ISOLATION AND CHARACTERIZATION OF ENVIRONMENTAL STRESS RELATED GENES FROM SOME WILD PLANTS

By

HAYTHAM MOHAMED ABD ELHALIM

B.Sc. Agric. Sc. (Genetics), Ain Shams University, 2003

A thesis submitted in partial fulfillment of the requirements for the degree of

in
Agricultural Science
(Genetics)

Department of Genetics
Faculty of Agriculture
Ain Shams University

Approval Sheet

ISOLATION AND CHARACTERIZATION OF ENVIRONMENTAL STRESS RELATED GENES FROM SOME WILD PLANTS

By

HAYTHAM MOHAMED ABD ELHALIM

B.Sc. Agric. Sc. (Genetics), Ain Shams University, 2003

This thesis for M.Sc. degree has been approved by:	
Prof. Dr. Mahmoud Imam Nasr	
Prof. Emeritus of Genetics, Genetic Engineer	ing and
Biotechnology Research Institute, Minufiya U	Jniversity
Prof. Dr. Samir Abd El-Aziz Ibrahim	
Prof. of Genetics, Faculty of Agriculture, Ain University	Shams
Prof. Dr. Ahmed Bahieldin Mohamed	•••••
Prof. of Genetics, Faculty of Agriculture, Ain University	Shams

Date of Examination: 28/4/2010

ISOLATION AND CHARACTERIZATION OF ENVIRONMENTAL STRESS RELATED GENES FROM SOME WILD PLANTS

By

HAYTHAM MOHAMED ABD ELHALIM

B.Sc. Agric. Sc. (Genetics), Ain Shams University, 2003

Under the supervision of:

Prof. Dr. Ahmed Bahieldin Mohamed

Prof. of Genetics, Department of Genetics, Faculty of Agriculture, Ain Shams University (Principal Supervisor)

Dr. Ahmed Mohamed Kamal Nada

Researcher of Genetics, Department of Plant Molecular Biology, Genetic Engineering Research Institute, Agricultural Research Center

ABSTRACT

Haytham Mohamed Abd-Elhalim Ismail: Isolation and Characterization of Environmental Stress Related Genes from Some Wild Plants. Unpublished M.Sc. Thesis, Department of Genetics, Faculty of Agriculture, Ain Shams University, 2010.

Proline is the most common organic solutes that play an active role in the mechanism of plant salt tolerance. Ornithine aminotransferase (-OAT) is a key enzyme in proline biosynthetic pathway and is implicated in salt tolerance in higher plants. In this study full-length coding sequence encoding the Ornithine aminotransferase enzyme was isolated from legume Vicia villosa by RT-PCR approach. *In-silico* mapping of *Vicia villosa* Ornithine aminotransferase (VvOAT) revealed that this gene is located on barrel medic chromosome 3, lotus japonicus chromosome 1, soybean chromosome 4 and common bean chromosome 1. Docking generated 3D model of the gene with its known substrate and inhibitors was performed, the results indicated that PHE170 and ASN171 in VvOAT are the important determinant residues in binding substrate and inhibitors. Obtained protein from gene expression showed a molecular mass compatible with the predicted mass, confirming that the protein is fully intact and not truncated. After gene delivery into E. coli host HB101, VvOAT gene enhanced its salt tolerance.

Key words: *Vicia villosa* - Salt tolerance – Osmoprotectants - Ornithine -aminotransferase - RT-PCR

ACKNOWLEDGMENT

Thanks for my God Allah, the great and almighty on his uncountable and infinite graces

I wish to express my deep gratitude and sincere appreciation to **Dr.** Ahmed Bahieldin, Professor of Molecular Genetics, Genetics Dept., Faculty of Agriculture, Ain Shams University and Director of AGERI/ARC for his inspiring and capable leadership, continuous supervision, kind encouragement, and precious advices during the progress of thesis work. Great appreciation is also expressed to **Dr. Fotouh M.A. El-Domyati**, Professor of Molecular Genetics, Genetics Dept., Faculty of Agriculture, Ain Shams University for his continuous supervision, kind encouragement, sincere support and appreciable invaluable criticism. Thanks are also extended to Dr. Ahmed Nada, Researcher, AGERI, ARC for his supervision, unlimited guidance and valuable assistance in writing the manuscript. Sincere thanks to Dr. Rania Mahmoud Ismail, Researcher, AGERI, ARC for here valuable advice and help during the course of this study and for revising this thesis. I am grateful to all my friends at (AGERI) who so helped with their encouragement. I must especially mention **Doaa El-Gharib**. Very special thanks to all my colleagues at (MMGT) laboratory, Mohamed Youssef, El-Zahraa Radwan, Lamiaa Salah, Marwa Samir, Asmaa Ezzat, Menna-T-Allah Mohamed, Aya Ahmed, Asmaa El Fiky, Afaf **Elgarhy** and **Asmaa Ghanem** who went out of their way so many times to help me.

CONTENTS

	Page
LIST OF TABLES	IV
LIST OF FIGURES	V
LIST OF ABBREVIATION	VII
I. INTRODUCTION	1
II. REVIEW OF LITERATURE	3
1. Domestication and Agriculture	3
2. Plant abiotic stress	4
2.1. Salinity stress	5
2.1.1. Salinity effects on plants	7
2.1.2. Plant response to salt stress	8
2.1.3. A key enzyme involved in proline biosynthesis, Ornithineaminotransferase (-OAT)	18
2.1.4. Improving crop salt tolerance	28
3. Selected recent studies of salt stress-related genes isolated and	
characterized through PCR based methods	30
III. MATERIALS AND METHODS	35
1. Plant materials	35
2. RNA manipulation	35
2.1. Purification of RNA from plant material	35
2.2. RNA Electrophoresis	36
2.3. RNA sample preparation	37
2.4. RNA gel run	37
2.5. Photography of RNA in agarose gel	37
2.6. Determination of RNA concentration	38
3. Primer design for PCR amplification	38
3.1. Primer preparation and dilution	39
4. RT-PCR (cDNA first strand synthesis)	39
5. RT-PCR (cDNA second strand synthesis)	40
5.1. DNA Electrophoresis	42

5.2. DNA sample preparation	43
5.3. DNA gel run	43
5.4. Photography of DNA in agarose gel	44
6. Purification of DNA from agarose gel	44
7. Cloning of OAT gene	45
7.1. DNA ligation	45
7.2. Preparation of competent cells	46
7.3. Transformation of competent cells	47
7.4. Miniprep of plasmid DNA	48
7.5. Screening of recombinants	50
7.5.1. Screening of recombinants by PCR	50
7.5.2. Screening of recombinants by restriction digestion	51
7.6. Bacterial glycerol stocks	52
8. Sequencing of cDNA insert	52
9. Identification of cDNA fragment using NCBI database	53
10. Bioinformatics analyses	53
10.1. DNA sequence analysis	53
10.1.1. In-silco mapping	53
10.2. Protein sequence analysis	54
10.2.1. Protein 3D modeling	54
10.2.2. Docking studies	54
10.2.3. Evolution studies	55
11. Detection of expression and function analysis	55
11.1. Subcloning of -OAT into prokaryotic expression vector	55
11.2. Detection of protein expression using SDS-PAGE	57
11.3. Function study of VvOAT into prokaryotic system	60
IV. RESULTS AND DISCUSSION	62
1. Reverse Transcriptase Polymerase Chain Reaction (RT-PCR) of	
Ornithine -aminotransferase (-OAT) gene	62
1.1. Abscisic acid (ABA) treatment for <i>Vicia villosa</i> seedlings1.2. RNA isolation and cDNA first strand synthesis	62
1.2. KINA ISOIAHOH AHU UDINA HISI SITAHU SYHURUSIS	63

1.3. Second strand synthesis	64
1.3.1. Primer design	64
1.3.2. Amplification of -OAT	
	64
2. Cloning and sequencing of -OAT gene	66
2.1. Cloning of the PCR product into pGEM–T Easy vector	66
2.2. Molecular screening of recombinant clones	67
2.2.1. Screening using restriction digestion	67
2.2.2. Screening using PCR	68
2.3. Sequencing of the PCR product	68
2.4. Homology analysis of VvOAT	72
3. In-silco analyses	78
3.1. Nucleic acid sequence analysis	78
3.1.1. Identification of putative chromosomal location	78
3.2. Protein sequence analysis	82
3.2.1. Homology modeling	82
3.2.2. Docking VvOAT protein	82
3.2.3. Evolution investigation	83
4. Studying the protein expression and function	84
4.1. Detection of expression	84
4.1.1. Subcloning of VvOAT into PET system	84
4.1.2. Detection of VvOAT expression	87
4.2. Detection of VvOAT function	90
V. SUMMARY	92
VI. REFERENCES	
VI. NITI ITALIA (P.)	95

LIST OF TABLES

No.		Pages
(1)	RT-PCR reaction assembly	40
(2)	Resolving and staking gels for 12.5% SDS-Gel	59
(3)	Designed Primer sequence for -OAT ge	ene 65
(4)	Docking VvOAT	83

LIST OF FIGURES

No.		Pages
(1)	The complexity of the plant response to abiotic stress.	19
(2)	pGEM-T Easy vector circular map	48
(3)	pET-30b circular map	57
(4)	Total RNA of Vicia villosa seedlings shoots	64
(5)	RT-PCR of -OAT	65
(6)	Candidate band after purification	66
(7)	LB-Agar plates containing transformed single white and single blue <i>E. coli</i> colonies	67
(8)	Digestion of candidate colonies plasmid DNA with EcoRI enzyme	69
(9)	Digestion of candidate colonies plasmid DNA with Pstl enzyme	69
(10)	Screening for OAT using Sp6 and T7 primers	70
(11)	Screening for OAT using gene specific primers	70
(12)	Nucleotide sequence of VvOAT gene	73
(13)	Calculated DNA statistics and calculated Protein	
	properties	74
(14)	Prediction of VvOAT secondary structure	74

(15)	Blastn results of VvOAT DNA	76
(16)	Blastp results of VvOAT Protein	76
(17)	Blastp results of VvOAT Protein, conserved domain hits	76
(18)	Codon usage of VvOAT	77
(19)	Comparative map 1	80
(20)	Comparative map 2	81
(21)	3D Model of <i>Vicia villosa</i> Ornithine - aminotransferase gene	82
(22)	Alignment of the predicted amino acid sequence of Vicia villosa (VvOAT) and those of several other	0.5
(23)	plants Phylogenetic analysis of OATs	85 86
(24)	pET-30b vector and VvOAT Insert purification	88
(25)	PCR screening for VvOAT inside p6X-VvOAT	88
(26)	Cloning of VvOAT into pET 30-b	89
(27)	Generation of chimera p6XVvOAT	89
(28)	VvOAT Expression in <i>E. coli</i>	90
(29)	Effects of NaCl on transformed E. coli strain HB101	91

VII

LIST OF ABBREVIATION

ABA Abscisic acid

ACP Annealing-control-primer

APX Ascorbate peroxidase

Blast Basic local alignment search tool

CBF C-repeat binding factors

Cd Conserved domain

DEGs Differentially expressed genes

DREB Dehydration response element binding protein

EST Expression sequence tags

FAO Food and Agriculture Organization

GSA Glutamate semialdehyde

Hsps Heat-shock proteins

IAA Indole-acetic acid

LEA Late embryogenesis abundant

MAPK Mitogen activated protein kinase

NJ Neighbor-joining

OAT Ornithine aminotransferase

ORF Open reading frame

Orn Ornithine

P5CDH P5C dehydrogenase

P5CR Pyrroline-5-carboxylate reductase

P5CS Pyrroline-5-carboxylate synthase

VIII

PAGE Polyacrylamid gel elecrtophresies

PCR-SSH PCR-based suppression subtractive hybridization

PDB Protein Data Bank

PDH Pro dehydrogenase

PLP Pyridoxal-5-phosphate

PRE Proline responsive elements

Pro Prolin

RACE Rapid amplification of cDNA ends

RT-PCR Reverse transcriptase-polymerase chain reaction

ROS Reactive Oxygen Specie

SOD Superoxide dismutase

SOS Salt overlay salinity

TFs Transcription factors

I. INTRODUCTION

Early communities were situated near reliable water sources with the consequence that crops became gradually adapted/selected for growth under near optimal conditions of water supply (Bressan et al., 2009), they established agrarian societies in order to give rise to the civilization.

One of the major obstacles encountering the development of agrarian societies is abiotic stress, such as drought, salinity, extreme temperatures, chemical toxicity and oxidative stress such stresses are serious threats to agriculture and result in the deterioration of the environment. Abiotic stresses are considered as the primary causes of crop loss worldwide, reducing average yields for most major crop plants by more than 50% (Boyer, 1982; Bray et al., 2000).

Salt stress is one of the abiotic stresses that impose a major environmental threat to agriculture. Its adverse impacts are getting more serious problem in regions where saline water is used for irrigation (**Türkana and Demiral, 2009**). The increased salinization of arable land showed that it is expected to have devastating global effects, resulting in 30% land loss within the next 25 years, and up to 50% by the year 2050 (Wang et al., 2003).

Salinity affects all the major processes such as growth, photosynthesis, protein synthesis, and energy and lipid metabolism (Parida and Das, 2005).

The final effect is estimated as death of plants or decrease in productivity. Suppression of growth occurs in all plants, but their tolerance levels and rates of growth reduction at lethal concentrations of salt vary widely among different plant species.

One of the most common stress responses in plants is overproduction of different types of compatible organic solutes (Serraj and Sinclair, 2002). Proline accumulation is one of the most frequently reported modifications induced by water and salt stresses in plants and is often considered to be involved in stress resistance mechanisms, although its precise role still remains a controversial subject. Cytoplasmic accumulation of this amino acid is thought to be involved in osmotic adjustment of stressed tissues (Delauney and Verma 1993, Kavi Kishor et al., 1995).

Ornithine -aminotransferase (OAT) is an important enzyme in proline biosynthetic pathway and is implicated in salt tolerance in higher plants. OAT transaminates ornithine to pyrroline-5-carboxylate, which is further catalyzed to proline by pyrroline-5-carboxylate reductase (Sekhar et al., 2007).

It was reported that the activity of -OAT as well as the proline contents in plants increased when treated with cold (Charest and Phan, 1990), salt (Roosens et al., 1998), dehydration (Hervieu et al., 1994) and ABA (Abscisic acid) (Yang et al., 2000).

There are two research topics which elucidate the significance of studying OAT: (i) production of genetically modified plants for increased production of the osmoprotectant proline, (ii) considering human OAT as a potential target for development of new therapeutic drugs (Stránská et al., 2008).

The present study aims to isolate and characterize ornithine -aminotransferase gene using ABA as inducer for gene expression and to give a primary estimates of the encoded protein function.