

**INCIDENCE AND FREQUENCY OF UREAPLASMA  
UREALYTICUM IN PREGNANT EGYPTIANS  
WITH ASYMPTOMATIC BACTERIURIA**

By

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# ERRATA

Page	Line	erratum	correct
3	12	or lysis osmotic	to lysis by osmotic
5	3	or	on
5	19	50 to 300 mm	50 to 300 nm.
8	7	lines cultures mycoplasmas	line cultures carry mycoplasmas
8	14	no growth at 24 C	No growth at 42 C.
9	5	aagar	agar
10	17	ureapasma	ureaplasmas
10	21	Therefore	therefore
16	24	injection	infection
19	13	20-30 $\mu$ m 200-300 $\mu$ m	20-30 $\mu$ m 200-300 $\mu$ m.
20	18	in	is
26	6	Brumell P	Brunnel P
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38	6	ans its	and it is
38	11	prevalance	prevalence
41	12	Charales	Charles
44	3	to enteric	the enteric
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54	last line	u derlying	underlying
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55	6	a ailable	available
55	12	ha	has
55	12	wel	well
55	13	in idence	incidence
55	20	bacteriura	bacteriuria
95	16	sodium	solid
98	last line	irone slides	iron tubes

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# ***Introduction***

## INTRODUCTION

Asymptomatic bacteriuria can be defined as the presence of single species of bacteriuria in a count of  $10^5$  organisms/ml, or more in clean voided specimen of urine collected from a person with no symptoms referable to the urinary tract.

Bacteriuria is more common in females than in males except in the elderly. In women the incidence rises progressively with the age from about 2% in schoolgirls and 5-10% during the childbearing years to 20% in old age.

Studies of asymptomatic bacteriuria during pregnancy have shown an increasing incidence with increasing parity.

It's generally agreed that, if untreated, asymptomatic bacteriuria early in pregnancy is associated with a high risk (30-50%) of symptomatic acute urinary tract infection later in pregnancy or in the puerperium. It follows that detection of asymptomatic bacteriuria early in pregnancy will prevent about 90% of incidents of acute pyelonephritis during pregnancy.

Although E-coli is the commonest cause of urinary tract infection and for asymptomatic bacteriuria, however other types of organisms could be encountered e.g. indole negative proteus, stap.-saphrophyticus, other gram-negative rods, enterococci, etc. In addition, a variety of fastidious microaerophilic and anaerobic bacteria e.g. Ureaplasma urealyticum have been isolated.

# ***Aim of the work***



AIM OF THE WORK

The aim of this work is to study the incidence of presence of urinary *Ureaplasma urealyticum* among pregnant females with asymptomatic bacteriuria and to correlate this with the presence of other microbes present and with the parity.

# ***Review of literature***

## MYCOPLASMAS

### Bacteriology :

Mycoplasmas are unique group of organisms with the following characteristics :

- 1- The smallest reproductive units have a size of 125-250 nm.
- 2- They are highly pleomorphic because they lack a rigid cell wall and instead are bounded by a triple-layered "unit membrane" "plasma membrane" (Bredt et al, 1973).
- 3- They are completely resistant to penicillin but inhibited by tetracycline or erythromycin.

The lack of the cell wall and the rigid peptidoglycan polymer explains the resistance of the organisms to lysis by lysozyme as well as their susceptibility to lysis osmotic shock and various agents causing the lysis of bacterial protoplasts (Razin 1978).

The mycoplasma membrane is a typical procaryotic plasma membrane built of amphipathic lipids (phospholipids, glycolipids, lipoglycans, sterols) and proteins.(Razin and Rottem 1976; Razin 1981).

Thin sections of mycoplasmas reveal that the cells are built of three organelles only, the cell membrane, the ribosomes and the characteristic procaryotic genome, definitely without mesosomes.(Razin et al 1980).

- 4- They can reproduce in cell-free media; on agar the center of the whole colony is characteristically embedded beneath the surface (fried egg appearance). (Razin and Oliver 1961)

- 5- Growth is inhibited by specific antibody.
- 6- Mycoplasmas don't revert to, or originate from bacterial parental forms.
- 7- Mycoplasmas have an affinity for cell membranes.(Razin et al 1983).
- 8- Usually non motile, but gliding motility have been demonstrated in some species (Bredt 1979). Facultatively anaerobes possessing a truncated flavin-terminated electron transport chain devoid of quinones and cytochromes. An atmosphere of 25% N<sub>2</sub> + 5% CO<sub>2</sub> is preferred for primary isolation.(Noel R.et al 1985).

Gram negative. Catalase negative. Chemo-organotrophic, using their sugar and arginine as the major energy source. Requires cholesterol or related sterols for growth. Parasites and pathogens of a wide range of mammalian and avian hosts. Usually ferment carbohydrates or hydrolyse arginine or urea. Readily destroyed by heat. Don't give rise to inclusion bodies in tissues. G + C content of DNA mostly 23 - 41 moles % (Noel R.et al., 1985).

#### **Morphology and identification :**

Mycoplasma cannot be studied by usual bacteriologic methods because of small size of their colonies, the plasticity and delicacy of their individual cells (due to lack of rigid wall) and their poor staining with aniline dyes. The morphology appears different according to the method of examination. e.g.dark-field, immunofluorescence, Giemsa-

stained films from solid or liquid media, agar fixation.(Razin and Tully, 1983).

Growth on fluid media gives rise to many different forms, including rings, bacillary and spiral bodies, filaments and granules. Filamentous growth is usually associated with young logarithmic cultures growing under optimum conditions. However, filamentous phase is transitory and the filaments transform into chains of cocci which later break apart.(Bred et al.(1973). Phase-contrast or dark-field microscopy of young log-phase broth cultures is the recommended procedure for microscopic examination of mycoplasmas, as it introduces minimal distortions in the shape of the plastic cells. Moreover, it enables observation of gliding motility that characterizes Mycoplasmas. Examination of methanol-fixed organisms stained with Giemsa solution is preferable to the Gram-stain. For electron microscope special attention should be paid to the osmolarity of the fixatives (Lemcke 1972) and the buffers (Cole et al.,1973), as these may drastically alter the actual size and the shape of the plastic organisms. Growth on solid media consists principally of plastic protoplasmic masses of definite shape that are easily distorted. These structures varies in size from 50 to 300 mm in diameter.

Many strains of mycoplasmas grow in heart infusion peptone broth with 2% agar ( pH 7.8 ) to which about 30% human ascitic fluid or animal serum (horse or rabbit) has added. Following incubation at 37° C for 48-96 hours, there may be no turbidity, but Giemsa stains of the centrifuged sediment show the characteristic pleomorphic structures,

and subcultures on solid media yield minute colonies. After 2 - 6 days on special agar medium incubated in a Petri dish that has been sealed to prevent evaporation, isolated colonies measuring 20-500  $\mu$ m can be detected with a hand lens. These colonies are round with a granular surface and a dark center nipple typically buried in agar. They can be subcultured by cutting out a small square of agar containing one or more colonies and streaking of this material on a fresh plate or dropping it into liquid medium. The organisms can be stained for microscopic study by placing a similar square on a slide and covering the colony with a cover glass onto which an alcoholic solution methylene blue and azure has been poured and then evaporated (agar fixation) (Ernest Jawetz et al 1982). Such slides can also be stained with specific fluorescent antibody.

The study of reproduction of mycoplasmas at the level of molecular biology showed clearly that replication of their genome, which must precede cell division, follows the same pattern as with other prokaryotes dividing by binary fission (Morowitz and Wallace 1973).

The mycoplasmas have limited biosynthetic abilities reflecting their small genome & parasitic mode of life. Consequently they require complex media for growth including the serum which provides fatty acids and cholesterol for membrane synthesis, which is unique, (Freundt et al 1980) in an assimilable, non toxic form.

Mycoplasma species are parasites of the mucous membranes and joints, Mycoplasma infection have been most frequently associated with

diseases of respiratory and urogenital tracts, where the parasites firmly adhere to and colonize the epithelial lining. The intimate association between the adhering mycoplasmas and their host cells provides an environment in which local concentrations of toxic by-products excreted by the organisms (i.e.  $H_2O_2$ ,  $NH_3$ ) can accumulate and cause tissue damage. (Razin 1978; Razin et al., 1983). Since no cell wall separates the plasma membrane of the parasite from that of its host, exchange of antigens may occur between the two membrane, an event which may trigger immunological responses of serious consequences to the host (Wise et al., 1978).

The intimate association between mycoplasmas and host cell membranes is reflected also by the "capping" of mycoplasma adhering to lymphocytes, followed by shedding of membrane vesicles of presumed host origin (Stanbridge and Weiss, 1978).

This phenomenon is apparently related to the well-known induction of blast transformation by mycoplasmas (Naot 1982).

#### **Growth characteristics :**

Mycoplasmas are unique in microbiology because of :

- 1- Their extremely small size.
- 2- Their growth on complex but cell-free media.

Mycoplasmas pass through filters with 450 nm pore size and thus are comparable to chlamydiae or large viruses. However, parasitic mycoplasmas grow on cell free media that contain lipoprotein & sterol.