بسم الله الرحمن الرحيم

Acknowledgement

I would like to express my deepest gratitude to *Prof. Dr. Ahmed Samy Khalifa*, Professor of Pediatrics, Ain Shams University, who gave me the honour of working under his supervision and who was kind enough to offer me much of his valuable time and sincere advices. His constant encouragement, close supervision and constructive guidance were the cornerstones in the selection, initiation and progress of this work.

My utmost thanks and appreciation are due to *Prof. Dr. Abdel-Baset M.S.*, Professor of Biophysics, National Research Center, for his generous support, valuable remarks and active skillfull cooperation throughout the practical part of this work.

I am deeply indebted to *Dr. Azza Abdel-Gawad*, Lecturer of Pediatrics, Ain Shams University for her great help and meticulous supervision of this work.

Last, but by no means least, I would like to thank the patients and the staff of the Hematology Oncology Unit, Children's Hospital, Ain Shams University, for without their understanding and cooperation this work would have never been accomplished.

1.



LIST OF CONTENTS

	Page
List of Abbreviations	i
List of Figures	ii
List of Tables	iii
Introduction and Aim of the Work	1
Review of Literature	
Structure of Hemoglobin	4
Normal Hemoglobin and its Variants	9
Chemically modified forms of hemoglobin	15
Hemoglobin Function and Structure Relationship	18
Absorption Spectrum of hemoglobin	21
The Thalassemias	
Definition	23
Geographic distribution	23
β-thalassemia	
Molecular pathology	26
Pathophysiology	30
Clinical picture	35
Diagnosis	44
Management	49
Other types of thalassemia	75
Subjects and Methods	80
Results	90
Discussion	133
Summary and Conclusion	160
Recommendations	165
References	167
Arabic Summary	

LIST OF ABBREVIATIONS

A⁰ : Angstrom unit AG : Adenine-Gaunine

Anti HBc : Hepatitis B core antibody
Anti HBs : Hepatitis B surface antibody

Baso : Basophils

Chel : Chelating agent
CONTROL : Control group
D.C. : Direct Current
Desfer. : Desferoxamine
Diff. : Differential
dl : deciliter
EOS : Eosinophils

fl : femtoliter
g : gram
gm : gram

GT : Guanine-Thymidine

Hb : Hemoglobin

HBsAg : Hepatitis B surface Antigen

Hct : Hematocrit

HETERO : Heterozygous group

HOMO-NONSPL: Homozygous nonsplenectomized group

HOMO-SPL: Homozygous splenectomized group
L3 and L6: Interleukin 3 and interleukin 6

I.V.S. : Intervening sequence

L : Liter

Lymph : Lymphocytes mA : milliampere

MCH : Mean corpuscular hemoglobin

MCHC: Mean corpuscular hemoglobin concentration

MCV : Mean corpuscular volume

Met-Hb : Methemoglobin

mM: millimol : Monocytes Mono Ν : Normal : nanometer nm No. : Number : picogram

Poly : Polymorphonuclear leucocytes

: Patients Pts

pg

RBCs : Red blood cells

: Voltage V : volume ٧

: White blood cells WBCs

LIST OF FIGURES

Fig. No.	Title	Page
1	Geographic Distribution of Thalassemia	24
II	The location of various classes of point mutations	27
	that cause β-thalassemia	
1	Hemoglobin Electrophoresis of the Studied Groups	93
2	Heme Concentration	102
3	Hemoglobin Conductivity	104
4	Methemoglobin Concentration	105
5-12	Correlation between Hemin Concentration and	107-114
	different studied parameters in Homozygous	
	Nonsplenectomized Thalassemia Group	
13-20	Correlation between Hemin Concentration and	115-122
	different studied parameters in Homozygous	
	Splenectomized Thalassemia Group	
21-25	Correlation between Hemin Concentration and	123-127
	different studied parameters in HETERO	
	Thalassemia Group	
26-27	Correlation between Hemoglobin Conductivity and	128-219
	Hemoglobin A ₂ concentration in HETERO Group	
28-30	Absorption Spectra of Isolated Heme in the studied	130-132
	groups	

LIST OF TABLES

Гаb. No.	Title	Page
1	Clinical Data of the Studied Groups	91
2	Hemoglobin Electrophoresis of the Studied Groups	92
3	Hemogram of the Studied Groups	95
4	Hemogram of the Studied Groups (Cont.)	96
5	Heme Concentration, Conductivity of Hemoglobin	101
	and Methemoglobin Concentration of the Studied	
	groups	
6	Correlation coefficient of the studied scatteroplots	129'

Introduction and Aim of The Work

INTRODUCTION AND

AIM OF THE WORK

The thalassemia syndromes constitute the most prevalent of all known genetic diseases. About 3% of world's population carry β-thalassemia genes (Lukens, 1993).

Each year, 100,000 children throughout the world are born with thalassemia major (Esposito, 1992).

In Egypt, β-thalassemia is the commonest chronic hemolytic anemia (Sabry, 1973).

Along the last 15 years, thalassemia represented 40% of the hematological problems in children attending The Hematology/Oncology Clinic, Children's Hospital, Ain Shams University (*Imam*, 1994).

To date, well in excess of 100 different thalassemia mutations have been identified (*Huisman*, 1992).

Although the genetic defects of β -thalassemia, as well as the molecular pathology of these defects have been studied intensively, the reasons leading to premature removal of abnormal RBCs in the marrow

and from peripheral circulation are not clearly understood (Shinar et al., 1987).

The extreme clinical heterogeneity of thalassemia phenotypes reflects an enormous diversity of genetic mutations. Some mutations interact to produce a severe thalassemia phenotype, whereas others result in relatively mild syndromes (McDonagh and Nienhuis, 1993).

The basic pathology in thalassemia is a decrease in either α or β chain synthesis which has several deleterious effects on red cell production and survival. Selective deficiency of one or more polypeptide chains has two immediate consequences: decreased hemoglobin synthesis and imbalance between α and non- α chain production. The absence of complementary globin chains with which to bind, chains whose synthesis is normal form aggregates, precipitate within the cytoplasm, damage cell membranes, and lead to premature cell destruction (Lukens, 1993).

The end product of the precipitated hemoglobin chains is heme, from which eventually iron and globin are liberated. Globin chains have been found to interact and disrupt the RBC membrane, leading to damage of cytoskeleton. Excess of iron is known to be a catalyst of peroxidation, causing damage to the various RBC membrane components. The role of heme has not yet been studied in detail in thalassemic RBCs. However,

there is some evidence that heme participates in damaging RBCs in other types of hemoglobinopathies (Shinar and Rachmitewitz, 1990).

Free heme is capable of interacting with either the lipid bilayer or cytoskeletal membrane protein, however, the direct contribution of the heme molecule to the creation of oxidative damage in thalassemia still needs to be elucidated (Wolfe, 1989).

Massive splenomegaly is a characteristic feature of the natural course of severe thalassemia. Splenectomy is often necessary in the management of a patient with severe β -thalassemia. Splenectomy offers the long lasting benefit of reduction of transfusion requirement together with improving the quality of life of β -thalassemia patient (*Pinna et al.*, 1988).

The objective of this study was to evaluate the possible role of heme in the pathophysiology of β -thalassemia through studying the conformational state of heme, heme concentration, conductivity of hemoglobin and methemoglobin concentration in β -thalassemia patients, and comparing these characteristics with those of heterozygous thalassemia individuals as well as with a normal control group. In addition, the effect of splenectomy on the heme pattern was assessed.

Review of Literature

STRUCTURE OF HEMOGLOBIN

One vital function of the red blood cell is to mediate the exchange of respiratory gases, oxygen and carbon dioxide, between the lungs and the tissues. Of fundamental importance to this process is the oxygen-transport red-pigmented protein, hemoglobin. It is the major constituent of the red cell cytoplasm, accounting for about 90% of the dry weight of the mature cell (Telen, 1993).

Hemoglobin is a tetramer with a molecular weight of 64,400 daltons, it consists of two pairs of unlike globin polypeptide chains. An iron-containing porphyrin derivative called heme, a ferroprotoporphyrin IX, is linked covalently at a specific site to each chain. When heme iron is in the reduced (ferrous) state, it can bind reversibly with gaseous ligands, such as, oxygen and carbon monoxide. In developing human erythroblasts, eight genes direct the synthesis of six structurally different globin polypeptide chains designated by a Greek letter α , β , γ , δ , ε , and ζ (Bunn, 1993).

GLOBIN

The globin polypeptide chains in hemoglobin differ from one another in amino acid sequence. The α -chain contains 141 amino acids, whereas β -chain (as well as γ , δ , and ϵ) have 146. The δ -hain differs from

the β -chain in only 10 of the 146 amino acid residues, whereas the γ - and β -chains differ by 39 amino acids (Bunn and Forget, 1986).

Protein structure is routinely described using four different aspects of structure: 1) Primary structure, or the linear sequence of amino acids; 2) Secondary structure, which describes how the amino acids within segments of the protein are spacially organized, e.g., by folding into an alpha helix or beta pleated sheet; 3) Tertiary structure, which refers to the steric relationships of sequence domains that are separate from each other when analysed as part of the linear sequence of the protein; and 4) Quaternary structure, or the way in which several polypeptide chains join to form a single molecule (Bunn and Forget, 1986).

The primary structure of the globin chains are different from each other, however, their secondary structures are remarkably similar. Each has eight helical segments designated by the letters A through H. The helices are of nearly identical length in all four normal chains except for the D helix, which contains seven amino acids in the β -, γ -, and δ -chains, but only two amino acids in the β -chain. The helices make up about 75% of the molecule. Interspersed between them are seven nonhelical segments. This arrangement is important structurally, because the helices are relatively rigid and linear, whereas the nonhelical segments allow bending (*Perutz*, 1987).