APPLICATION OF SOME METHODS FOR DETECTING ADULTERATION IN OLIVE OIL

By

RANIA IBRAHIM MOHAMMAD ALMOSELHY

B.Sc. Agric. Sc. (Food Technology), Ain Shams University, 2000

A thesis submitted in partial fulfillment of the requirements for the degree of

MASTER OF SCIENCE

in

Agricultural Science (Food Science and Technology)

Department of Food Science
Faculty of Agriculture
Ain Shams University

2010

Approval Sheet

APPLICATION OF SOME METHODS FOR DETECTING ADULTERATION IN OLIVE OIL

By

RANIA IBRAHIM MOHAMMAD ALMOSELHY

B.Sc. Agric. Sc. (Food Technology), Ain Shams University, 2000

This	This thesis for M.Sc. degree has been approved by:								
							Tashualası		
		eulture, Cai				and	Technology,	racuity	01
Prof.	Dr.	Mamdouh	Hel	my El-	-Kalyoub	i			
		Emeritus culture, Ain				and	Technology,	Faculty	of
Prof.	Dr.	Magda Ha	beel	b Allar	n				
		Emeritus culture, Ain				and	Technology,	Faculty	of

Date of Examination: 29/8/2010

APPLICATION OF SOME METHODS FOR DETECTING ADULTERATION IN OLIVE OIL

By

RANIA IBRAHIM MOHAMMAD ALMOSELHY

B.Sc. Agric. Sc. (Food Technology), Ain Shams University, 2000

Under the supervision of:

Prof. Dr. Magda H. Allam

Prof. Emeritus of Food Science and Technology, Department of Food Science, Faculty of Agriculture, Ain Shams University (Principal Supervisor)

Prof. Dr. Atef A. Kotb

Prof. of Food Science and Technology, Department of Food Science, Faculty of Agriculture, Ain Shams University

Prof. Dr. Ahmad A. El-Sharkawy

Head of Research Emeritus of Oils and Fats Technology, Food Technology Research Institute, Agricultural Research Center

ABSTRACT

Rania Ibrahim Mohammad Almoselhy. Application of Some Methods for Detecting Adulteration in Olive Oil. Unpublished M.Sc. Thesis, Department of Food Science, Faculty of Agriculture, Ain Shams University, 2010.

This study aimed to maintain the high quality of olive oil by investigating some analytical methods as a powerful tool to determine extra virgin olive oil adulteration with relatively cheap seed and vegetable oils such as sunflower oil, corn oil and refined olive oil.

The analytical methods ranged from the familiar manual physical and chemical tests such as refractive index (RI) which gives a good idea about the degree of unsaturation of the oil under investigation, as well as its correlation with iodine value (IV); acid value (AV) as indication of free fatty acid content of the oil; peroxide value (PV) which determines the amount of primary oxidation products and UV absorbencies at 232 and 270 nm, that measure the formation of conjugated dienes and trienes, respectively due to the formation of secondary oxidation products.

The analytical methods extended to the more sophisticated instrumental methods of analysis such as chromatographic separation and determination techniques which involved gas chromatography (GC) with flame ionization detector (FID) in order to investigate the composition of the fatty acids of the oils under investigations.

Fourier Transform Infra Red (FTIR) spectroscopic determination technique was employed and applied as a potent, nondestructive and effective analytical tool to study its potency to investigate the functional groups with their relative absorbencies or transmittances according to their concentrations in samples and their characteristic fingerprints. FTIR spectroscopy was used also to determine extra virgin olive oil adulteration with sunflower, corn and refined olive oils in their binary admixtures at different concentrations of 0, 5, 10, 20, 30, 40, 50, 100%; w/w.

FTIR spectral data collected in MIR range 4000-400 cm⁻¹ showed major peaks representing triglyceride functional groups which could be observed around 2925 cm⁻¹ [C–H stretching (asymmetry)], 2854 cm⁻¹ [C–H stretching (symmetry)], 1747 cm⁻¹ [C=O stretching], 1463 cm⁻¹ [C–H bending (scissoring)], 1238, 1163, 1118 and 1097 cm⁻¹ [C–O stretching] and 722 cm⁻¹ [C–H bending (rocking)]. A peak around 1653 cm⁻¹ is attributed to C=C stretching (*cis*). The spectral region (1300-1000 cm⁻¹) which contains FTIR fingerprints of the used oils was found to be very useful in detecting extra virgin olive oil adulteration.

A band shift at 3005 cm⁻¹; assigned to C–H stretching vibration of *cis*- double bond (=C–H) characteristic to extra virgin olive oil, was observed at higher wavenumbers with increasing adulterant concentration which allowed the determination of adulteration of extra virgin olive oil.

The absorption intensity values of the spectral bands at 1163 cm⁻¹ (assigned to C–O stretching vibration and CH₂ bending vibration) increased with increasing adulterant concentration.

There was a pronounced shift of the peak at 912 cm⁻¹ (assigned to –HC=CH– of *cis*- double bond, bending out-of-plane) for extra virgin olive oil to higher wave numbers with increasing adulterant concentration.

Absorbance ratios (R1118/1097 and R1747/2925 cm⁻¹) decreased with increasing the concentrations of added adulterant oils (sunflower, corn and refined olive oils) with a fairly good linear relationship.

The spectral region selected between 1800-900 cm⁻¹ mostly represented the combination of C–H bending, C=O stretching and C=C stretching and hence it was directly related to the unsaturated C=C bond. It played a very important role in the discriminant analysis.

In conclusion, FTIR spectroscopy proved its potency to detect extra virgin olive oil adulteration at 5% level of adulterant oils (sunflower, corn and refined olive oils) which is much lower than the limit at which there exists a threatening of adulteration of extra virgin olive oil.

KEY WORDS: Adulteration; extra virgin olive oil; FTIR spectroscopy; UV spectroscopy; GC analysis; sunflower oil; corn oil; refined olive oil.

ACKNOWLEDGMENT

In the beginning; I would like to express my deep gratefulness, thankfulness and indebtedness to Allah who enabled me to accomplish this research work to come to light in its final form.

I wish to acknowledge the prior and continuing efforts of my teachers and colleagues, especially the major contributions of the Principal Supervisor; **Prof. Dr. Magda H. Allam**; Prof. Emeritus of Food Science and Technology, Faculty of Agriculture, Ain Shams University, for her great help in a thoroughly professional approach, sincere guidance, valuable comments, scientific supports and continuous supervision. Her constant interest and input have had a significant impact on the final structure of this work to be presented in an excellent form.

Acknowledgement is extended to **Prof. Dr. Atef A. Kotb**; Prof. of Food Science and Technology, Faculty of Agriculture, Ain Shams University and **Prof. Dr. Ahmad A. El-Sharkawy**; Head of Research Emeritus of Oils and Fats Technology, Food Technology Research Institute, Agriculture Research Center, for their precious technical recommendations concerning the experimental work.

I am highly thankful to all the staff members in Food Technology Research Institute, Agriculture Research Center for providing the necessary laboratory facilities, instrumentation availability and valuable contribution to carry out the experimental work.

Finally, I wish to express my sincere gratitude and appreciation to my family for their persistent encouragement, support and sympathy.

CONTENTS

Title	Page
LIST OF TABLES	iv
LIST OF FIGURES	vi
LIST OF ABBREVIATIONS	vii
1. INTRODUCTION	1
2. REVIEW OF LITERATURE	7
2.1 -Physicochemical Quality Parameters of Virgin Olive Oil	7
2.2 -Fatty Acid Composition of Virgin Olive Oil	8
2.3 -Unsaponifiable Matter Composition of Virgin Olive Oil	10
2.4 -Detection of Adulteration of Virgin Olive Oil by DSC	11
2.5 -Detection of Adulteration of Virgin Olive Oil by	14
Hyphenated Chromatographic Separation and	
Determination Techniques	
2.6 -Detection of Adulteration of Virgin Olive Oil by UV	28
Spectroscopy	
2.7 -Detection of Adulteration of Virgin Olive Oil by IR	28
Spectroscopy	
2.8 -Detection of Adulteration of Virgin Olive Oil by NMR	42
Spectroscopy	
2.9 -Detection of Adulteration of Virgin Olive Oil by Mass	46
Spectroscopy	
2.10-Detection of Adulteration of Virgin Olive Oil by CL	48
Spectroscopy	
2.11-Detection of Adulteration of Virgin Olive Oil by	50
Fluorescence Spectroscopy	
2.12-Detection of Adulteration of Virgin Olive Oil by XRF	52
Spectroscopy	
2.13-Detection of Adulteration of Virgin Olive Oil by	54
Electronic Nose	

3. MATERIALS AND METHODS	56
3.1- MATERIALS	56
3.1.1- Oils	56
3.1.1.1- Extra Virgin Olive Oil (EVOO)	56
3.1.1.2- Refined Olive Oil (ROO)	56
3.1.1.3- Sunflower Oil (SO)	56
3.1.1.4- Corn Oil (CO)	56
3.1.2- Solvents	56
3.1.3- Standards	56
3.2- METHODS	57
3.2.1- Preparation of Binary Admixtures Containing Different	57
Adulterant Oils; (w/w %)	
3.2.1.1- Binary admixtures of SO in EVOO	57
3.2.1.2- Binary admixtures of CO in EVOO	57
3.2.1.3- Binary admixtures of ROO in EVOO	57
3.2.2- ANALYTICAL METHODS	58
3.2.2.1- Refractive Index (RI)	58
3.2.2.2- Acid Value (AV) and Free Acidity	58
3.2.2.3- Peroxide Value (PV)	58
3.2.2.4- Iodine Value (IV)	58
3.2.3- Ultraviolet (UV) Spectroscopic Characteristics	58
3.2.4- Gas Chromatography (GC) Analysis	59
3.2.4.1- Gas Chromatography Analysis of Fatty Acids	59
3.2.4.2- Useful Relationships Obtained from Fatty Acid	60
Composition	
3.2.4.3- Determination of Total Unsaponifiable Matter	60
3.2.5- FTIR Spectroscopic Characteristics	61
3.2.5.1- FTIR Spectral Data Analysis	61
3.2.5.2- Useful Relationships Obtained from FTIR Spectral	62
Data Analysis	

4. RESULTS AND DISCUSSION		
4.1- Physical and Chemical Properties		
4.1.1- Refractive Index	63	
4.1.2- Acid Value and Free Acidity	67	
4.1.3- Peroxide Value	72	
4.1.4- Iodine Value	75	
4.2- Ultraviolet (UV) Spectroscopic Characteristics	80	
4.3- Gas Chromatography (GC) Analysis	89	
4.3.1- GC Analysis of Fatty Acids	89	
4.3.1.1- Detection of adulteration of EVOO with	91	
adulterant SO using fatty acids composition		
4.3.1.2- Detection of adulteration of EVOO with	92	
adulterant CO using fatty acids composition		
4.3.1.3- Detection of adulteration of EVOO with	95	
adulterant ROO using fatty acids composition		
4.3.2- Total Unsaponifiable Matter	97	
4.4- Fourier Transform Infra Red Spectroscopic Characteristics	99	
4.4.1- FTIR Spectral Data Analysis	99	
4.4.2- Detection of adulteration of virgin olive oil by FTIR	100	
Spectroscopy		
4.4.3- Detection of adulteration of EVOO with adulterant SO	115	
using FTIR spectral data analysis		
4.4.4- Detection of adulteration of EVOO with adulterant CO	126	
using FTIR spectral data analysis		
4.4.5- Detection of adulteration of EVOO with adulterant	142	
ROO using FTIR spectral data analysis		
4.4.6- Detection of adulteration of EVOO with adulterant oils	144	
(SO, CO and ROO) using FTIR spectral data analysis		
and useful relationships		
5-SUMMARY	156	
6-REFERENCES		
ARABIC SUMMARY		

LIST OF TABLES

$N^{\underline{o}}$	Title	Page
1	Refractive index values of EVOO in binary admixtures with different adulterant oils (SO, CO and ROO) at various concentrations, at 20°C	64
2	Acid values of EVOO in binary admixtures with different adulterant oils (SO, CO and ROO) at various concentrations	68
3	Free acidity values of EVOO in binary admixtures with different adulterant oils (SO, CO and ROO) at various concentrations	69
4	Peroxide values of EVOO in binary admixtures with different adulterant oils (SO, CO and ROO) at various concentrations	73
5	Iodine values of EVOO in binary admixtures with different adulterant oils (SO, CO and ROO) at various concentrations	76
6	Conjugated diene absorbency (at 232 nm) of EVOO in binary admixtures with different adulterant oils (SO, CO and ROO) at various concentrations	83
7	Conjugated triene absorbency (at 270 nm) of EVOO in binary admixtures with different adulterant oils (SO, CO and ROO) at various concentrations	84
8	Determination of ΔK of EVOO in binary admixtures with adulterant SO at various concentrations	85
9	Determination of ΔK of EVOO in binary admixtures with adulterant CO at various concentrations	86
10	Determination of ΔK of EVOO in binary admixtures with adulterant ROO at various concentrations	87

11	Fatty acids composition percent of EVOO in binary	90				
	admixtures with adulterant SO at various concentrations					
12	Fatty acids composition percent of EVOO in binary	93				
	admixtures with adulterant CO at various concentrations					
13	Fatty acids composition percent of EVOO in binary	96				
	admixtures with adulterant ROO at various concentrations					
14	Total unsaponifiable matter composition percents of EVOO	98				
	in binary admixtures with different adulterant oils (SO, CO					
	and ROO) at various concentrations					
15	Comparison of FTIR spectral data of pure oils (EVOO, SO,	111				
	CO and ROO)					
16	Absorbance ratio values (R1118/1097 cm ⁻¹) of EVOO in	146				
	binary admixtures with different adulterant oils (SO, CO					
	and ROO) at various concentrations					
17	Absorbance ratio values (R1747/2925 cm ⁻¹) of EVOO in	148				
	binary admixtures with different adulterant oils (SO, CO					
	and ROO) at various concentrations					
18	Absorption intensity values (at 1163.83 cm ⁻¹) of EVOO in	150				
	binary admixtures with different adulterant oils (SO, CO					
	and ROO) at various concentrations					
19	Shift of absorption peak (at 912.78 cm ⁻¹) of EVOO in	152				
	binary admixtures with different adulterant oils (SO, CO					
	and ROO) at various concentrations					
20	Shift of absorption peak (at 3005.52 cm ⁻¹) of EVOO in	154				
	binary admixtures with different adulterant oils (SO, CO					
	and ROO) at various concentrations					

LIST OF FIGURES

$N^{\underline{o}}$	Title	Page
1	Typical FTIR Absorption spectra (4000-400 cm ⁻¹) of pure	101
2	oils (SO, CO, EVOO and ROO)	104
2	Typical FTIR transmittance spectra of pure EVOO (top)	104
	mixed with different ratios (0, 5, 10, 20, 30, 40, 50 and	
	downwards till 100%) of SO (bottom)	
3	Typical FTIR transmittance spectra of pure EVOO (top)	105
	mixed with different ratios (0, 5, 10, 20, 30, 40, 50 and	
	downwards till 100%) of CO (bottom)	
4	Typical FTIR transmittance Spectra of pure EVOO (top)	106
	mixed with different ratios (0, 5, 10, 20, 30, 40, 50 and	
	downwards till 100%) of ROO (bottom)	
5	Typical FTIR Absorption Spectra of EVOO	107
6	Typical FTIR Absorption Spectra of 100% SO	108
7	Typical FTIR Absorption Spectra of 100% CO	109
8	Typical FTIR Absorption Spectra of 100% ROO	110
9	Typical FTIR Absorption Spectra of 5% SO	116
10	Typical FTIR Absorption Spectra of 10% SO	117
11	Typical FTIR Absorption Spectra of 20% SO	118
12	Typical FTIR Absorption Spectra of 30% SO	119
13	Typical FTIR Absorption Spectra of 40% SO	120
14	Typical FTIR Absorption Spectra of 50% SO	121
15	Typical FTIR Absorption Spectra of 5% CO	127
16	Typical FTIR Absorption Spectra of 10% CO	128
17	Typical FTIR Absorption Spectra of 20% CO	129
18	Typical FTIR Absorption Spectra of 30% CO	130
19	Typical FTIR Absorption Spectra of 40% CO	131
20	Typical FTIR Absorption Spectra of 50% CO	132

21	Typical FTIR Absorption Spectra of 5% ROO	136
22	Typical FTIR Absorption Spectra of 10% ROO	137
23	Typical FTIR Absorption Spectra of 20% ROO	138
24	Typical FTIR Absorption Spectra of 30% ROO	139
25	Typical FTIR Absorption Spectra of 40% ROO	140
26	Typical FTIR Absorption Spectra of 50% ROO	141
27	The relationship between changes in Absorbance Ratio	147
	(R1118/1097 cm ⁻¹) and concentrations of:	
	sunflower oil (a), corn oil (b), and refined olive oil (c)	
28	The relationship between changes in Absorbance Ratio	149
	(R1747/2925 cm ⁻¹) and concentrations of:	
	sunflower oil (a), corn oil (b), and refined olive oil (c)	
29	The relationship between changes in Absorption Intensity	151
	values (at 1163.83 cm ⁻¹) and concentrations of:	
	sunflower oil (a), corn oil (b), and refined olive oil (c)	
30	The relationship between Shift of Absorption Peak	153
	(at 912.78 cm ⁻¹) and concentrations of:	
	sunflower oil (a), corn oil (b), and refined olive oil (c)	
31	The relationship between Shift of Absorption Peak	155
	(at 3005.52 cm ⁻¹) and concentrations of:	
	sunflower oil (a), corn oil (b), and refined olive oil (c)	

LIST OF ABBREVIATIONS

APCI Atmospheric Pressure Chemical Ionization
APPI Atmospheric Pressure Photospray Ionization

ATR Attenuated Total Reflectance

AV Acid Value

CEC Capillary Electrochromatography

CL Chemiluminescence

CLA Conjugated Linoleic Acids

CO Corn Oil

CVA Canonical Variate Analysis

DEPT Distortionless Enhancement by Polarization Transfer

DSC Differential Scanning Calorimetry
DTGS Deuterated Tri-Glycine Sulfate
ECN Equivalent Carbon Number

ED Energy-Dispersive

EEFM Excitation–Emission Fluorescence Matrices

EEFS Excitation–Emission Fluorescence Spectroscopy

ESI Electrospray Ionization EVOO Extra Virgin Olive Oil

FA Fatty Acid

FAME Fatty Acid Methyl Ester

FFA Free Fatty Acid

FID Flame Ionization Detector

FIR Far-IR

FTIR Fourier Transform Infra Red

GC Gas Chromatography

GC-FID Gas Chromatography – Flame Ionization Detector

GC-MS Gas Chromatography–Mass Spectrometry

GILS Genetic Inverse Least Squares
GLC Gas Liquid Chromatography

HPLC High-Performance Liquid Chromatography

HPLC- High-Performance Liquid Chromatography combined APCI- with Atmospheric Pressure Chemical Ionization coupled

MS with Mass Spectrometry

HT-GC High Temperature – Gas Chromatography

ID Internal Diameter

IOOC International Olive Oil Council

IV Iodine Value

LC-MS Liquid Chromatography with Mass Spectrometry

LDA Linear Discriminant Analysis

MIR Mid-infrared

MRI Magnetic Resonance Imaging

MS Mass Spectroscopy

MSC Multiplicative Signal Correction
MUFA Mono-unsaturated Fatty Acid

NIR Near-infrared

NMR Nuclear Magnetic Resonance

OPO Olive-Pomace Oil

OSC Orthogonal Signal Correction
PCA Principal Component Analysis
PCR Principal Component Regression
PDO Protected Denomination of Origin

PLS Partial Least Square

PLS-DA Partial Least Square-Discriminant Analysis

PR Pattern Recognition

PRESS Prediction Residual Error Sum of Squares

PUFA Poly-unsaturated Fatty Acid

PV Peroxide Value QC Quality Control

QqTOF Quadrupole Time-Of-Flight

R Absorbance Ratio ROO Refined Olive Oil