

**EVALUATION OF N-ACETYLCYSTEINE
AND VITAMIN "E" IN PROPHYLAXIS
AND TREATMENT OF NEUROTOXICITY
INDUCED BY "2,5-HEXANEDIONE"**

THESIS

SUBMITTED IN THE PARTIAL FULFILLMENT FOR
THE DEGREE OF M.D. IN
CLINICAL TOXICOLOGY

BY

GAMAL NASSER EID EL SAYED

LECTURER OF FORENSIC MEDICINE & CLINICAL TOXICOLOGY
AIN SHAMS UNIVERSITY
FACULTY OF MEDICINE

SUPERVISED BY

FROM AIN SHAMS UNIVERSITY- FACULTY OF MEDICINE
DEPARTMENT OF FORENSIC MEDICINE AND CLINICAL TOXICOLOGY

PROF. DR.
ASSEM ABD EL REHEEM EL HABASHI
THE HEAD OF THE DEPARTMENT

PROF. DR.
MOSTAFA ABD EL LATIF KAMEL
PROF. OF FORENSIC MEDICINE & TOXICOLOGY

DR. HANY A. GAMAL EL DIN
ASSIS. PROF. OF FORENSIC MEDICINE & TOXICOLOGY

FROM DUKE UNIVERSITY MEDICAL CENTER "USA"
DEPARTMENT OF PHARMACOLOGY

PROF. DR. MOHAMED B. ABOU DONIA
PROF. OF PHARMACOLOGY

1989

بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

وَمَا أُوتِيتُمْ مِنَ الْعِلْمِ إِلَّا قَلِيلًا

صَدَقَ اللَّهُ الْعَظِيمُ



TO MY PARENTS

TO MY BELOVED WIFE

TO MY SWEETHEART AHMED

AND USAMA

ACKNOWLEDGMENT

I would like to express my gratitude and thanks to Prof. Dr. Assem Abd El Reheem El Habashi, head of the Department of Forensic Medicine and Clinical Toxicology, Ain Shams University, for his supervision and unlimited helpful support to me all through the stages of this work.

My sincere appreciation and grateful thanks are due to Prof. Dr. Mostafa Abd El Latif Kamel, Prof. of Forensic Medicine and Clinical Toxicology, Ain Shams University, for his invaluable suggestion of the original detailed protocol of this work, upon which every thing was built up.

My thanks and appreciation are deserved to Assis. Prof. Hany A. Gamal El Din, Assis. Prof. of Forensic Medicine and Clinical Toxicology, Ain Shams University, for his efforts in initiation and establishing of the Scientific Channel System through which the work was carried out.

I am really indebted and grateful to Prof. Dr. M. B. Abou Donia, Prof. of Pharmacology, Duke University Medical Center, North Carolina, U.S.A., who gave me the chance and unlimited facilities to carry out my

research in his laboratory under his direct supervision.

Grateful thanks are due to Prof. Dr. D.M. Lapadula, Laboratory of Neurotoxicology, Duke University Medical Center, U.S.A, from whom I got a lot of invaluable knowledge especially the techniques of electrophoresis and immunobinding.

My sincere thanks are deserved to Prof. Dr. H.A. Tilson, National Institute of Environmental Health Sciences, N.C. U.S.A. for his supervision and guidance during performing the neurobehavioral tests.

My thanks and good wishes are to Prof. Dr. C. Clark, Prof. Dr. R. Gupta and Prof. Dr. K. Jane, Laboratory of Neurotoxicology, Duke University Medical Center, U.S.A. for their great help to me.

Great thanks are deserved to M. Green, Secretary of the Laboratory of Neurotoxicology, Duke University Medical Center, U.S.A. for her efforts in typing this thesis.

7

LIST OF ABBREVIATIONS

n-Hx	n-Hexane
2, 5-HD	2, 5-Hexanedione
2, 5-HDiol	2, 5-Hexanediol
MEK	Methyl ethyl ketone
Mn-BK	Methyl n-butyl ketone
IDPN	B, B-iminodipropionitrile (neurotoxic compound)
NAC	N-Acetylcysteine
IF(s)	Intermediate filament(s)
NF(s)	Neurofilament(s)
NFP(s)	Neurofilament protein(s)
NFL, NFM, NFH	Neurofilament, low, medium and high respectively
Mt	Microtubules
MAP(s)	Microtubule associated protein(s)
Sca	Slow component _a of axonal transport
OSHA	Occupational Safety and Health Administration
ACGIA	American Conference of Governmental Industrial Hygienists
U. S. DHEW	United States Department of Health Education and Welfare
TIV(s)	Threshold Limit Value(s)

TLV-TWA	Threshold Limit Value-Time Weighted Average
LD ₅₀	Lethal dose ₅₀
NCV	Nerve Conduction Velocity
NAP	Nerve Action Potential
MAP	Muscle Action Potential
GSH	Reduced glutathione
GSSG	Oxidized glutathione
LFB	Luxol Fast Blue
DTT	Dithioreitol (sulfhydryl donor)
GAPDH	Glyceraldehyde 3 - phosphate dehydrogenase
PFK	Phosphofructokinase
GSSG-R	Glutathione reductase
GSH-PX	Glutathione peroxidase
SOD	Superoxide dismutase
XOD	Xanthine Oxidase
SDS-PAGE	Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis
ANOVA	Analysis of Variance

* * *

CONTENTS

	Page
* INTRODUCTION.....	1
* HISTORICAL REVIEW.....	3
* ANATOMICAL PRINCIPLES	5
The Neurone	6
Neuronal Cytoskeleton.....	7
Neurofilaments.....	9
Axonal Transport.....	12
* NEUROTOXIC HEXACARBONS.....	14
- N- Hexane.....	14
- 2, 5-Hexanedione.....	15
- Methy n-butyl Ketone.....	16
* TOXICOLOGY OF NEUROTOXIC HEAXACARBONS.....	17
* PHARMACOKINETICS AND BIOTRANSFORMATION OF NEUROTOXIC HEXACARTONS.....	19
* HUMAN HEXACARBON TOXICITY	24
- Sources of Human Exposure.....	24
- Effects of Acute Human Exposure.....	25
- Effects of Subacute and Subchronic Exporure.....	25
Clinical picture.....	25
Electrophysiological studies in man.....	28
Laboratory findings.....	29
Neuropathological findings.....	29
Diagnosis	30
Defferential diagnosis.....	30

* EXPERIMENTAL TOXICOLOGY OF HEXACARBONS.....	31
Effects of Repetitive or Continuous Exposure.....	31
- General changes.....	31
- Body weight.....	31
- Behavioral changes.....	32
- Electrophysiological studies.....	32
- Functional sings.....	33
- Neuropathological findings.....	33
- Therapeutic trials.....	38
* PATHOGENESIS OF HEXACARBON NEUROPATHY	
* STRUCTURE ACTIVITY RELATINSHIFS	39
* THEORIES OF HEXACARBON NEUROPATHY	40
A - Interruption of Energy Metabolison.....	40
B - Inhibition of Sterologogenesis.....	43
C - Disruption of Neurofilaments.....	43
D - Inhibition of Normal Turnover of Neurofilaments.....	49
* N-ACETYL CYSTEINE	50
* VITAMIN E.....	53
* METHODOLOGY.....	61
- Experimental Animals.....	61
- Feeding.....	61
- Chemicals.....	61
- Animal Treatment Protocol.....	62
- Experimental Parameters.....	63
Body weight changes.....	64
Water intake and dosage of 2 5-HD.....	64
Clinical neurological evaluation.....	64

Assessment of neurobehavioural function	65
Animals survival.....	69
Sacrifice of the animals.....	69
Histopathlogy.....	70
Enzymes assay.....	72
Gutathione reduactase.....	73
Glutathione peroxidase.....	74
Superoxide dismutase.....	76
Preparation of the spinal cord cytoskeleton	77
Separation of neurofilament proteins.....	78
The electrophoretic run.....	82
Electrotransfer and immunobinding.....	90
Statistical Analysis.....	94
* RESULTS DISCUSSION AND CONCLUSION.....	
* ROLE OF N-ACETYLCYSTEINE.....	96
- Role of N-Acetylcysteine in Prevention and Treatment of 2,5-HD Neurotoxicity.....	96
Results.....	96
Dicussion	124
- Effect of N-acetylcysteine on recovery of 2,5-HD neurotoxicity.....	130
Results.....	130
Discussion.....	136
Conclusion.....	137
* ROLE OF VITAMIN E.....	138
- Role of Vitamin E in Prevention and Treatment of 2,5-HD Neurotoxicity.....	

Results.....	138
Discussion.....	153
Effect of Vitamin E on Recovery of 2,5-HD Neuro-	
toxicity.....	155
Results.....	155
Discussion.....	162
Conclusion.....	163
SUMMARY.....	164
REFERENCES.....	171
BIC SUMMARY	

* * *

10

INTRODUCTION
AND
AIM OF THE WORK

INTRODUCTION

As a result of industrial exposure (Henskowitz et al., 1971) or deliberate inhalation of glues containing hexacarbon solvents (Goto et al., 1974), hexacarbon neurotoxicity is a world-wide problem. Numerous cases of hexacarbon-induced neuropathy have been reported in Italy (Abbritti et al., 1976; Battistini et al., 1974), Japan (Yamamura, 1969), Morocco (Cavignaux, 1972), France (Gaultier et al., 1973), and the United States (Mendell et al., 1974; Allen, 1975; Allen et al., 1975). A similar clinical neuropathy is readily produced in experimental animals (Mendell et al., 1974; Spencer et al., 1978). This syndrome is characterized by degeneration of long-central and peripheral myelinated nerve fibers, resulting in eventual neuromuscular paralysis and sensory alterations (Schaumburg and Spencer, 1976).

It is important to study hexacarbon-induced neurotoxicity not only because of the great potential for human exposure to these solvents, but also, because of the possibility of demonstrating a common mechanism of action of these and other chemicals causing a similar neuropathy.

This study examined the roles of *N*-acetyl-cysteine and Vitamin E in preventing, treating, and/or enhancing recovery of hexacarbon-induced axonopathies.

In addition to the therapeutic benefit, identification of substances that may block or modify the neurotoxic response to hexacarbon compounds might provide clues to the biochemical mechanisms underlying distal axonopathy.

Although the use of 2,5-hexanedione (2,5-HD) in industry is rather restricted, it has been employed as an experimental model of this neuropathy (Spencer and Schaumburg, 1975, 1976) because it is recognized as the ultimate toxic metabolite of the most commonly used neurotoxic hexacarbon compounds in several species, including man (Krasavage et al., 1980). It was selected also

because it is completely soluble in water and can be administered by the oral route (simple) to produce a neurotoxicity identical to that produced by exposure to other hexacarbons (Spencer and Schaumburg, 1975).

Rats were employed because they metabolize the hexacarbons like man and 2,5-HD has the same effect on them as on human (Gilbert and Maurissen, 1982).

In an attempt to participate in the debate regarding the molecular pathogenesis of hexacarbon-induced central-peripheral-distal axonopathy, the tested therapeutic compounds were scrutinized for their effect on neurobehavior, on indigenous protective antioxidant enzyme systems, and on the cytoskeleton of the tested rats.

Employment of N-acetyl-cysteine as a therapeutic agent grew out of the theories proposed by Sabri et al. (1979a) implicating inhibition of sulfhydryl containing glycolytic enzymes as the underlying biochemical lesion for hexacarbon neuropathy.

The popularity of the theory regarding pyrrole adduct formation and auto-oxidation as an essential step in the pathogenesis of hexacarbon neuropathy provided the impetus for selecting Vitamin E (antioxidant). Another reason for this was the work done by Rosenberg et al. (1987b) which demonstrated exaggeration of hexacarbon (2,5-hexanedione) neurotoxicity by hyperbaric oxygen in rats.