

Cairo University
Faculty of Veterinary Medicine
Department of Medicine and Infectious Diseases

# Title of Thesis Epidemiological and immunological studies on some viral equine infections causing reproductive problems in Egypt

Thesis presented by

Samer Fouad Said Khalil

(Master degree of Infectious Diseases, Cairo University, 2012)

For the degree of

Ph.D. of V.Sc. (Infectious Diseases)

**Under Supervision of** 

Prof.Dr. Raouf R. Youssef

Professor of Infectious Diseases
Department of Medicine and Infectious
Diseases
Faculty of Veterinary Medicine
Cairo University

Prof.Dr. Fayez A. Salib

Prof. Dr. Hany M. Hassan

Professor of Infectious Diseases
Department of Medicine and Infectious
Diseases
Faculty of Veterinary Medicine
Cairo University

Chief researcher Immunology Diseases and Toxicology Animal Reproduction Research Institute ,Haram, Giza

Prof. Dr. Magda A. Kalad

Chief researcher
Department of Equine Viral Diseases
Veterinary Serum and vaccine Research
Institute (VSVRI), Abbasia, Cairo.

Cairo University
Faculty of Veterinary Medicine
Department of Medicine and Infectious Diseases



## **Supervision Sheet**

## **Supervisoes**

#### Prof.Dr. Raouf R. Youssef

Professor of Infectious Diseases Department of Medicine and Infectious Diseases Faculty of Veterinary Medicine Cairo University

#### Prof. Dr. . Fayez Awad Salib

Professor of Infectious Diseases Department of Medicine and Infectious Diseases Faculty of Veterinary Medicine Cairo University

#### Prof. Dr. Hany Mohamed Hassan

Chief researcher Immunology Diseases and Toxicology Animal Reproduction Research Institute ,Haram, Giza.

#### Prof. Dr. Magda Anis Kalad

Chief researcher Department of Equine Viral Diseases Veterinary Serum and vaccine Research Institute (VSVRI), Abbasia, Cairo. Cairo University Faculty of Vet. Medicine

Name: Samer Fouad Said Khalil Date of Birth: 19-6-1979

**Degree: Ph.D.** in Veterinary Sciences **Specialization:** Infectious Diseases

Title of Thesis: Epidemiological and immunological studies on some viral equine

**Dept.:** Medicine and Infectious Diseases

Nationality: Egyptian

Place of Birth: Cairo, Egypt

infections causing reproductive problems in Egypt

Supervisors:

**Prof. Dr. Raouf R. Youssef:** Professor of Infectious Diseases, Department of Medicine and Infectious Diseases, Faculty of Veterinary Medicine, Cairo University

**Prof. Dr. Fayez A. Salib:** Professor of Infectious Diseases, Department of Medicine and Infectious Diseases

, Faculty of Veterinary Medicine, Cairo University

**Prof. Dr. Hany M. Hassan:** Chief researcher, Immunology Diseases and Toxicology, Animal Reproduction Research Institute, Haram, Giza

**Prof. Dr. Magda A. Kalad:** Chief researcher, Department of Equine Viral Diseases Ve terinary Serum and vaccine Research Institute (VSVRI), Abbasia, Cairo.

#### **Abstract**

Studying the prevalence of Equine herpes virus-1(EHV-1), Equine herpes virus-4(EHV-4) and Equine Viral Arthritis (EVA) infection among equines in different governorates of Egypt. Evaluating the indirect Enzyme-Linked Immuno-Sorbent Assay (ELISA) for the diagnosis of them in comparison with the virus isolation. Assessing the efficacy of Polymerase Chain Reaction (PCR) for the diagnosis of the viral infection by using the horses nasopharyngeal swabs isolated virus and the aborted foeti tissues. One hundred eighty two equines of different ages, sexes, localities and breeds showing signs of fever, respiratory manifestations, abortions, ataxia, hind limb paralysis, limb edema, foal depression and death; were examined. The viral isolation on both histopathology and Baby Hamster Kidney (BHK) cell culture, the aborted foeti were histologically examined for the detection of inclusion bodies, the serum samples were collected to detect Immunoglobulin-G specific to EHV-1,EHV-4 and EVA by using ELISA, EHV-4 and EVA were negative .the horses nasopharyngeal swabs isolated virus and the aborted foeti tissues were tested by PCR using specific primers to EHV-1 to confirm the infection with EHV-1. The prevalence of EHV-1 infection in the examined animals were recorded as 4.94%, it was prevalent in Cairo, Giza, Kafr Elsheikh, Monofeia, El Sharkia governorates. The EHV-1 was isolated on the embryonated chicken eggs and the pock lesion was appeared on the chorio-allantoic membranes. The cytopathic effects were also observed on the tissue culture. The Liver of aborted foeti showing necrosis of all hepatic tissue and activation of kuffer cells with heamosidren and the detection of

i

esinophilic Intranuclear inclusion bodies. Indirect ELISA could detect IgG in all infected equines(N=9) from which EHV-1 isolated. PCR proved the infection with EHV-1 in the aborted foeti (N=3) tissues and gave the similar results by using the horses nasopharyngeal swabs isolated virus where 489 bp PCR products were detected in both. EHV-1 is prevalent in Egypt in different governourates. EHV-1 infection could diagnosed by intranuclear esinophilic inclusion bodies in the aborted foeti tissues. The indirect ELISA could diagnose EHV-1 infection in all ages and sexes groups. PCR on aborted foeti tissues is better for the diagnosis of EHV-1 infection than that on the horses nasopharyngeal swabs isolated virus because it save time and money, the study indicated that there are no EHV-4 and EVA infection found in Egypt.

Keywords: EHV-1, EHV-4, EVA, ELISA, PCR, diagnosis, Egypt.

# **DEDICATION**

I dedicate my thesis
to my family especially my parents, my
wife,
my daughters and my Son

## Acknowledgment

I wish to express my appreciation to Prof.**Dr. Raouf R. Youssef,** Professor of Infectious Diseases, Faculty of Veterinary Medicine, Cairo University, for his various attributions have led to the completion of this thesis.

It is with the deepest gratitude that I wish to thank Prof. **Dr. Fayez A. Salib,** professor of Infectious Diseases, Faculty of Veterinary Medicine, Cairo University, for his mentorship, guidance and encouragement to complete this thesis.

I would also like to extend my gratitude and appreciation to **Prof. Dr.Hany M. Hassan,** Chief researcher of Immunology Disease and Toxicology Animal Reproduction Research Institute, Haram, Giza for his support, guidance, continuous help and encouragement throughout the work.

I would also like to extend my gratitude and appreciation to **Prof. Dr. Magda A. Kalad,** Chief researcher Department of Equine viral Diseases (VSVRI), Abbassia, Cairo for indispensable support, advise and help throughout the work.

I would also like to extend my gratitude and appreciation to **Dr. Jehan A. Gafer,** Chief researcher & Head of biotechnology unit Animal Reproduction Research Institute, Haram, Giza for her help in PCR to complete this research work.

I would also like to extend my gratitude and appreciation to **Dr. Thanaa K. Hassanien**, Assistant Chief researcher of pathology Animal Reproduction Research Institute, Haram, Giza for her help in pathology and histopathology to complete this research work.

I am appreciating all stuff members of department of Animal reproduction research institute for their cooperation

To all who have assisted me in any aspect of this work, I humbly thank you

# **CONTENTS**

ITEM	PAGE
ABESTRACT	i
DEDICATION	iii
ACKNOWLEDGEMENT	iv
CONTENTS	V
LIST of TABLES	vi
LIST of PHOTOS	vii
LIST of CHARTS	ix
1.INTRODUCTION	1
2.REVIEW of LITERATURES	5
2.1.Equine herpes viruses (EHV)	5
2.1.2. Diagnosis and PCR for equine herpes viruses (1	21
and 4)	56
2.1.3.Clinical, Haematological, Pathological and Clinicopathological for	30
2.2. Equine Viral Arteritis (EVA)	60
2.2.1 Epidemiology of equine viral arteritis (EVA)	60
2.2.2. Diagnosis of equine viral arteritis(EVA)	57
3-MATERIAL AND METHODS	91
3.1. Materials	91
3.2.Methodes	98
4 – RESULTS	105
4.I.Experiment-1: Clinical and epidemiological examination	103
of horses	105
4.2. Experiment-2: Pathology and Histopathology of the	
aborted foeti	112
4.3.Experiment-3: Polymerase chain reaction for the	
diagnosis of EHV-1	119
5-DISCUSSION	123
6-Summary	131
7- REFERENCES	133
List of abbraviation	150
الملخص العربي	
المستخلص العربي	

# LIST of TABLES

No. of Table	Page
Table (1). The recorded Clinical signs of the EHV-1 infected	
equines	106
Table (2) . The number of the collected samples from the	
examined infected equines	107
Table (3) . Epidemiological data of the examined infected	
equines	108
	100
Table (4). The number of the examined animals	109
Table (5) . Virus isolation and Serology of the infected animals	110
Table (6) . Gross pathology and histopathology of the aborted foeti	
infected by EHV1	119
Table (7) Results of Indirect ELISA and PCR for diagnosis of	
EHV-1 in examined equines	120
Table (8) . Comparison of indirect ELISA on sera of the aborted	
mares and PCR on the aborted foeti	120

## **LIST of PHOTOS**

No. of Photo	Page
Photo (1): Severe conjunctivitis due to trauma resulted from nervous	111
signs	
photo(2): Hind limb paralysis	111
Photo(3): Pock lesion on chorio-allantoic membrane	112
Photo (4): Aborted equine fetus	113
Photo (5): Aborted foal showing Abnormal coloration of the eye.	113
Photo (6): Aborted foal showing congested viscera	114
Photo (7): Intestine of aborted foal showing congestion and edema.	114
Photo (8): Lung of aborted foal appeared edematous with areas were	115
rubbery in texture area with whitish coloration on the urface	
Photo (9): Liver of aborted foal appeared enlarged, friable ,dark and congested areas at the priphery of the lobules with whitish	
colouration on the surface.	115
Photo (10): Spleen of aborted foal appeared congested, morbid and	110
edematous organ	116
Photo (11): Liver of aborted foal showed degenerative changes of	
hepatocytes with focal area of inflammatory cell aggregations	
at the portal area. H&E.X100.	116
Photo (12): liver of aborted foal showing areas of hydropic	
degeneration of hepatocytes with focal areas of hepatic	
necrosis. Focal inflammatory cell aggregation.H&E.X400.	117
Photo (13): Liver of aborted foal showing necrosis of all hepatic	
tissue and activation of kuffer cells with heamosidren.	
Esinophilic Intranuclear inclusion bodies (black arrow).	117
(H&E,X400).	117
Photo (14): Lung of aborted foal showing degenerated bronchiolar	
epithelium and the alveolus lined by degenerated epithelial cells (vacuolar) and surrounded by a fibrous tissue	
proliferation .The alveolar space filled by numerous	
inflammatory cells, primarily neutrophils.H&E.X400.	118
Photo (15): Spleen of aborted foal showed congestion with depletion	110
of spleen follicles and necrosis of others. H&E. X100	
r	118
Photo(16): PCR techniques applied on the EHV-1 DNA extracted	
from aborted foeti tissues and its products showed that EHV-1	
specific band at 489 bp.	121

# LIST OF CHARTS

No. of Chart	Page
Figure (1). The recorded Clinical signs of the infected	
equines	106
Figure(2). The tested animals	109

#### 1-INTRODUCTION

There is a great interest with equine industry all over the world, especially in Egypt and Arabian countries which plays an important role in the national income. Abortion in horses may result from a variety of infectious agents, such as bacteria, viruses or fungi that attack the fetus or its membranes, resulting in fetal death and expulsion.

EHV-1 is one of group of alpha-herpes viruses. Both EHV-1 and EHV-4 have extensive antigenic cross-reactivity and were previously considered subtypes of the same virus (EHV-1). DNA fingerprinting has demonstrated both the genetic divergence of the two virus species (Radostitis, *et al.* 2007). EHV-1 is an imported pathogen and has been responsible for causing four recognized clinical syndromes in horses throughout the world: upper respiratory disease in young horses, abortions in late pregnant mares, perinatal foal mortality and occasionally neurological disorders (Reed and Toribio, 2004).

Economic losses caused by equine herpes virus -1 is summarized in abortion and loss of fetus, death of horse due to encephalomyeilitis, loss of semen of infected stallion, loss of infected newborn foals and loss of training time and opportunities to perform during convalescence and quarantine. Abortion in affected mares can occur at any time within 1-14 days after the onset of the disease. Most mares abort during the last half of gestation (5-10months). Incidence of abortion is up to 50 percent. The disease

can cause death in horses with a course of disease of 2-15 days. In some cases, the foal will be born alive at term and will die shortly after birth due to infection by the virus. The abortion rate may approach 100% in a herd of susceptible mares. The vaccines that are currently available are the best we have but are of questionable value in preventing abortion, **Amer** *et al.* (2011).

Diagnosis of EHV-1 must be rapid and sensitive so early intervention policies aimed to reduce the virus spread. Routine diagnosis of EHV-1 infection in live animals is usually achieved by virus isolation in SPF embryonated chicken egg, cultured cells from nasopharyngeal secretions or from the tissues of aborted fetus (Elia et al., 2006). Several rapid and innovative diagnostic techniques based on PCR and enzyme linked immunosorbent assay (ELISA) were applied for the diagnosis of EHV-1 infection (Ataseven et al., 2009).

Equine Herpesvirus-1 (EHV-1) abortion virus is most often associated with abortions in mares, while Equine Herpesvirus (EHV-4) rhinopneumonitis virus is usually associated with respiratory disease in young horses. Both subtypes have the potential to cause respiratory disease and abortion. Vaccines are available as aids to prevent abortion due to EHV-1 infections. The herpes family of viruses has the capacity to persist in the body of its host in a dormant state as an inapparent carrier after the primary infection. Months or years after the primary infection, the latent herpesvirus may again become manifest with renewed replication and with the potential for initiating new outbreaks of disease in its host as well as susceptible

stable mates. Therefore, it is the existence of these latently infected carrier horses, from which the virus is re-activated by stress-induced circumstances and shed into the environment to infect other individuals, that initiates a new outbreak of the disease **OIE** (2015).

Equine Viral Arteritis virus (EVA) has the potential to cause abortion as well as the more commonly observed contagious respiratory disease and semen-shedding state in infected carrier stallions. No EVA-related abortions were diagnosed at the Animal Health Laboratory, University of Guelph, from 1998 to 2004. By inhalation primarily, also transmission from infected weanlings to broodmare. Mares aborting can transmit virus from aborted fetus, placenta and fluids. This virus can also be spread by anyone handling infective material. Abortion in affected mares can occur at anytime within 1-14 days after the onset of the disease. Most mares abort during the last half of gestation (5-10 months). Incidence of abortion up to 50 percent. The disease can cause death in horses - course of disease from 2-15 days (Afify *et al.* 2013).

The importance of the equine viruses causing reproductive problems forcing us to studying them as summarized in *our* research work goals as follows:

1- Recording the epidemiological data of equine viral diseases causing reproductive problems (equine herpes viruses 1(EHV-1), equine herpes virus 4 (EHV-4) and equine viral arteritis virus (EVA).

- 2- Identifying the equine viruses causing reproductive problems by the virus isolation and characterization.
- 3- Investigating the foetal tissues alterations by Gross and histopathological examination of the aborted foeti and their foetal membranes.
- 4- Screening the infected equines and their contacts by indirect enzyme immunosorbent assay on the collected sera for EHV-1, EHV-4 and EVA.
- 5- Evaluating polymerase chain reaction on the isolated virus and the aborted foeti tissues for the diagnosis of the equine viral infections causing reproductive problems.

## 2 - REVIEW OF LITERATURES

## 2.1. EquineHerpes viruses (EHV):

#### 2.1.1 Epidemiology:

Foote, et al. (2004) indicated that a silent cycle of equine herpesvirus 1 infection has been described following epidemiological studies in unvaccinated mares and foals. In 1997, an inactivated whole virus EHV-1 and EHV-4 vaccine was released commercially in Australia and used on many stud farms. However, it was not known what effect vaccination might have on the cycle of infection of EHV-1. To investigate whether EHV-1 and EHV-4 could be detected in young foals from vaccinated mares. Nasal and blood samples were tested by PCR and ELISA after collection from 237 unvaccinated. unweaned foals and vaccinated nonvaccinated mares during the breeding season of 2000. EHV-1 and EHV-4 DNA was detected in nasal swab samples from foals as young as age 11 days. These results confirm that EHV-1 and EHV-4 circulate in vaccinated populations of mares and their unweaned, unvaccinated foals. The evidence that the cycle of EHV-1 and EHV-4 infection is continuing and that very young foals are becoming infected should assist stud farms in their management of the threat posed by these viruses.

Borchers, *et al.* (2005) collected a total of 51 sera from a migratory population of Burchell's zebras (Equusburchelli) in the Serengeti National Park (Tanzania) between 1999 and 2001 to assess