD: glutathione reductase, CAT: catalase, MDA: malondialdehyde.

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especially in the G + L + Se treatment compared to the control bean group (B) with high significant difference (p < 0.001)(Fig. 4D). The G + L + Se, L + Se and Se treatments significantly increased the total protein content of dry faba bean seeds compared to the control bean group (B) and the other treatments with high significant difference (p < 0.001) (Fig. 4E).

To determine the antioxidant capacity, a DPPH radical scavenging capacity assay was employed to test the antioxidant capacity of the phenolic and flavonoid contents of the dry faba bean seed extract and the results are shown in Fig. 4F. The IC50% value (defined as the concentration of the sample required to scavenge 50% of DPPH) was chosen to indicate the antioxidant activity. Therefore, a lower IC50% indicates higher antioxidant activity. The data obtained indicated that the antioxidant capacity of seeds from the G + L + Se, L + Se and Se treatments was significantly higher than that of seeds from the control bean group (B) and the other treatments with a high significant difference (p < 0.001).

The correlations between the antioxidant capacity ($IC_{50\%}$) value), TPC, TFC, N content, K content, Se content and protein content were established, and the correlation coefficients (r) are tabulated in Table 2. A significant positive correlation was found between TPC and TFC (r = 0.923, P < 0.01), TPC and N content (r = 0.555, P < 0.01), TPC and protein content (r = 0.888, P < 0.01), TVC and K content (r = 0.629, P < 0.01), TVC and protein content (r = 0.937, P < 0.01), N content and K content (r = 0.659, P < 0.01)0.01), N content and Se content (r = 0.779, P < 0.01), N content and protein content (r = 0.627, P < 0.01), K content and Se content (r = 0.755, P < 0.01), K content and protein content (r = 0.665, P < 0.01) and Se content and protein content (r = 0.579, P < 0.01)P < 0.01). Significant negative correlations were observed between the antioxidant capacity (IC50% value) and TPC, TVC, nitrogen content, potassium content and protein content (r = -0.836, -0.896, -0.588, -0.651 and -0.827, respectively, *P* < 0.01).

3.5 Relationship between the TPC, TFC, N content, K content, total Se content, total soluble protein content and the antioxidant capacity (IC50%) of the dry faba bean seed extract and oxidative stress and the antioxidant indices in colon tissues

The results shown in Table 3 indicate that no significant correlations were found between TPC, TFC, N content, K content, total Se content, total soluble protein content and the antioxidant capacity (IC50%) of the dry faba bean seed extract and both colonic MDA level and CAT activity (p > 0.05). Conversely, significant positive correlations were found between the TPC, TFC, K content and protein content of the dry faba bean seed extract and colonic SOD activity, GSH level and GRD activity (p < 0.001). Furthermore, significant negative correlations were observed between the antioxidant capacity (IC50% value) and colonic SOD activity, GSH level and GRD activity (p < 0.001).

4 Discussion

In a previous study,²¹ we studied the effect of soaking bean seeds before planting in garlic peel extract and lemon peel

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Fig. 3 Effect of feeding with different types of beans on the colonic oxidative and antioxidant profiles in acetic acid (AA)-induced ulcerative colitis. (A) Effect on the colonic malondialdehyde level, (B) effect on colonic superoxide dismutase activity, (C) effect on the colon reduced glutathione (GSH) level, (D) effect on colonic glutathione reductase activity and (E) effect on colonic catalase activity. Data are represented as mean \pm SD, P < 0.001, n = 6. a: Significant *versus* the negative control (NC) group. b: Significant *versus* the positive control (PC) group. c: Significant *versus* the bean (B) group. d: Significant *versus* the selenium (Se) group. e: Significant *versus* the garlic (G) group. f: Significant *versus* the lemon (L) group. g: Significant *versus* the garlic + selenium (G + Se) group. h: Significant *versus* the lemon + selenium (L + Se) group.

extract with or without selenium on increasing fresh and dry weights, plant height, and nitrogen and potassium content, in addition to the effect on TFC and TVC during different growth stages, which was reflected in the final productivity. In the current study, we have evaluated the antioxidant activity of faba bean seeds harvested from plants grown from seeds after being subjected to the previously mentioned soaking treatments and investigated the extent of their prophy-



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Fig. 4 Effect of different treatments on (A) the total phenolic content (TPC), (B) total flavonoid content (TFC), (C) total nitrogen and potassium contents, (D) total selenium content, (E) total protein content and (F) the antioxidant capacity ($IC_{50\%}$) of the dry faba bean seed extract. Data are represented as mean \pm SD, P < 0.001. a: Significant *versus* the untreated beans (B). b: Significant *versus* the selenium pre-treated beans (Se). c: Significant *versus* the garlic husk extract pre-treated beans (G). d: Significant *versus* the lemon peel extract pre-treated beans (L). e: Significant *versus* the lemon peel extract + selenium pre-treated beans (L + Se).

lactic effects in preventing acetic acid (AA)-induced colitis in experimental rats. The AA-induced colitis model usually mimics the pathogenesis of human UC.^{42,43}

Herein, clinical manifestations of colitis were evaluated for the experimental groups. The DAI was used to assess the percentage weight loss, stool consistency, and rectal bleeding. The obtained results showed that the rats fed with the different types of beans showed significant reductions in the DAI scores, especially in the Se and G + L + Se groups. Regarding macroscopic injury, feeding with different types of beans significantly reduced the severity of all macroscopic damage, especially in the Se, L + Se and G + L + Se groups.

An AA-induced colitis model can represent several histopathological features that are similar to human ulcerative

Table 2 Correlation analysis of the total phenolic content (TPC), total flavonoid content (TVC), nitrogen (N) content, potassium (K) content, selenium (Se) content, protein content and antioxidant capacity (IC_{50%})

	TPC	TVC	N content	K content	Se content	Protein content	IC _{50%}
TPC	1.000 ()	0.923** (0.000)	0.555** (0.009)	0.530* (0.013)	$0.440^{*}(0.046)$	0.888** (0.000)	-0.836** (0.000)
TVC	0.923** (0.000)	1.000(-)	0.492* (0.023)	$0.629^{**}(0.002)$	0.415(0.061)	0.93/**(0.000)	-0.896** (0.000)
N content	0.555** (0.009)	0.492* (0.023)	1.000(-)	0.659** (0.001)	$0.7/9^{**}(0.000)$	0.62/**(0.002)	-0.588** (0.005)
K content	0.530* (0.013)	0.629** (0.002)	0.659** (0.001)	1.000 (—)	0.755** (0.000)	0.665** (0.001)	-0.651** (0.001)
Se content	0.440* (0.046)	0.415 (0.061)	0.779** (0.000)	0.755** (0.000)	1.000 (—)	0.579** (0.005)	-0.499* (0.041)
Protein content	$0.888^{**}(0.000)$	0.937** (0.000)	0.627** (0.002)	$0.665^{**}(0.001)$	0.579** (0.005)	1.000 (—)	$-0.827^{**}(0.000)$
$IC_{50\%}$	-0.836** (0.000)	0.896** (0.000)	-0.588** (0.005)	-0.651** (0.001)	-0.499* (0.041)	-0.827** (0.000)	1.000(-)

**Correlation is significant at the 0.01 level (2-tailed).

Table 3 Correlation analysis between the TPC, TFC, N content, K content, Se content, protein content and the antioxidant capacity (IC_{50%}) of the dry faba bean seed extract and the oxidative stress and antioxidant profiles in colon tissues

MDA SOD CAT GSH GRD TPC -0.326 (0.149) 0.608** (0.003) 0.007 (0.976) 0.778** (0.000) 0.643** (0.002) TVC 0.385 (0.85) 0.807** (0.000) 0.271 (0.234) 0.839** (0.000) 0.824** (0.000) N content -0.51 (0.825) 0.313 (0.167) -0.271 (0.235) 0.417 (0.060) 0.168 (0.467) K content -0.164 (0.478) 0.611** (0.003) 0.161 (0.486) 0.551** (0.010) 0.454* (0.039) Se content 0.159 (0.491) 0.305 (0.179) -0.265 (0.247) 0.287 (0.207) 0.039 (0.867) Protein content -0.162 (0.484) 0.679** (0.001) 0.011 (0.964) 0.805** (0.000) 0.616** (0.003) IC _{50%} 0.333 (0.140) -0.713** (0.000) -0.175 (0.448) -0.768** (0.000) -0.698** (0.000)						
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		MDA	SOD	CAT	GSH	GRD
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	TPC	-0.326 (0.149)	0.608** (0.003)	0.007 (0.976)	0.778** (0.000)	0.643** (0.002)
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	TVC	0.385 (0.085)	0.807** (0.000)	0.271(0.234)	0.839** (0.000)	0.824** (0.000)
K content $-0.164(0.478)$ $0.611^{**}(0.003)$ $0.161(0.486)$ $0.551^{**}(0.010)$ $0.454^{*}(0.039)$ Se content $0.159(0.491)$ $0.305(0.179)$ $-0.265(0.247)$ $0.287(0.207)$ $0.039(0.867)$ Protein content $-0.162(0.484)$ $0.679^{**}(0.001)$ $0.011(0.964)$ $0.805^{**}(0.000)$ $0.616^{**}(0.003)$ IC _{50%} $0.333(0.140)$ $-0.713^{**}(0.000)$ $-0.175(0.448)$ $-0.768^{**}(0.000)$ $-0.698^{**}(0.000)$	N content	-0.51(0.825)	0.313 (0.167)	-0.271(0.235)	0.417 (0.060)	0.168 (0.467)
Se content $0.159(0.491)$ $0.305(0.179)$ $-0.265(0.247)$ $0.287(0.207)$ $0.039(0.867)$ Protein content $-0.162(0.484)$ $0.679^{**}(0.001)$ $0.011(0.964)$ $0.805^{**}(0.000)$ $0.616^{**}(0.003)$ IC_{50\%} $0.333(0.140)$ $-0.713^{**}(0.000)$ $-0.175(0.448)$ $-0.768^{**}(0.000)$ $-0.698^{**}(0.000)$	K content	-0.164(0.478)	0.611** (0.003)	0.161 (0.486)	0.551^{**} (0.010)	$0.454^{*}(0.039)$
Protein content $-0.162(0.484)$ $0.679^{**}(0.001)$ $0.011(0.964)$ $0.805^{**}(0.000)$ $0.616^{**}(0.003)$ IC_{50\%} $0.333(0.140)$ $-0.713^{**}(0.000)$ $-0.175(0.448)$ $-0.768^{**}(0.000)$ $-0.698^{**}(0.000)$	Se content	0.159 (0.491)	0.305 (0.179)	-0.265(0.247)	0.287 (0.207)	0.039 (0.867)
$IC_{50\%} 0.333 (0.140) -0.713^{**} (0.000) -0.175 (0.448) -0.768^{**} (0.000) -0.698^{**} (0.000) $	Protein content	-0.162(0.484)	0.679** (0.001)	0.011 (0.964)	0.805** (0.000)	0.616** (0.003)
	IC _{50%}	0.333 (0.140)	-0.713** (0.000)	-0.175(0.448)	-0.768** (0.000)	-0.698** (0.000)

**Correlation is significant at the 0.01 level (2-tailed). TPC: total phenolic content, TVC: total flavonoid content, N content: nitrogen content, K content: potassium content, Se content: selenium content, MDA: malondialdehyde, SOD: super oxide dismutase, CAT: catalase, GSH: reduced glutathione, GRD: glutathione reductase.

colitis, such as mucosal ulceration in which neutrophils infiltrate into the lamina propria and intestinal crypts.⁴² Other histological features include the depletion of goblet cells.⁴³ Our findings showed that feeding with different types of beans significantly improved all the histopathological findings. In this regard, the G + L + Se group exhibited dramatic preventive and curative achievements, which was significantly different from the other groups. Based on these results, we conclude that consuming faba bean seeds pre-treated with a mixture of lemon peel, garlic husk extract and selenium before planting has prophylactic effects in rats with AA-induced colitis.

It has been previously reported that soaking seeds before planting in Se solution increases the rate of selenium uptake and its quantity in the final crop.⁴⁴ Moreover, Se supplementation increases the antioxidant activity of plants.^{45,46} This is clearly evident in this study, where the different soaking treatments led to a significant increase in the Se content of dry faba bean seeds. Also, a significant positive correlation was found between the Se content and the antioxidant capacity of the dry faba bean seed extract.

In this study, no significant correlation was found between the Se content of dry faba bean seeds and the antioxidant indices in the colon tissues (SOD activity, GSH level and GRD activity). However, there was a significant correlation between the Se content and the total protein content, potassium content and antioxidant capacity ($IC_{50\%}$) of dry faba bean seeds, which are all significantly related to the antioxidant indices in the colon tissues. This may confirm the indirect relationship between the Se content of dry faba bean seeds and the antioxidant indices in the colon tissues.

As for garlic, previous studies showed that garlic is a source of antioxidants (phenols and flavonoids) and contains various growth-promoting compounds such as organo-sulphur compounds (allicin and diallyl disulphide) that improve the growth and yield of faba bean plants.^{47,48} Phenolics and flavonoids are two of the major bioactive substances present in citrus fruits such as lemon, with a higher concentration in the peels than in the fruits.⁴⁹ Citrus phenolics and flavonoids are powerful antioxidants and potent free radical scavengers that help in the prevention of diseases that occur due to ROS.⁵⁰ In the current study, soaking faba bean seeds in garlic and/or lemon peel extract mixed with Se solution before planting significantly increased the TPC, TVC, total N content, total K content, total Se content and total protein content and finally the antioxidant capacity (IC_{50%}) of dry faba bean seeds.

Faba beans are known to be rich in proteins and contain abundant dietary fibers that could contribute to anti-IBD activities through modulating gut microbiota and preventing gut dysbiosis.^{51,52} A previous study conducted by Papoutsis *et al.*⁵³ investigated the role of protein and fiber fractions of faba beans for colonic health and microbiota composition in a low-grade inflammation mouse model. They found that faba bean fractions had minor effects on inflammatory parameters and colonic microbiota. Studying the effect of dietary intake of faba bean seeds on colonic microbiota was beyond the scope of the current study and the main concern was the effect on the antioxidant indices in colon tissues. A significant correlation was found between the total protein content of dry faba bean seeds and the antioxidant indices in the colon tissues. This effect of faba bean proteins may be attributed to the amino acid profiles or the presence of bioactive peptides produced during the digestion of plant proteins as previously reported.^{54,55}

ROS play an essential role in the pathophysiology of UC. In fact, UC patients exhibit lower antioxidant capacity and reveal greater levels of oxidative DNA damage than healthy individuals.^{56,57} SOD, an endogenous antioxidant enzyme, converts superoxide to H₂O₂ in the colonic epithelium, while GSH, a non-enzymatic antioxidant, captures ROS and is converted to the oxidized form GSSG. GSH reductase (GRD) keeps GSH in its reduced state.⁵⁸ MDA is an end product of the lipid peroxidation process, which induces metabolic aberrations and results in cross-links with DNA proteins with consequent DNA breaks.⁵⁹ Such an oxidative imbalance stimulates the inflammatory cells to produce peroxynitrite, which establishes oxidative stress in UC.⁶⁰ The results of the current study are in agreement with these findings, as AA administration led to a remarkable oxidative imbalance. Feeding with pre-treated faba bean seeds, especially in the Se, L + Se and G + L + Se groups, decreased oxidative stress (decreased colonic MDA) and increased antioxidant defense mechanisms (increased colonic GSH, GRD, SOD and CAT) as a protective approach. Likewise, treatment with antioxidants such as N-acetylcysteine, the GSH precursor,⁶¹ or the flavonoid quercetin⁶² reduced colitisinduced oxidative stress.

Phenolics and flavonoids are among the most important phytonutrients contained in faba beans. These compounds occur ubiquitously in plant-based diets or medicinal plants, and faba beans are rich in these compounds.⁶³ Previous studies reported different therapeutic effects of phenolics and flavonoids due to their natural anti-oxidant, anti-carcinogenic,⁶⁴ anti-ulcer,⁶⁵ anti-inflammatory,⁶⁶ immunomodulatory,⁶⁷ and anti-microbial⁶⁸ activities.

In order to show the impact of the different treatments on the phenolic and flavonoid content of faba bean seeds, the TPC, TFC and antioxidant capacity of the dry faba bean seed extract were determined. The obtained results showed high levels of phenolic and flavonoid contents of faba bean seeds in different groups and this confirms the fact that faba bean is a rich source of phenolic and flavonoid compounds. These results are consistent with those reported previously for faba bean seeds.^{69,70} Interestingly, the G + L + Se, L + Se and Se treatments, respectively, increased the TPC, TVC and antioxidant capacity in faba bean seeds as compared to the untreated beans (control group).

Correlations between the TPC and TFC and antioxidant capacity of the dry faba bean seed extract were analyzed. The obtained data revealed that the TPC and TFC in the faba bean seed extract strongly influenced the antioxidant capacity of the extract. Moreover, significant correlations were found between the TPC, TFC and antioxidant capacity of the dry faba bean seed extract and the antioxidant indices in the colon tissues (SOD activity, GSH level and GRD activity). Therefore, phenolic and flavonoid compounds may contribute to disease attenuation through their ability to reduce oxidative stress. In addition, previous studies, using various experimental colitis models, found down-regulation of pro-inflammatory cytokines in animals fed pure phenolic compounds.^{71,72} Since faba beans are a rich source of phenolic compounds,^{69,70} it is possible that the reduction in colitis symptoms observed in this study is due to the down-regulation of pro-inflammatory cytokines, but this needs to be investigated in further studies.

5 Conclusions

In conclusion, it became clear to us that planting faba bean seeds pre-treated with a mixture of selenium, garlic husk extract and lemon peel extract increased the faba beans' phenolic, flavonoid, Se, N, K and protein content with a subsequent enhancement in their antioxidant capacity. Consumption of such faba beans has potential protective and therapeutic effects during experimental colitis, as it reduces colonic oxidative stress and increases colonic antioxidant defense mechanisms. Further research is required to understand the mechanisms by which such faba beans influence colitis, their effects on various inflammatory biomarkers and their impact on the severity of colitis in humans.

Author contributions

Salwa M. El-sayed and Mona I. Nossier conceived and designed the research, performed the research and acquired the data; Ahmed I. Nossier analyzed and interpreted the data and drafted the manuscript. All authors were involved in revising the manuscript.

Conflicts of interest

There are no conflicts to declare.

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References

1 K. Crépon, P. Marget, C. Peyronnet, B. Carrouee, P. Arese and G. Duc, Nutritional value of faba bean (*Vicia faba* L.) seeds for feed and food, *Field Crops Res.*, 2010, **115**, 329– 339.

- 2 M. V. Patto, R. Amarowicz, A. Aryee, J. Boye, H. Chung, M. MartÃn-Cabrejas and C. Domoney, Achievements and challenges in improving the nutritional quality of food legumes, *Crit. Rev. Plant Sci.*, 2015, **34**, 105–143.
- 3 A. Septembre-Malaterre, F. Remize and P. Poucheret, Fruits and vegetables, as a source of nutritional compounds and phytochemicals: Changes in bioactive compounds during lactic fermentation, *Food Res. Int.*, 2018, **104**, 86–99.
- 4 S. Boukhanouf, H. Louaileche and D. Perrin, Phytochemical content and in vitro antioxidant activity of faba bean (*Vicia faba* L.) as affected by maturity stage and cooking practice, *Int. Food Res. J.*, 2016, 23(3), 954–961.
- 5 J. Ryu, D.-G. Kim, M.-K. Lee, J. M. Kim, M. J. Hong, K.-Y. Kang, S. H. Eom, S.-Y. Kang, J.-B. Kim and S.-J. Kwon, Fatty acid composition, isoflavone and L-3, 4-dihydroxyphenylalanine (L-dopa) contents in different parts of faba bean (*Vicia faba*) genotypes, *Plant Breed. Biotechnol.*, 2017, 5, 314–324.
- 6 E. B. Kurutas, The importance of antioxidants which play the role in cellular response against oxidative/nitrosative stress: Current state, *Nutr. J.*, 2015, **15**, 1–22.
- 7 M. Kieliszek and S. Błażejak, Current knowledge on the importance of selenium in food for living organisms: A review, *Molecules*, 2016, **21**, 609.
- 8 S. Misra, M. Boylan, A. Selvam, J. E. Spallholz and M. Björnstedt, Redox-active selenium compounds – From toxicity and cell death to cancer treatment, *Nutrients*, 2015, 7, 3536–3556.
- 9 S. P. Short and C. S. Williams, Selenoproteins in tumorigenesis and cancer progression, *Adv. Cancer Res.*, 2017, 136, 49–83.
- 10 R. Yang, Y. Liu and Z. Zhou, Selenium and selenoproteins, from structure, function to food resource and nutrition, *Food Sci. Technol. Res.*, 2017, **23**, 363–373.
- 11 F. Combs Jr., Biomarkers of selenium status, *Nutrients*, 2015, 7, 2209–2236.
- 12 G. Diretto, A. Rubio-Moraga, J. Argandoña, P. Castillo, L. Gómez-Gómez and O. Ahrazem, Tissue-specific accumulation of sulfur compounds and saponins in different parts of garlic cloves from purple and white ecotypes, *Molecules*, 2017, **22**, 1359.
- 13 K. A. Szychowski, K. Rybczynska-Tkaczyk, K. Gawel-Beben, M. Swieca, M. Karas, A. Jakuczyk, M. Matysiak, U. E. Binduga and J. Gminski, Characterization of active compounds of different garlic (*Allium sativum L.*) cultivars, *Pol. J. Food Nutr. Sci.*, 2018, **68**(1), 73–81.
- 14 A. Shang, S. Cao, X. Xu, R. Gan, G. Tang, H. Corke, V. Mavumengwana and H. Li, Bioactive Compounds and Biological Functions of Garlic (Allium sativum L.), *Foods*, 2019, 8(246), 1–31.
- 15 M. Ichikawa, K. Ryu, J. Yoshida, N. Ide, Y. Kodera, T. Sasaoka and R. T. Rosen, Identification of six phenylpropanoids from garlic skin as major antioxidants, *J. Agric. Food Chem.*, 2003, **51**, 7313–7317.
- 16 S. S. Ferreira, A. L. M. Silva and F. M. Nunes, *Citrus reticulata* Blanco peels as a source of antioxidant and anti-prolif-

erative phenolic compounds, Ind. Crops Prod., 2018, 111, 141-148.

- 17 A. T. Abegunde, B. H. Muhammad and T. Ali, Preventive health measures in inflammatory bowel disease, *World J. Gastroenterol.*, 2016, **22**, 7625.
- 18 H. S. De Souza and C. Fiocchi, Immunopathogenesis of IBD: Current state of the art, *Nat. Rev. Gastroenterol. Hepatol.*, 2016, **13**, 13–27.
- 19 Z. Wang, S. Li, Y. Cao, X. Tian, R. Zeng, D.-F. Liao and D. Cao, Oxidative stress and carbonyl lesions in ulcerative colitis and associated colorectal cancer, *Oxid. Med. Cell. Longevity*, 2016, 9875298.
- 20 B. D'Autréaux and M. B. Toledano, ROS as signalling molecules: mechanisms that generate specificity in ROS homeostasis, *Nat. Rev. Mol. Cell Biol.*, 2007, **8**, 813–824.
- 21 M. I. Nossier, S. H. Abd-Elrahman and S. M. El-Sayed, Effect of using garlic and lemon peels extracts with selenium on Vicia faba productivity, *Asian J. Agric. Biol.*, 2022, **4**.
- 22 L. Carbone and J. Austin, Pain and laboratory animals: publication practices for better data reproducibility and better animal welfare, *PLoS One*, 2016, **11**, e0155001.
- 23 P. Hawkins, D. Morton, O. Burman, N. Dennison, P. Honess, M. Jennings, S. Lane, V. Middleton, J. Roughan and S. Wells, A guide to defining and implementing protocols for the welfare assessment of laboratory animals: Eleventh report of the BVAAWF/FRAME/RSPCA/UFAW Joint Working Group on Refinement, *Lab. Anim.*, 2011, 45, 1–13.
- 24 A. Rashidian, P. Dejban, K. K. Fard, A. Abdollahi, M. Chamanara, A. Dehpour and A. Hasanvand, Bupropion ameliorates acetic acid-induced colitis in rat: The involvement of the TLR4/NF-κB signaling pathway, *Inflammation*, 2020, 43, 1999–2009.
- 25 A. Rashidian, H. Keshavarz-Bahaghighat, A. Abdollahi, M. Chamanara, H. Faghir-Ghanesefat, M. Hoseini-Ahmadabadi and A. R. Dehpour, Agmatine ameliorates acetic acid-induced colitis in rats: involvement of nitrergic system, *Immunopharmacol. Immunotoxicol.*, 2019, **41**, 242–249.
- 26 X. Niu, H. Zhang, W. Li, Y. Wang, Q. Mu, X. Wang, Z. He and H. Yao, Protective effect of cavidine on acetic acidinduced murine colitis via regulating antioxidant, cytokine profile and NF-κB signal transduction pathways, *Chem.-Biol. Interact.*, 2015, **239**, 34–45.
- 27 M. M. Pastrelo, C. C. D. Ribeiro, J. W. Duarte, A. P. B. Gollücke, R. Artigiani-Neto, D. A. Ribeiro, S. J. Miszputen, C. T. F. Oshima and A. P. R. Paiotti, Effect of concentrated apple extract on experimental colitis induced by acetic acid, *Int. J. Mol. Cell. Med.*, 2017, 6, 38.
- 28 J. D. Bancroft and M. Gamble, *Theory and practice of histological techniques*, Elsevier health sciences, 2008.
- 29 H. Ohkawa, N. Ohishi and K. Yagi, Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction, *Anal. Biochem.*, 1979, **95**, 351–358.
- 30 M. Nishikimi, N. A. Rao and K. Yagi, The occurrence of superoxide anion in the reaction of reduced phenazine methosulfate and molecular oxygen, *Biochem. Biophys. Res. Commun.*, 1972, 46, 849–854.

- 31 H. Aebi, [13] Catalase in vitro, *Methods Enzymol.*, 1984, 105, 121–126.
- 32 D. Goldberg, and R. Spooner, Assay of Glutathione Reductase, *In: Bergmeyen, H.V., Ed., Methods of Enzymatic Analysis*, 3rd Edition, 1983 vol. 3, pp. 258–265.
- 33 E. Beutler, Improved method for the determination of blood glutathione, *J. Lab. Clin. Med.*, 1963, **61**, 882–888.
- 34 W. Gargouri, S. M. Osés, M. A. Fernández-Muiño, M. T. Sancho and N. Kechaou, Evaluation of bioactive compounds and biological activities of Tunisian propolis, *LWT*, 2019, **111**, 328–336.
- 35 A. Cottenie, M. Verloo, L. Kiekens, G. Velghe and R. Camerlynck, *Chemical analysis of plant and soil laboratory of analytical and agrochemistry*, State Univ., Ghent, Belgium, 1982.
- 36 C. A. Black, Method of soil analysis part 2, *Chemical* and microbiological properties, 1965, vol. 9, pp. 1387– 1388.
- 37 M. L. Jackson, *Soil chemical analysis: Advanced course*, UW-Madison Libraries Parallel Press, 2005.
- 38 J. B. Jones, Laboratory guide for conducting soil tests and plant analysis, CRC Press, 2001.
- 39 M. M. Bradford, A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein–dye binding, *Anal. Biochem.*, 1976, 72, 248–254.
- 40 F. Ribarova and M. Atanassova, Total phenolics and flavonoids in Bulgarian fruits and vegetables, *J. Univ. Chem. Technol. Metall.*, 2005, **40**, 255–260.
- 41 A. Duca, A. Sturza, E.-A. MoacÄf, M. Negrea, V.-D. Lalescu, D. Lungeanu, C.-A. Dehelean, D.-M. Muntean and E. Alexandra, Identification of resveratrol as bioactive compound of propolis from western Romania and characterization of phenolic profile and antioxidant activity of ethanolic extracts, *Molecules*, 2019, 24, 3368.
- 42 R. Fabia, R. Willén, A. Ar'Rajab, R. Andersson, B. Ahren and S. Bengmark, Acetic acid-induced colitis in the rat: A reproducible experimental model for acute ulcerative colitis, *Eur. Surg. Res.*, 1992, **24**, 211–225.
- 43 R. Caruso, B. C. Lo and G. Núñez, Host-microbiota interactions in inflammatory bowel disease, *Nat. Rev. Immunol.*, 2020, 20, 411–426.
- 44 I. N. Mona, S. M. Gawish, T. Taha and M. Mubarak, Response of wheat plants to application of selenium and humic acid under salt stress conditions, *Egypt. J. Soil Sci.*, 2017, 57, 175–187.
- 45 J. Xu, F. Yang, L. Chen, Y. Hu and Q. Hu, Effect of selenium on increasing the antioxidant activity of tea leaves harvested during the early spring tea producing season, *J. Agric. Food Chem.*, 2003, **51**, 1081–1084.
- 46 J. Xu and Q. Hu, Effect of foliar application of selenium on the antioxidant activity of aqueous and ethanolic extracts of selenium-enriched rice, *J. Agric. Food Chem.*, 2004, **52**, 1759–1763.
- 47 N. L. Martins, S. Petropoulos and I. C. Ferreira, Chemical composition and bioactive compounds of garlic (Allium

sativum L.) as affected by pre- and post-harvest conditions: A review, *Food Chem.*, 2016, **211**, 41–50.

- 48 M. H. Mohamed, E. A. Badr, M. S. Sadak and H. H. Khedr, Effect of garlic extract, ascorbic acid and nicotinamide on growth, some biochemical aspects, yield and its components of three faba bean (*Vicia faba* L.) cultivars under sandy soil conditions, *Bull. Natl. Res. Cent.*, 2020, **44**, 1–8.
- 49 R. Casquete, S. M. Castro, A. Martín, S. Ruiz-Moyano, J. A. Saraiva, M. G. Córdoba and P. Teixeira, Evaluation of the effect of high pressure on total phenolic content, antioxidant and antimicrobial activity of citrus peels, *Innovative Food Sci. Emerging Technol.*, 2015, **31**, 37–44.
- 50 B. Singh, J. P. Singh, A. Kaur and N. Singh, Phenolic composition, antioxidant potential and health benefits of citrus peel, *Food Res. Int.*, 2020, **132**, 109114.
- 51 T. P. Trinidad, A. C. Mallillin, A. S. Loyola, R. S. Sagum and R. R. Encabo, The potential health benefits of legumes as a good source of dietary fibre, *Br. J. Nutr.*, 2010, **103**, 569–574.
- 52 S. R. Hertzler, J. C. Lieblein-Boff, M. Weiler and C. Allgeier, Plant proteins: Assessing their nutritional quality and effects on health and physical function, *Nutrients*, 2020, **12**, 3704.
- 53 D. Papoutsis, S. D. C. Rocha, A. M. Herfindal, S. K. Bøhn and H. Carlsen, Intestinal effect of faba bean fractions in WD-fed mice treated with low dose of DSS, *PLoS One*, 2022, 17, e0272288.
- 54 F. Kamran and N. Reddy, Bioactive peptides from legumes: Functional and nutraceutical potential, *Recent Adv. Food Sci.*, 2018, **1**, 134–149.
- 55 L. López-Barrios, J. A. Gutiérrez-Uribe and S. O. Serna-Saldívar, Bioactive peptides and hydrolysates from pulses and their potential use as functional ingredients, *J. Food Sci.*, 2014, **79**, R273–R283.
- 56 J. Pravda, Radical induction theory of ulcerative colitis, *World J. Gastroenterol.*, 2005, **11**, 2371.
- 57 D. Achitei, A. Ciobica, G. Balan, E. Gologan, C. Stanciu and G. Stefanescu, Different profile of peripheral antioxidant enzymes and lipid peroxidation in active and non-active inflammatory bowel disease patients, *Dig. Dis. Sci.*, 2013, **58**, 1244–1249.
- 58 Y.-F. Hsiao, S.-B. Cheng, C.-Y. Lai, H.-T. Liu, S.-C. Huang and Y.-C. Huang, The prognostic role of glutathione and its related antioxidant enzymes in the recurrence of hepatocellular carcinoma, *Nutrients*, 2021, 13, 4071.
- 59 X.-F. Leong, Lipid oxidation products on inflammationmediated hypertension and atherosclerosis: A mini review, *Front. Nutr.*, 2021, **8**, 717740.
- 60 J. Toro-Pérez and R. Rodrigo, Contribution of oxidative stress in the mechanisms of postoperative complications and multiple organ dysfunction syndrome, *Redox Rep.*, 2021, **26**, 35–44.
- 61 I. Amrouche-Mekkioui and B. Djerdjouri, N-Acetylcysteine improves redox status, mitochondrial dysfunction, mucindepleted crypts and epithelial hyperplasia in dextran sulfate sodium-induced oxidative colitis in mice, *Eur. J. Pharmacol.*, 2012, **691**, 209–217.

- 62 C. F. Guazelli, V. Fattori, B. B. Colombo, S. R. Georgetti, F. T. Vicentini, R. Casagrande, M. M. Baracat and W. A. Verri Jr., Quercetin-loaded microcapsules ameliorate experimental colitis in mice by anti-inflammatory and antioxidant mechanisms, *J. Nat. Prod.*, 2013, **76**, 200–208.
- 63 I. M. Abu-Reidah, M. del Mar Contreras, D. Arráez-Román,
 A. Fernández-Gutiérrez and A. Segura-Carretero,
 UHPLC-ESI-QTOF-MS-based metabolic profiling of *Vicia faba* L.(Fabaceae) seeds as a key strategy for characterization in foodomics, *Electrophoresis*, 2014, 35, 1571–1581.
- 64 W. Y. Jeong, J. S. Jin, Y. A. Cho, J. H. Lee, S. Park, S. W. Jeong, Y. H. Kim, C. S. Lim, A. A. El-Aty and G. S. Kim, Determination of polyphenols in three *Capsicum annuum* L.(bell pepper) varieties using high-performance liquid chromatography-tandem mass spectrometry: Their contribution to overall antioxidant and anticancer activity, *J. Sep. Sci.*, 2011, 34, 2967–2974.
- 65 Z. Zakaria, E. A. Hisam, M. Rofiee, M. Norhafizah, M. Somchit, L. Teh and M. Salleh, In vivo antiulcer activity of the aqueous extract of *Bauhinia purpurea* leaf, *J. Ethnopharmacol.*, 2011, 137, 1047–1054.
- 66 I. N. Beara, M. M. Lesjak, D. Z. Orčić, N. Đ. Simin, D. D. Četojević-Simin, B. N. Božin and N. M. Mimica-Dukić, Comparative analysis of phenolic profile, antioxidant, anti-inflammatory and cytotoxic activity of two closely-related Plantain species: *Plantago altissima* L. and

Plantago lanceolata L, LWT – Food Sci. Technol., 2012, 47, 64–70.

- 67 A. R. Zimmer, B. Leonardi, D. Miron, E. Schapoval, J. R. de Oliveira and G. Gosmann, Antioxidant and anti-inflammatory properties of *Capsicum baccatum*: From traditional use to scientific approach, *J. Ethnopharmacol.*, 2012, **139**, 228– 233.
- 68 J. C. Silva, S. Rodrigues, X. s. Feás and L. M. Estevinho, Antimicrobial activity, phenolic profile and role in the inflammation of propolis, *Food Chem. Toxicol.*, 2012, **50**, 1790–1795.
- 69 Y.-h. Lu, C.-r. Tian, C.-y. Gao, B.-n. Wang, W.-y. Yang, X. Kong, L.-q. Chai, G.-c. Chen, X.-f. Yin and Y.-h. He, Phenolic composition, antioxidant capacity and inhibitory effects on α -glucosidase and lipase of immature faba bean seeds, *Int. J. Food Prop.*, 2018, **21**, 2366–2377.
- 70 W. Rybiński, M. Karamać, K. Sulewska and R. Amarowicz, in *Plant Extracts*, IntechOpen, 2019.
- 71 K. H. Kwon, A. Murakami, T. Tanaka and H. Ohigashi, Dietary rutin, but not its aglycone quercetin, ameliorates dextran sulfate sodium-induced experimental colitis in mice: attenuation of pro-inflammatory gene expression, *Biochem. Pharmacol.*, 2005, **69**, 395–406.
- 72 M.-Y. Park, G. E. Ji and M.-K. Sung, Dietary kaempferol suppresses inflammation of dextran sulfate sodium-induced colitis in mice, *Dig. Dis. Sci.*, 2012, **57**, 355–363.