COMPARATIVE STUDIES ON VIRAL INDUCIBLE PROTEIN IN SUSPENSION CULTURE OF SOME SOLANACEAE PLANTS

By

ALAA MOHAMED EL-SAEED EL-MINISY

B.Sc. Agric. Sci. (Biotechnology), Fac. Agric., Cairo Univ., 2008

THESIS

Submitted in Partial Fulfillment Of the Requirements for the Degree of

MASTER OF SCIENCE

In

Agricultural Sciences

Department of Genetics
Faculty of Agriculture
Cairo University
EGYPT
2017

APPROVAL SHEET

COMPARATIVE STUDIES ON VIRAL INDUCIBLE PROTEIN IN SUSPENSION CULTURE OF SOME SOLANACEAE PLANTS

Master Thesis In Agric. Sci. (Genetics)

 $\mathbf{B}\mathbf{y}$

ALAA MOHAMED EL-SAEED EL-MINISY

B.Sc. Agric. Sci. (Biotechnology), Fac. Agric., Cairo Univ., 2008

APPROVAL COMMITTEE

Date: / /

SUPERVISION SHEET

COMPARATIVE STUDIES ON VIRAL INDUCIBLE PROTEIN IN SUSPENSION CULTURE OF SOME SOLANACEAE PLANTS

Master Thesis
In
Agric. Sci. (Genetics)

By

ALAA MOHAMED EL-SAEED EL-MINISY

B.Sc. Agric. Sci. (Biotechnology), Fac. Agric., Cairo Univ., 2008

SUPERVISION COMMITTEE

Dr. MOHAMED HASSANIEN SOLIMAN Professor of Genetics, Fac. Agric., Cairo University

Dr. SALAH EL-DIN SAYED MOHAMED EL-ASSAL Professor of Genetics, Fac. Agric., Cairo University

Dr. ADEL EL-SAWY MOHAMED
Professor, Plant Biotechnology department, National Research Center

ACKNOWLEDGEMENT

All thanks and praises to thank ALMIGHTY ALLAH, the most merciful and beneficent, who gave me the ability and knowledge to complete this study. I am grateful to my family, my parents and to my brothers and sisters for supporting me spiritually not only throughout writing this thesis but also my life in general. Also to friends who have supported me along the way.

I would like to express appreciation to a number of people whose contributions have served to make this work possible. First and foremost, I am grateful to my adviser, **Dr. Mohamed H. Soliman,** Professor of Genetics, Faculty of Agriculture, Cairo University for his supervision

I would like also to express my deep appreciations and utmost gratitude to **Dr. Salah el-Din Sayed El-Assal**, Professor of Genetics, Faculty of Agriculture, Cairo University for his supervision of this investigation, motivation, valuable guidance, and helping me during the course of this work.

My special thanks to **Dr. Adel El-Sawy**, Professor, Plant biotechnology Department, National Research Center, for his valuable help and encouragement during this work.

Also, my special and deep thanks to my advisor and mentor, **Dr. Hattem M. El-Shabrawi**, Associate professor, Plant biotechnology Department, National Research Center, for his supervision, unlimited help, and encouragement during this work. I am also grateful to him for his patience and support in overcoming numerous obstacles I have been facing through my research.

I am also grateful to the following department staff: **Dr. Ahmed Magdy**, **Dr.Hussein Taha** for their unfailing support and assistance.

Thanks are also extended to the staff members of the Plant Biotechnology Department, National Research Center for their help, encouragement, and the facilities provided during the work of this thesis.

And finally, last but by no means least, also to everyone in the laboratory... it was great sharing laboratory with all of you during last six years.

Name of Candidate: Alaa M.El-Minisy Degree: M.Sc.

Title of Thesis: Comparative Studies on Viral Inducible Protein in Suspension Culture of *Solanaceae* Plants

Supervisors: Dr. Mohamed Hassanein Soliman

Dr. Salah El-Din Sayed Mohamed El-Assal

Department: Genetics Approval: 4 /7/ 2017

ABSTRACT

Potato virus Y (PVY), a member of Potyvirus genus, is one of the most important potato viruses and is the causal agent of several plant diseases in a wide range of host species, causing important economic losses in agriculture. These studies were conducted during the period from 2013 to 2016 at the Department of Plant Biotechnology, National Research Center, Cairo, Egypt, to identify and detect the proteins involved in response to PVY infections in N.tabacum Samsun CV. In the first experiment, Total soluble proteins were extracted from N.tabacum leaf tissue inoculated with PVY, Protein changes in both healthy and inoculated tobacco leaves were revealed using one dimensional gel electrophoresis (1D-SDS). Proteins that were highly and significantly changed are selected for identification by liquid chromatography mass spectrometry (LC-MS/MS) combined with bioinformatics. These data provide a valuable resource for discovering novel proteins involved in the pathogen response. Such proteins could be introduced into agronomical important species to develop a viral resistant crop plants. A total of 470 proteins were confidently identified, with a predominance of proteins associated with photosynthesis, energy and metabolism and response to stimulus. The second experiment was conducted to study the role of PAP in plant response against PVY infection using N.tabacum cv. Samsun. To achieve this goal, pokeweed micropropagation protocol was achieved. After 12 days of inoculation, N. tabacum plants pre-treated with 0.1 mg/mL pokeweed crude leaf extract before inoculation with PVY showed less symptoms compared to that inoculated with PVY only. Furthermore, the leaf total soluble proteins of PAP treated and untreated PVY infected tobacco plants were analyzed by 1D-SDS. The electrophoresis profile showed variant accumulation levels at two different molecular weight protein bands. Moreover, the spectrometric analysis of those two bands using LC-MS/MS, has identified about 50 different proteins. According to their functions prediction, it has been classified into seven groups including (Energy& metabolism, stress defense response, carbon fixation, photosynthesis, plant cytoskeleton, structural proteins and signaling). We conclude that the PAP may has a direct role in suppression of the PVY infection mechanism. Moreover, PAP protein could be used as a foliar protective agent against PVY infection in tobacco system.

Key words: *Solanaceae*, PVY, Viral inducible protein, 1 D-SDS, LC-MS/MS.

CONTENTS

INTRODUCTION	
REVIEW OF LITERATURE	
1. Solanaceae family and Plant viruses	
2. Potato virus Y (PVY)	
a. Host range	
b. Symptomology	
c. Virus particles	
d. Molecular characterization	
e. Detection methods	
f. Natural resistance to PVY	
g. Spread and accumulation of Potyviruses in pla	ant
3. Changes in PVY infected plant at metabolic levels	vel
a. Antioxidant metabolism	
b. Pathogenesis-Related Proteins	
c. Heat shock proteins	
4. Antiviral protein (Pokeweed)	
5. Mass spectrometry analysis in protein sequence	ing
MATERIALS AND METHODS	
1. Plant materials	
a. In vitro culture	
b. N.tabacum cv. Samsun Plants	
2. Virus source	
3. Confirmation of PVY isolate	
4. Detection of plant protein response due to P	
infection and PAP treatment	
RESULTS AND DISCUSSION	
1. In vitro culture	
2. Confirmation of PVY isolate	
a. Symptomology on different differential host	
b. Detection of PVY using Transmission electron	
microscopy	

c.	Detection of PVY using RT-PCR	58
infect	etection of plant protein response due to PVY tion and exogenous application of PAP crude extract (Pokeweed Antiviral Protein)	61
a.	Phenotypic reactions of PVY inoculated <i>N.tabacum</i> and non-inoculated one	61
	Histochemical detection of superoxide anion accumulation and hydrogen peroxide using Nitro blue tetrazolium (NBT) and Diaminobenzidine tetrahydrochloride(DAB), respectively	63
	infection	68
d.	Identification and analysis of the excised proteins by LC-MS/MS	71
e.	PAP treatment before PVY inoculation	82
SUM	MARY	101
REF	ERENCES	114
ARA	BIC SUMMARY	

LIST OF TABLES

rate	18
rate	18
rate	18
rate	
rate	
of	51
	53
<i>N</i> .	
	86
<i>N</i> .	
	88
<i>N</i> .	92
N.	98
62	
	of N N

LIST OF FIGURES

No.	Title	Page
1	PVY genome annotation	7
2	Schematic presentation of plant-virus interaction	11
3.a	In vitro germination of cucumber seeds	44
3.b	Four weeks old cucumber callus derived from	
	hypocotyl explant	44
4.a	Two weeks old <i>Datura metel</i> germinated in pot	46
4.b	Six weeks old Datura metel Callus derived from leaf	
	explant	46
5.a	Shoot multiplication of potato sprout	48
5.b	Four weeks old Potato (Solanum tuberesium) Callus	
	derived from leaf	40
	discs	48
6	Micropropagation of pokeweed	54
7	Symptoms on Nicotiana tabacum ev. Samsun	57
8	Symptoms on <i>Datura metel</i> plants	57
9	Datura metel. Crude sap examined under transmission electron microscope	58
10	PCR results of inoculated <i>Datura metel</i> plants	60
11	Different phenotypic response of N. tabacum plants to	00
	PVY	
	infection	62
12	Different response of N. tabacum plants to PAP and	
	PVY infection	62
13	Histochemical detection of superoxide anion	
	accumulation in N. tabacum inoculated with	
	PVY	65
14	Histochemical detection of superoxide anion	
	accumulation in N. tabacum treated with PAP before	
	inoculated with PVY	66
15	Histochemical detection of hydrogen peroxide	
	accumulation in N. tabacum inoculated with	
	PVY	67
16	Histochemical detection of hydrogen peroxide	
	accumulation in N. tabacum treated with PAP before	
	inoculated with PVY	68

17	12 % SDS-PAGE stained with coomassie blue of total soluble protein extracted from <i>N.tabacum</i> inoculated with PVY	69
18	12 % SDS-PAGE stained with coomassie blue of total soluble protein extracted from <i>N.tabacum</i> inoculated with PVY	70
19	12 % SDS-PAGE stained with coomassie blue of proteins extracted from PAP pretreated <i>Nicotiana</i>	
20	tabacum cv. Samsun plants Functional categorization of identified proteins in Non-inoculated <i>N.tabacum</i>	70 72
21	Functional categorization of identified proteins in PVY inoculated sample at 205 kDa.	72
22	Functional categorization of identified proteins in PVY inoculated sample at 32 kDa	73
23	Functional categorization of identified proteins in PAP treated sample at 62 kDa	73
24	Proteins identified by Thermo EASY nLC II LC system.	74

INTRODUCTION

Potato plants are subjected to attack by several viral diseases, causing harmful effects and reduce crop yield and tuber quality. Viral diseases, particularly potato leaf roll virus (PLRV) and potato virus Y (PVY), are severe problems for Egyptian potato production.

PVY is one of the most important potato viruses and is spread worldwide and recognized as the fifth most important plant virus regarding its scientific and economic importance (Scholthof *et al.*, 2012). It is mainly infect some plants of family *Solanaceae* and causes great yield losses in major crops.

Agricultural crops worldwide suffer from a vast array of pathogens including bacteria, fungi, and viruses which cause tremendous losses in yield and quality of production (Rodoni, 2009). For a long time, these pathogens have been controlled through conventional measures like crop rotation and other cultivation techniques, early detection, destruction of infected source plants, cross-protection, breeding for resistance, and chemical control (Goldbach *et al.* 2003). It has been known that some plants possess special metabolic pathways to synthesize a number of valuable proteins which can be used for prevention and treatment of diseases (Calixto, 2000). For example, plant genes encoding ribosome-inactivating proteins (RIPs) have been shown to confer disease resistance in recent years. RIPs are found not only in a few higher plants but also in fungi, bacteria, and at least one alga (Stirpe and Battelli, 2006).

Attempts to transform potato plants with the pokeweed antiviral protein (PAP), a ribosome-inactivating protein found in the cell walls of *Phytolacca Americana* (pokeweed), thought to give a broad-spectrum protection against several viruses, gave only limited protection against aphid transmitted PVY. When PAP was applied exogenously it protected potato plants from mechanical inoculation with PVY (Lodge *et al.*, 1993).

In recent years, several efforts have been made to study plantvirus interaction at the proteome level in both compatible (susceptible host) and incompatible (resistant host) infections. In both compatible and non-compatible interactions, viruses utilize plant host proteins to complete the infection process. In case of incompatible hostpathogen interactions, damage caused by the pathogen remains restricted because of the plant's defensive response. Plant defensive response is hypersensitive reaction (HR), in which the cells around the infection site rapidly undergo necrosis, integrated with set of metabolic alterations that restricts pathogen ingress (Van Loon and 1999). Strien, Metabolic alterations included defense mechanisms like physical strengthening of the cell wall through lignification, suberization, callose deposition and synthesis of the pathogenesis related (PR) proteins, which include β-1,3-glucanases (PR-2), chitinases (PR-3, -4, -8, and -11) and thaumatin-like proteins(PR-5) (Bowles, 1990). Other HR responses to contain the pathogen is changes in ion fluxes, lipid hyper peroxidation, protein phosphorylation, nitric oxide generation, antimicrobial compounds and burst of reactive oxygen species (ROS).

The ability of plants to defend themselves against pests and disease is associated with a number of proteins that can be up or down-regulated (Afroz, 2011) after challenged. Thus, proteomic analysis to identify host's proteins and their changes in abundance linked to biochemical and cellular processes that control pathogen recognition, defense signal transduction and confer resistance (Zimaro, 2001; Mandelc, 2013) is of paramount importance.

The objective of our study is to detect and identify the proteins involved in the response to PVY infection and exogenous application of crude extract of PAP in tobacco plants. The scope of the study was:

- isolate and identify potato virus Y (PVY).
- mechanically inoculate plants of host range with PVY.
- confirm the inoculation with PVY.
- Pokeweed micropropagation and extract pokeweed antiviral protein (PAP).
- detect the plant protein response due to PVY infection and exogenous application of crude extract of PAP
- in gel digestion of excised selected protein bands for liquid chromatography mass spectrometry (LC MS/MS) analysis.
- LC MS/MS analysis and *in silico* analysis for identification and functional prediction of sequenced proteins (proteomics).

REVIEW OF LITERATURE

1. Solanaceae family and plant viruses

Solanaceae is a family of flowering plants that consists of about 100 genera and 2500 species (Olmstead et al., 2008). Many of them are considered from the world's most important agricultural species, including tomato (Solanum lycopersicum), potato (Solanum tuberosum), egg- plant (Solanum melongena), tobacco (Nicotiana tabacum), pepper (Capsicum annuum) and petunia (Petunia spp.). Also, Solanaceae has many species that have wide variety of uses (Mueller et al., 2005). Considering the economic importance, some Solanaceae plants are important model systems for biology. For example, tomato for fruit ripening and plant defense, tobacco for plant defense, and petunia for the biology of anthocyanin pigments. Wide range of diseases affects solanaceous vegetables. The most important viruses isolated from potato are Potato leaf roll virus (PLRV), Potato virus Y (PVY), Potato virus A (PVA), Potato virus X (PVX) and Potato virus M (PVM) (Valkonen, 1994).

Potato Virus Y (PVY; family Potyviridae, genus Potyvirus), and Potato Leafroll Virus (PLRV; family Luteoviridae, genus Polerovirus), are considered highly important. However, PVY regarded as the most economically significant virus of potato (De Bokx and Van der Want, 1987). Potato leaf roll virus (PLRV) and potato virus Y (PVY), are critical problems for Egyptian potato production. They decrease crop yield and tuber quality.