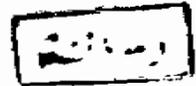


MEASUREMENT OF THE FREE ERYTHROCYTE
PROTOPORPHYRIN IN CHILDREN WITH
MICROCYTOSIS

THESIS

Submitted to the Graduate Division
in Partial Fulfilment of the
Requirement for the Degree of
MASTER OF SCIENCE
IN
Clinical Pathology



BY

NANCY LEWIS WASSEF

SUPERVISORS

Professor A.S. KHALIFA
Professor Of Pediatrics

Dr. FADILA H. SABRY
Assistant Professor of Clinical Pathology

Handwritten signatures and initials, including a large signature and the initials 'CS'.

FACULTY OF MEDICINE
AIN SHAMS UNIVERSITY



1983

IN THE NAME OF GOD



**TO MY PARENTS AND MY DAUGHTER,
FOR THEIR CONTINUOUS ENCOURAGEMENT AND LOVE**

Aknowledgement

ACKNOWLEDGEMENT

I wish to express my appreciation and gratitude to **Professor A.S. Khalifa**, Professor of pediatrics, to whom I am deeply indebted for his helpful supervision.

Gratefully, I thank **Dr. Fadila H. Sabry**, Assistant Professor of Clinical pathology, for the helpful and cogent advices. Her valuable help in providing me with a flow of essential informations together with useful references calling my attention to an error, and generously encouraging me was particularly important to achieve this work.

Finally, I appreciate the help and encouragement from all my colleagues in the Blood Transfusion Centre, Clinical Pathology and Paediatric Departments.

TABLE OF CONTENTS

	<u>Page</u>
INTRODUCTION AND AIM OF WORK.....	1
REVIEW OF LITERATURE.....	
- 1- <u>Haemoglobin</u> :.....	4
- Molecular Structure of Haemoglobin.....	4
- Haemoglobin Biosynthesis.....	8
- 2- <u>Iron Metabolism</u> :.....	18
- Iron Compartments.....	18
- Components of Internal Iron Exchange....	28
- Iron Deficiency.....	29
- 3- <u>Beta Thalassaemia Trait</u> :.....	31
- Historical Perspectives.....	31
- Heterozygous B-thalassaemia.....	32
- Molecular Basis.....	33
- Pathophysiology.....	34
- 4- <u>Erythrocyte Protoporphyrin</u> :.....	40
- Historical perspectives.....	40
- Synthesis of Protoporphyrin.....	41
- Significance of E.P.....	44
MATERIAL AND METHODS.....	49
Statistical Analysis.....	61
RESULTS.....	64
DISCUSSION AND CONCLUSION.....	78
SUMMARY.....	83
REFERENCES.....	85
ARABIC SUMMARY.....	

INTRODUCTION & AIM OF WORK

INTRODUCTION AND AIM OF WORK

Thalassaemia syndromes and iron deficiency are considered to be the commonest haematologic disorders that pose major public health problems in many parts of the world, particularly in the underprivileged communities. Reports throughout the world support the high prevalence of iron deficiency anaemia in young children, especially in the developing countries (Bothwell et al., 1979); meanwhile, heterozygous B-thalassaemia is considered as the most common type of the thalassaemias (Pierce et al., 1977). In Egypt, Sabry et al. (1973) found that its incidence was 1.3%.

B-thalassaemia trait mimicks iron deficiency in the presence of both microcytosis and suppressed haemoglobin synthesis (Bainton Finch, 1964; Nathan, 1972; and Ward, 1979), and it is usually not possible to make a distinction between both disorders on examination of the blood smear.

For identification of iron deficiency, it was found that determination of the plasma ferritin concentration, which closely parallels body iron reserves, is the most

valuable criterion (Jacobs et al., 1972; Lipschitz, et al., 1974) on the other hand, globin synthetic rate measurement provides the most definitive method to diagnose heterozygous B-thalassaemia (Beris et al., 1980 and Congote, 1981). But since these procedures of diagnosis are not suitable for general use in the routine clinical laboratory, a simple, accurate and precise method is needed to differentiate between iron deficiency as a cause of anaemia and B-thalassaemia trait.

Dagg, Goldberg and Lockhead (1966) found that the estimation of free erythrocyte protoporphyrin (FEP), together with percentage saturation of the total iron binding capacity, allowed an accurate diagnosis of latent iron deficiency as supported by Langer et al., (1972) and Van Eijk et al. (1974).

Meanwhile, Pearson, O'Brien and McIntosh (1973) have demonstrated that the electronic measurement of mean corpuscular volume (MCV) was a reliable screening test for thalassaemia trait among adults; but this was not suitable procedure in children due to the age related changes occurring in MCV during childhood, as

described by Berman et al. (1980), and because of the incidence of microcytosis caused by iron deficiency.

AIM OF WORK:

The aim of this work is to measure the free erythrocyte protoporphyrin (FEP) simultaneously with the determination of the mean corpuscular volume (MCV) in children with microcytosis, since heterozygous B-thalassaemia and iron deficiency anaemia are known to be common haematological disorders among children in the region.

An attempt to evaluate the usefulness of such test in the differential diagnosis between both conditions will be the matter of this study.

Review of Literature

1- HAEMOGLOBIN

Molecular Structure of Haemoglobin:

Haemoglobin is one of the most important biological constituents of the body. It has been known for over a century that human haemoglobin is a heterogenous intra-erythrocytic respiratory protein of a molecular weight 64 000 daltons, and it is thought to be ellipsoid in shape with a central cavity (Ingram et al., 1956).

The basic structure of haemoglobin consists of two alpha and two non alpha polypeptide chains of globin, colourless protein, on the surface of which are attached four small prosthetic groups of haem molecules that impart to haemoglobin its red colour and its unique oxygen carrying properties (Granick, 1949) (Figure 1).

GLOBIN

In normal adult haemoglobin (A), there are two alpha-chains made up of 141 amino-acids each, and two Beta-chains each of which are 146 amino-acids long (Rhine-Smith et al., 1957).

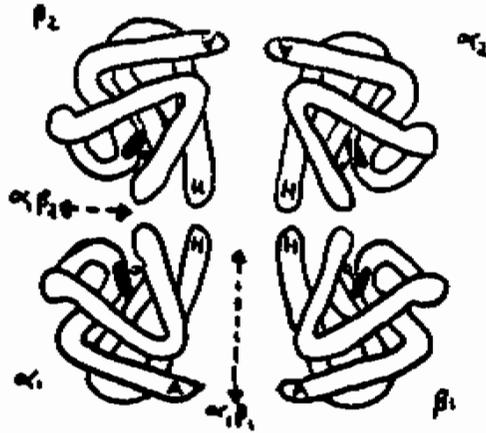


Figure (1): The arrangement of the 4 polypeptide chains of haemoglobin.

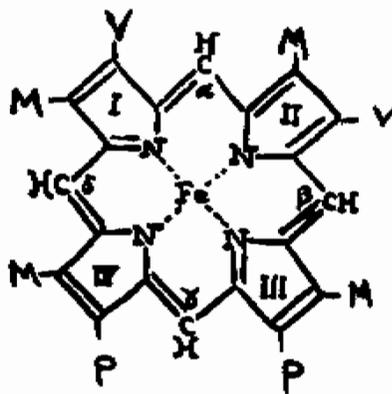


Figure (2): Fe protoporphyrin.

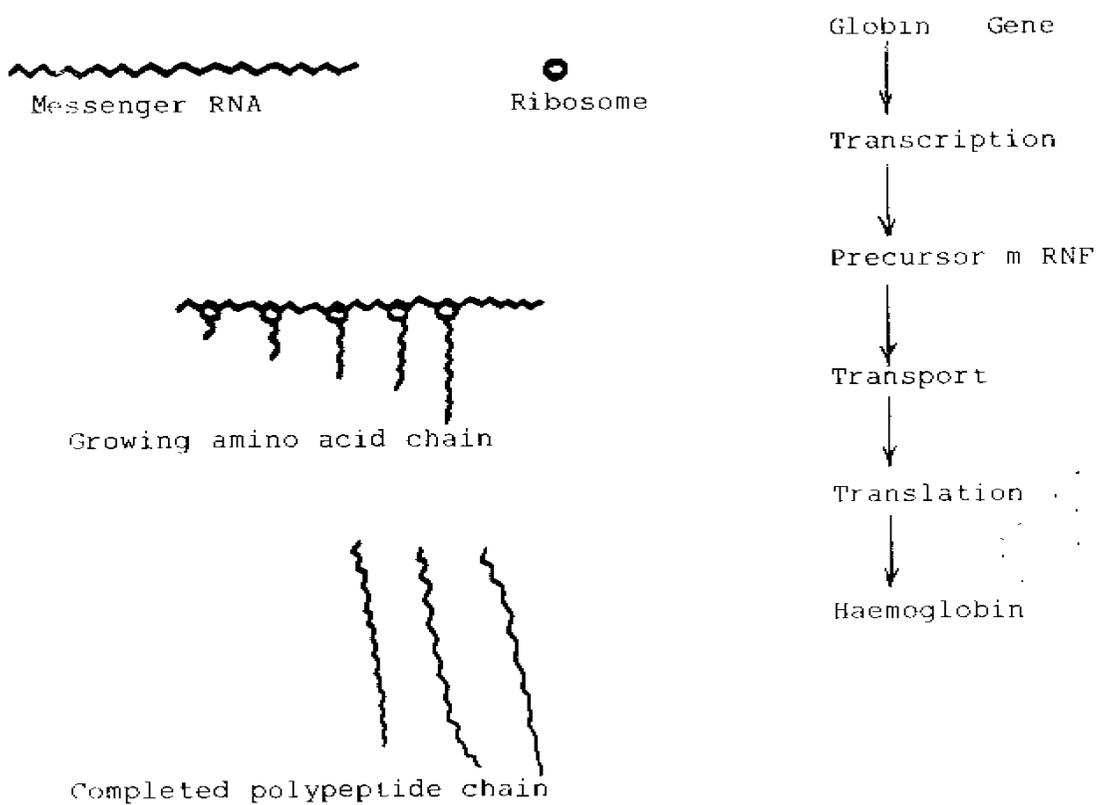


Figure (3): Messenger RNA carries the coding for the sequence of amino acids required to build globin. A ribosome moves to mRNA, and the appropriate a.a. on its specific transfer RNA is brought. Further a.a. are joined on to form the polypeptide globin chain.