ROLE OF PLATELET DERIVED GROWTH FACTOR IN CORNEAL HEALING

Thesis
Submitted for partial fulfillment of M.D. degree
In Ophthalmology

By Tamer Hassan El-Sersy (M.B.,B.Ch.,M.Sc.) Under The Supervision Of

Professor Dr. Amira Mohammed Mounir
Professor of Ophthalmology
Ain Shams University

Professor Dr. Mamdouh Hamdy El-Kafrawy
Professor of Ophthalmology
Ain Shams University

Professor Dr. Rafik Mohammed El- Ghazzawy
Professor of Ophthalmology
Ain Shams University

Professor Dr. Sanaa Abd El-Maged Sammour
Professor of Histopathology
Ain Shams University

Dr. Manal Fawzy Ghazlan
Assistant Professor of Clinical Pathology
Ain Shams University

Faculty of Medicine Ain Shams University Cairo-2006

دور عنصر النمو المشتق من الصفائح الدمويه في التئام القرنيه

رساله مقدمه توطئة للحصول على درجة الدكتوراه في طب و جراحة العيون مقدمه من الطبيب/ تامر حسن السرسي تحت اشراف

الاستاذه الدكتوره/ اميره محمد منير استاذ طب و جراحة العيون كلية الطب جامعة عين شمس

الاستاذ الدكتور/ ممدوح حمدى الكفراوى استاذ طب و جراحة العيون كلية الطب جامعة عين شمس

الاستاذ الدكتور/ رفيق محمد الغزاوى استاذ طب و جراحة العيون كلية الطب جامعة عين شمس

الاستاذه الدكتوره/ سناء عبد الماجد سمور استاذ الباثولوجي كلية الطب جامعة عين شمس

دكتوره/ منال فوزى غزلان استاذ مساعد الباثولوجيا الإكلينيكيه كلية الطب جامعة عين شمس

> كلية الطب جامعة عين شمس القاهره- 2006

Acknowledgement

I wish to express my deepest thanks and profound gratitude to **Professor Dr. Amira Mohammed Mounir**, Professor of Ophthalmology, Ain Shams University, to whom I am deeply indebted for her encouragement valuable supervision and continuous help during the developement of this work.

I am also extremely grateful to **Professor Dr.**Mamdouh El-Kafrawy, Professor of Ophthalmology Ain

Shams University, for his invaluable instructions, inspiring guidance and support which were crucial in the completion of this work.

I sincerely appreciate all the encouragement and support given to me by **Professor Dr. Rafik El-Ghazawy**, Professor of Ophthalmology ,Ain Shams University, whose invaluable guidance was of a pivotal role in the completion of this work.

May deepest appreciation and thanks are in order to **Professor Dr. Sanaa Samoor**, Professor of histopathology, Ain Shams University, towhom I am indebted for her meticulous supervision and kindness.

My greatest appreciation and thanks are due to **Professor Dr. Manal Ghazlan,** Assistant Professor of Clinical Pathology, Ain Shams University, whose continuous remarks and unforgettable support were of great importance in accomplishing this work.

FIGURE	TITLE	PAGE
Figure 1	Genomic organizations of PDGF	6
	family genes.	
Figure 2	Pseudomonas bacterial corneal ulcer.	30
Figure 3	Herpes Simplex corneal ulcer.	33
Figure 4	Fungal Keratitis.	37
Figure 5	Neurotrophic corneal ulcer.	39
Figure 6	Recurrent corneal erosion.	40
Figure 7	Mooren's ulcer.	42
Figure 8	Variable-speed centrifuge.	47
Figure 9	Supernatent platelet-rich plasma.	47
Figure 10	Bottle containing thrombin.	48
Figure 11	Ulcer size among human infective and	55
	non-infective eyes (pre-treatment).	
Figure 12	Ulcer size among human infective and	56
	non-infective eyes (1 week post	
	treatment).	
Figure 13	Ulcer size among human infective and	57
	non-infective eyes (2 weeks post	
	treatment).	
Figure 14	Ulcer size among human infective and	58
	non-infective eyes (4 weeks post	
	treatment).	
Figure 15	Ulcer size among non-infective eyes	60
	(pre-treatment to 1 week post PDGF).	
Figure 16	Ulcer size among non-infective eyes	61
	(pre-treatment to 2 week post PDGF).	
Figure 17	Slit lamp photography of a patient with	62
	peripheral corneal melting	
	secondary to rheumatoid arthritis pre-	
	treatment and 2weeks post PDGF.	
Figure 18	Ulcer size among non-infective eyes	63
	(pre-treatment to 4 week post PDGF).	

Figure 19	Slit lamp photography of a patient with non-infective neurotrophic ulcer pretreatment and 4 weeks post PDGF.	64
Figure 20	Ulcer size among non-infective eyes (1 week to 2 weeks post PDGF).	61
Figure 21	Ulcer size among non-infective eyes (1 week to 4 weeks post PDGF).	66
Figure 22	Slit lamp photography of a patient with an non-infective ulcer 1 week and 4 weeks post PDGF.	67
Figure 23	Ulcer size among non-infective eyes (2 week to 4 weeks post PDGF).	68
Figure 24	Ulcer size among human infective eyes (pre-treatment to 1 week post PDGF).	69
Figure 25	Slit lamp photography of a patient with an infective ulcer pre-treatment and 1 week post PDGF.	70
Figure 26	Ulcer size among human infective eyes (pre-treatment to 2 week post PDGF).	71
Figure 27	Slit lamp photography of a patient with an infective ulcer pre-treatment and 2 week post PDGF.	72
Figure 28	Ulcer size among human infective eyes (pre-treatment to 4 week post PDGF).	73
Figure 29	Slit lamp photography of a patient with an infective ulcer pre-treatment and 4 week post PDGF.	74
Figure 30	Ulcer size among human infective eyes (1 week to 2 weeks post PDGF).	75
Figure 31	Ulcer size among human infective eyes (1 week to 4 weeks post PDGF).	76
Figure 32	Ulcer size among human infective eyes (1 week to 4 weeks post PDGF).	77

Figure 33	Ulcer size among human infective eyes (2 week to 4 weeks post PDGF).	78
Figure 34	Sex distribution among non-infected eyes group in human.	79
Figure 35	Sex distribution among infected eyes group in human.	79
Figure 36	Histopathology of normal rabbit cornea. H & EX 200	83
Figure 37	Histopathological section of infected rabbit cornea 1 week after ulcer infliction without PDGF. H & EX 200	84
Figure 38	Histopathological section of infected rabbit cornea 1 week after ulcer infliction with PDGF. H & EX 200	84
Figure 39	Histopathological section of non- infected rabbit cornea 1 week after ulcer infliction without PDGF. H & EX 200	85
Figure 40	Histopathological section of non- infected rabbit cornea 1 week after ulcer infliction with PDGF. H & EX 200	85
Figure 41	Histopathological section of infected rabbit cornea 2 weeks after ulcer infliction without PDGF. H & EX 200	86
Figure 42	Histopathological section of non- infected rabbit cornea 2 weeks after ulcer infliction without PDGF. H & EX 200	86

Figure 43	Histopathological section of infected	87
	rabbit cornea 3 weeks after ulcer	
	infliction without PDGF.	
	H & EX 200	
Figure 44	Histopathological section of infected	87
	rabbit cornea 3 weeks after ulcer	
	infliction with PDGF. H&EX200	
Figure 45	Histopathological section of non-	88
	infected rabbit cornea 3 weeks after	
	ulcer infliction without PDGF.	
	H & EX 200	
Figure 46	Histopathological section of non-	88
	infected rabbit cornea 3 weeks after	
	ulcer infliction with PDGF.	
	H & EX 200	
Figure 47	Histopathological section of infected	89
	rabbit cornea 6 weeks after ulcer	
	infliction with PDGF.	
	H & EX 200	
Figure 48	Histopathological section of non-	89
	infected rabbit cornea 6 weeks after	
	ulcer infliction with PDGF.	
	H & EX 200	
Figure 49	Epithelization among rabbits.	91
Figure 50	Vascularization among rabbits	93
Figure 51	Stromal infiltration among rabbits	95
Figure 52	Stromal fibrosis among rabbits	97
Figure 53	Stromal hyalinization among rabbits	99

LIST OF ABBREVIATIONS

bFGF basic Fibroblast growth factor BMP Bone morphogenic protein

C3 Complement type 3

cAMP cyclic Adenosine mono phosphate CCLRU Cornea and Contact lens Research

CNS Central nervous system
CSF-1 Colony stimulating factor-1
EGF Epidermal growth factor

FN Fibronectin GC Glucocorticoid

HGF Hepatocyte growth factor HSV Herpes Simplex Virus

HZO Herpes Zoster Ophthalmicus

Ig Immunoglobulin

IGF Insulin Growth Factor

IL-1 Interleukin-1 IL-6 Interleukin-6

KGF Keratinocyte growth factor
MALT Mucosal Associated Lymphoid
MMP Matrix metalloproteinases
PAP PDGF-associated protein
PDGF Platelet derived growth factor

PKC Protein kinase C

proteinases

SCF Stem cell factor

SPE Sis proximal element

TGF Transforming growth factor TIMP Tissue inhibitors of metallo-

Tissue

TNF Tumour necrosis factor

Unit

US United States

VEGF Vascular endothelial growth factor

VZV Varicella-Zoster Virus

LIST OF TABLES

TABLE	TITLE	PAGE
Table 1	Normal cell types expressing PDGF	7
Table 2	Cellular effects mediated by PDGF α- and	11
	β- receptor homodimers	
Table 3	Cell type expression PDGF receptors	12
Table 4	Chi square comparing ulcer size among non	55
	infective and infective eyes (before starting	
	treatment).	
Table 5	Chi square comparing ulcer size among non	56
	infective and infective eyes (1 week post treatment).	
Table 6	Chi square comparing ulcer size among non	57
Table 0	infective and infective eyes (2 weeks post	37
	treatment).	
Table 7	Chi square comparing ulcer size among non	58
	infective and infective eyes (4 weeks post	
T. 11.0	treatment).	60
Table 8	Chi square comparing ulcer size among non infective eyes before treatment and 1 week	60
	post PDGF treatment.	
Table 9	Chi square comparing ulcer size among non	61
Tuble	infective eyes before treatment and 2 weeks	01
	post PDGF treatment	
Table 10	Chi square comparing ulcer size among non	63
	infective patients before treatment and 4	
Table 11	weeks post PDGF treatment.	65
Table 11	Chi square comparing ulcer size among non infective eyes at 1 week and 2 weeks post	05
	PDGF treatment.	
Table 12	Chi square comparing ulcer size among non	66
	infective eyes at 1 week and 4 weeks post	
	PDGF treatment.	
Table 13	Chi square comparing ulcer size among non	68
	infective eyes at 2 weeks and 4 weeks post PDGF treatment.	
Table 14	Chi square comparing ulcer size among	69
1 avic 14	infective eyes before starting treatment to 1	UF
	week post PDGF therapy.	

LIST OF TABLES

Chi square comparing ulcer size among	71
infective eyes before starting treatment to 2	
weeks post PDGF therapy.	
Chi square comparing ulcer size among	73
infective eyes before starting treatment to 4	
weeks post PDGF therapy.	
Chi square comparing ulcer size among	75
infective eyes 1 week and 2 weeks post PDGF	
therapy.	
Chi square comparing ulcer size among	76
infective eyes 1 week and 4 weeks post PDGF	
therapy.	
Chi square comparing ulcer size among	78
infective eyes 2 and 4 weeks post PDGF	
therapy.	
Epithelization among rabbit eyes.	90
Vascularization among rabbit eyes.	92
Stromal infiltration among rabbit eyes.	94
Stromal fibrosis among rabbit eyes.	96
Stromal hyalinization among rabbit eyes.	98
	infective eyes before starting treatment to 2 weeks post PDGF therapy. Chi square comparing ulcer size among infective eyes before starting treatment to 4 weeks post PDGF therapy. Chi square comparing ulcer size among infective eyes 1 week and 2 weeks post PDGF therapy. Chi square comparing ulcer size among infective eyes 1 week and 4 weeks post PDGF therapy. Chi square comparing ulcer size among infective eyes 2 and 4 weeks post PDGF therapy. Epithelization among rabbit eyes. Vascularization among rabbit eyes. Stromal infiltration among rabbit eyes. Stromal fibrosis among rabbit eyes.

CONTENTS

INTRODUCTION	1-2
AIM OF WORK	3
REVIEW OF LITERATURE	4-44
 Mechanism of action & in vivo role of PDGF 	4-20
 Mechanism of corneal ulceration 	21-27
■ Types of corneal ulcers	28-44
PATIENTS AND METHODS	45-53
RESULTS	54-100
DISCUSSION	101-108
CONCLUSION AND RECOMMENDATION -	109-110
SUMMARY	111-113
REFERENCES	114-123
ARABIC SUMMARY	

AIM OF WORK

AIM OF WORK

Clinical evaluation of the rate of corneal healing under the effect of Platelet Derived Growth Factor, together with histopathological examination of the corneal tissues of the experimental animals.

CONCLUSION & RECOMMENDATION

Our study included both human and animal studies and was targeted to evaluate the role of PDGF in the healing of resistant infective and non-infective corneal ulcers.

The animal study was done to endorse our clinically based study through histopathological examination of the rabbits corneas.

The human study showed the following facts:

Although there was a statistically significant difference in ulcer size between the infective and non-infective groups at the beginning of the study, however, by the end of the study we found this difference did not influence the response of both groups top the PDGF.

Concerning both the infective and non- infective groups:

There was a highly statistically significant decrease in ulcer size during the first week of the study. The size reduction continued during the second week, however, during the period extending along the third and fourth weeks there was no statistical difference in size reduction in both the infective and non- infective groups. Also there was no regression in the healing of the corneal ulcer in the last two weeks of the study (i.e. a relatively stationary course in the last two weeks).

From this we can deduce that the maximal effect of PDGF occurs during the first week of therapy and it continues till the second week thereafter no change will occur denoting that the followup period for the improvement can be restricted to two weeks only.

CONCLUSION & RECOMMENDATION

The animal study showed the following facts:

There marked difference in the rate of epithelization in the first week being more rapid with the use of PDGF in both the infective and non-infective eyes.

By the end of the second week there complete epithelization of all eyes with or without PDGF.

From this we can deduce that the maximum effect of PDGF on epithelial healing occurs during the first week of therapy regardless of the nature of the ulcer.

Concerning vascularization, there is decreased vascularization in the eyes receiving PDGF in the first week with equal mild vascularization by the second week of the study. There is earlier disappearance of vascularization in the eyes Receiving PDGF.

Thus it is safe to say that PDGF decreases initial vascularization of the ulcer and also promote early regression of the vascularization.

There was not a statistically significant impact of PDGF on stromal infiltration.

As regards stromal fibrosis, there was a statistically significant difference in the degree of fibrosis being more in the eyes receiving PDGF. This endorses the possibility of presence of chemtactic properties of PDGF attracting fibroblasts to the ulcer site.

As regards stromal hyalinization, there was a statistically significant difference in the degree of hyalinization denoting that PDGF promotes stromal hyalinization.

INTRODUCTION

INTRODUCTION

Throughout the 1980's, the focal point of wound healing knowledge was the role of tissue oxygenation. Today, however, the focal point of an expanding knowledge base are the identification, understanding, and now the means to use growth factors to enhance wound healing (Andresen et al., 1997).

The mechanism of corneal wound healing has not been clarified yet. However, evidence has accumulated that various kinds of growth factors such as epidermal growth factor (EGF), fibroblast growth factor (FGF), transforming growth factor (TGF) and platelet derived growth factor (PDGF) play a key role in corneal wound healing. For example, these factors are expressed in corneal epithelial cells, keratocytes and endothelial cells, and their receptors are expressed in the corneal cells. Thus these growth factors function in the regulation of corneal cell proliferation and in the maintenance of corneal transparency (Imanishi et al., 2000).

PDGF is a glycoprotein with a molecular mass of approximately 30 kd. Although it is the primary growth factor in platelets, it is also synthesized and secreted by other cells, such as macrophages and endothelial cells (**Tsubota et al., 1999**).

Platelets are concentrated in a small volume of plasma to form platelet rich plasma (PRP). If thrombin and calcium are added to the PRP, the platelets are activated to release their content, including PDGF. The thrombin and calcium also initiate clotting, including the conversion of fibrinogen to fibrin, resulting in a clinically useful platelet-rich gel (Andresen and Ehlers, 1998).