Effect of Postpartum Maternal Antibiotics On Colonization of Gastrointestinal Flora in Breast Fed Infants

Thesis
Submitted of Partial Fulfillment of M.Sc. Degree
In Pediatrics

By
Essam Saad Ibrahem Mohamed
(M.B.B.Ch)

Under Supervision of
Prof. Dr. Hisham Abdel Samei Awad
Professor of Pediatrics
Faculty of Medicine – Ain Shams University

Prof. Dr. Laila Mostafa Al Kady Professor of Microbiology and Immunology Faculty of Medicine – Zagazig University

Dr. Soha Mohamed Khafagy Lecturer of Pediatrics Faculty of Medicine – Ain Shams University

> Faculty of Medicine Ain Shams University 2012

إِنَّمَا يَخْشَكُ اللَّهُ مِنْ عَبَادِهِ الْحُلَمَاءُ إِنَّ اللَّهَ عَزِيزٌ غَفُورٌ

صدق الله الهظيم (فاطر آية ۲۸)



First and before all, I thank **Allah** who made all things possible.

I thank him for his great mercy, generous blesses, and for his continuous gifts.

I would like to express my very great appreciation to Professor **Prof. Dr. Hisham Abdel Samei Awad,** Professor of Pediatrics, Faculty of Medicine, Ain Shams University, for his generous help, advice and encouragement throughout this work.

I would like to offer my special thanks to **Prof. Dr. Laila**Mostafa Al Kady, Professor of Micrrobiology and Immunology,

Faculty of Medicine, Zagazig University, for her kind supervision and continuous guidance throughout the course of this work.

My great appreciation and deep thanks to **Dr. Soha**Mohamed Khafagy, Lecturer of pediatric, Faculty of Medicine,

Ain Shams University. It must be mentioned here that this work

wouldn't have been seen without her encouragement and immeasurable

assistance on designing and executing good experimental research.

Essam Saad



I dedicate this work

То

My Mother, Wife, Daughters and My Family.

Content

Subject	Page
List of abbreviations	I
List of Figures.	II
List of Tables.	III
Introduction	1
Aim of the work	3
Review of literature	4
Chapter 1: Infant GIT	4
Chapter 2: Breastfeeding	27
Chapter 3: Effect of maternal antibiotics on infant	51
Subject and methods	80
Results	96
Discussion	108
Summary & Conclusion	117
Recommendations	120
References	121
Arabic summary	-

List Of Abbreviation

AAD Antibiotic-Associated Diarrhea AAP American Academy of Pediatrics

B. bifidum
B. fragilis
B. infantis
BHIAbifidobacteria bifidum
bifidobacteria infantis
brain-heart infusion agar

C. difficileC. perfringensCfuClostridia difficileClostridia perfringenscolony flowing unit.

ELBW extremely low birth weight

F6PPK Fructose-6-phosphate phosphoketolase

G6PD glucose-6-phosphate deficiency
GAC germfree animal characteristics
GPW glucose-phosphate peptone water
HIV human immunodeficiency virus

Ig immunoglobulin KOH potassium hydroxide

LBS Agar Lactobacillus Selection Agar

M/P (milk to plasma)

MACs microflora associated characteristics

MR Methyl- Red

MRS De Man Rosa Sharpe

NaCl sodium chloride

NEC neonatal necrotizing enterocolitis

P. morgani proteus morgani
P. vulgaris proteus vulgaris

Pg Pico gram

pps peptone physiological saline

sp. species

TB tuberculosis

TPY trypticasephytone- yeast TSI agar triple sugar iron agar VLBW very low birth weight

WHO World Health Organization

List Of Figures

Figure (1): Sabouraud dextrose agar plate culture of Candida.	90
Figure (2): A MacConkey agar plate with an active Enterobacteria culture.	91
Figure (3): E. coli on MacConkey agar	91
Figure (4): Reaction of indole positive and MR positive by	92
E.coli.	
Figure (5): Reaction on TSI agar by E. coli.	92
Figure (6): Urease test for E. coli.	93
Figure (7): E. coli with positive MR test	93
Figure (8): Lactobacilli on LBS agar.	94
Figure (9): Catalase test results. (Top) The positive (bottom) the	94
negative	
Figure (10): Bifidobacteria gram-Stained	94
Figure (11): Bifidobacteria on Beerens agar	95

List Of Tables

Table 1: Approximate clearance values at different ages (Bennett and the WHO Working, 1997).	48
Table 2: Summary of distribution of Antibiotics into breast milk (Speight and Holford, 1997).	49
Table (3): Comparison between group 1 and group 2 as regards descriptive data.	96
Table (4): Oral swab results for oral moniliasis among group 1 and group 2.	97
Table (5): Comparison between group 1 and group 2 as regards Intestinal Enterobacteria count in Stool culture	98
Table (6): Comparison between group 1 and group 2 as regards Intestinal E. coli count in stool culture.	99
Table (7): Comparison between group 1 and group 2 as regards Intestinal Lactobacilli count in stool culture.	100
Table (8): Comparison between group 1 and group 2 as regards Intestinal Bifidobacteria count in stool culture.	101
Table (9): Correlation between the expressions of intestinal flora count & Antibiotic duration and Number of breast feds.	102
Table (10): Correlation between oral swab results and (antibiotic duration and number of breast feds).	103
Table (11): Correlation between the expressions of Intestinal Enterobacteria count and antibiotic duration.	104
Table (12): Correlation between the expressions of Intestinal E. coli count and antibiotic duration.	105
Table (13): Correlation between the expressions of Intestinal Lactobacilli count and antibiotic duration.	106
Table (14): Correlation between the expressions of Intestinal Bifidobacteria count and antibiotic duration.	107

Introduction

Antibiotic usage is fairly common among breastfeeding mothers and there is potential for transfer to infants through breast milk. While most medicines taken by lactating women cause no harm to their babies, at times, it can result in serious consequences (Mathew, 2004).

For various reasons, antibiotics are one of the commonest groups of drugs prescribed to postpartum mothers. This trend is noticeable both in the developing and developed countries (*Thomas*, 1994).

The majority of breast milk components act similar to plasma components and theoretically all drugs have the potential of crossing over from maternal plasma into breast milk. Molecular transfer depends on the usual mechanisms; passive diffusion, active transport against a concentration gradient and transcellular diffusion (*Chung et al.*, 2002).

The fetal gastrointestinal tract is sterile until birth when microbes colonize the gastrointestinal tract, and a dense complex microbiota are develops (*De la Cochetier et al.*, 2004).

In general, infants initially colonized by *enterobacteria* and gram-positive cocci, which are thought to create a reduced environment that is favorable for the establishment of *Bacteroides, Bifidobacterium*, and *Clostridium* by day seven (*Favier*, 2002).

A full term breast-fed infant has an intestine microbiota in which *bifidobacteria* and *lactobacillus* predominate over potentially harmful bacteria. A diet of breast milk induces the development of a flora rich in *Bifidobacterium* (*Dia and Walker*, 1999).

Loss of normal GIT flora and GIT colonization is associated with the use of antibiotics particularly third generation cephalosporin (Saiman et al., 2001).

All antibiotics can cause three potential problems for nursing infants. Firstly, they can modify the bowel flora and alter gut defense mechanisms; this can result in diarrhea and mal absorption of nutrients. Secondly, they may have direct effects that may or may not be dose related. Lastly, and often ignored, is that antibiotics can alter and interfere with microbiological culture resulting in babies being investigated for sepsis (American Academy of Pediatrics, 1994).

Aim of The Work

The aim of this study was determine the effects of postpartum maternal intake of systemic antibiotics on type of flora colonizing the GIT in breast fed infants and to trace such colonization pattern for the 1st 6 months of life.

Also, we aimed at detecting the effect of postpartum maternal antibiotics intake on the incidence and duration of oral moniliasis in their breast fed babies.

Chapter 1 INFANT GIT

Intestinal Microbiota in Neonates:

The fetal gastrointestinal tract is sterile until birth when microbes colonize the gastrointestinal tract, and a dense, complex microbiota develops. This enormous cell mass performs a variety of activities that affect both the intestinal and systemic physiology. The microbiota provides nutritional, metabolic, immunological, and protective functions. The neonatal gastrointestinal tract is an organ at risk. Increasing awareness that the human flora is a major factor in both health and disease has led to different strategies to manipulate the flora (*Fanaro et al.*, 2003a).

During the birth Process and shortly thereafter, microbes from the mother and the surrounding environment colonize the gastrointestinal Tract of the infant until a dense, complex microbiota Develops. The establishment of the gut microbial population is not strictly a succession in the ecological sense rather, this colonization is a complex process influenced by microbial and host interactions as well as by internal and

External factors. The climax intestinal flora is attained in successive stages (Favier et al., 2002).

Primary portals of entry for these bacteria include the birth canal during delivery and contact with other humans and the surrounding environment. After birth, many factors influence the types and quantities of intestinal micro flora of the infant. Besides extrinsic factors such as the mother's dietary intake or use of probiotics, type of birth (vaginal or surgical), gestational age, and primary source of nutrition (bottle or breast fed), intrinsic factors including underlying neonatal health, immunologic status, gastrointestinal transit time, pH, and stress all affect the process of colonization and the types of organisms established (Hooper, 2004).

Intestinal microflora in early infancy: composition and development:

The neonatal intestinal microbiota is a complex ecosystem composed of numerous genera, species and strains of bacteria. This enormous cell mass performs a variety of unique activities that affect both the colonic and systemic physiology. Its primary activities include nutritive, metabolic, immunological and protective functions (*Fanaro et al.*, 2005).

The intestinal microflora is estimated to harbour about 400 different microbial species, mostly bacteria. More than 99% of

the cultivable fecal microbiota is represented by 30–40 bacterial species (*Tannock*, 2001).

Other numerical predominant groups are bifidobacteria, eubacteria, clostridia and lactobacilli and the population level of each species appears to be directly regulated by the competition for nutrients and by physical space. On the other hand, some bacteria do not adhere to the intestinal mucosa and prefer different microhabitats (*Schrezenmeir and de Vrese*, 2001).

Four distinct bacterial microhabitats can be identified in the intestinal tract: (*Penders et al.*, 2006a)

- a) The surface of the epithelium cells,
- b) The crypts,
- c) The mucous gel layer, and
- d) The intestinal lumen.

The components of the bacterial ecosystem have a high degree of heterogeneity and compete to occupy the receptors of the different adhesion sites (*Schrezenmeir and de Vrese*, 2001).

The normal microflora represents the first line of defense against several potential pathogens by direct interference with the settlement of newly arrived species and is in close contact and balance with the defense system of the host (Edwads and Parrett, 2002).

The different bacterial strains present an important immunological challenge to the whole immune system and may influence immunological challenges as well as immunological tolerance. The normal host resident organisms enhance humeral immune response, activate non-specific host resistance to pathogens, modulate the host immune response to potential harmful antigens and down-regulate hypersensitivity reactions (Isolauri et al., 2001).

It is increasingly clear that the gut-associated lymphoid tissue, which constitutes one of the most important elements in the total immunological capacity of the host, is crucial for the recognition of several kinds of bacterial, and non-bacterial, antigens and for the subsequent immune response or tolerance (Forchilli and Walker, 2005).

Physiological Colonization of Microflora:

In general, infants are initially colonized by enterobacteria and gram-positive cocci, which are thought to create a reduced environment that is favorable for the establishment of Bacteroides, Bifidobacterium, and Clostridium by day seven (*Park et al.*, 2005).

In full-term infants, a diet of breast milk induces the development of a flora rich in Bifidobacteria and Lactobacillus.