

Intralesional Tuberculin as an Immunotherapy in Warts

Thesis

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List of Abbreviations

AKs	Actinic keratoses
ALA	α -aminolaevulinic acid
APCs	Antigen-presenting cells
BCG	Bacille Calmette-Guerin
CIN	Cervical intraepithelial neoplasia
CO ₂	Carbon dioxide
DNA	Deoxyribonucleic acid
DNCB	Dinitrochlorobenzene
DPCP	Diphencyprone
E	Early region
Er:YAG	Erbium: Yttrium/Aluminum/Garnet
EV	Epidermodysplasia verruciformis.
FDA	Food and drug administration
FISH	Filter in situ hybridization
GM-CSF	Granulocyte-macrophage colony stimulating factor
HIV	Human immunodeficiency virus
HPV	Human papilloma virus
HSIL	High-grade squamous intraepithelial lesion
IFN	Interferon
IL	Interleukin
ISH	Tissue in situ hybridization
Kb	Kilobases
L	Late region
LCs	Langerhans' cells
Mw	Mycobacterium w
Nd:YAG	Neodymium: Yttrium/Aluminum/Garnet

List of Abbreviations (Cont.)

NF-K β	Nuclear factor κ B
NKCs	Natural killer cells
ORFs	Open reading frames
p 53	Protein 53
PCR	Polymerase chain reaction
PPD	Purified protein derivative
pRB	Retinoblastoma tumor suppressor protein
RRP	Recurrent respiratory papillomatosis
SADBE	Squaric acid dibutylester
SCC	Squamous cell carcinoma
STH	Southern transfer hybridization
STI	Sexually transmitted infections
TCA	Trichloroacetic acid
Th 1	T-helper 1
Th 2	T-helper 2
TLR	Toll-like receptor
TNF	Tumor necrosis factor
Tu	Tuberculin unit
UK	United Kingdom
URR	Upstream regulatory region.
US	United States
VLP	Virus-like particles
VV	Verruca vulgaris
5 -fu	5 -Fluorouracil
5 -FU+LE	5 -fluorouracil, lidocaine, and epinephrine

Introduction

Warts are tumours or growths of the skin caused by infection with Human Papillomavirus (HPV). More than 100 HPV subtypes are known (*Pre'tet et al., 2004*).

Warts are particularly common in childhood and are spread by direct contact or autoinoculation. This means if a wart is scratched, the viral particles may be spread to another area of skin. The incubation period varies from 1 to 12 months and averages 2 to 3 months (*Karagas et al., 2004*).

Warts have a hard warty or verrucous surface. Clinically, warts are classified by location into three main subtypes: cutaneous, anogenital/mucosal and epidermo-dysplasia verruciformis (EV) (*Pfister et al., 2003*).

There are various types of viral warts including common warts, plantar warts, mosaic warts, plane or flat warts, periungual warts, filiform warts, oral warts and genital warts (*Aubin and Laurent, 2004*).

There are many therapeutic modalities available for the treatment of warts including occlusion by duct tape, chemical treatment using salicylic acid or similar compounds, cryotherapy with liquid nitrogen, electrosurgery (curettage and cautery), and laser vaporization, but none of them offers a guarantee of cure and recurrence is common (*Scheinfeld and Lehman, 2004*).

There are new trends towards the use of immunotherapy in treatment of warts, as the immune system seems to play an important role in the control of warts infection. Although the exact

mechanisms are unclear but most evidences suggest that cell mediated rather than humoral immunity plays an important role in control of HPV infection as the incidence of warts increases in subjects with cell mediated immune defects e.g. (HIV infection patients, malignant diseases...etc) (*Frazer, 1991 and Contant, 2000*).

Various methods have been used to stimulate immunological response e.g. imiquimod, intralesional interferons, intralesional immunotherapy using mumps, Candida or trichophyton skin test antigens (*Horn et al., 2000*).

In the trial of mumps and candida antigens, patients with warts were tested for immunity to mumps and Candida using commercial antigens then according to the response to the test injection some patients did not respond and some patients had detectable immunity. The immune group received intralesional mumps and Candida antiserum. About 42% of patients had complete clearance of the treated warts and about 48% of patients had complete resolution of their immunotherapy treated warts and also complete resolution of the untreated distant warts so the intralesional mumps and Candida immunotherapy proved efficacy in treatment of warts (*Johnson et al., 2001*).

Recently, intralesional tuberculin has been used for treatment of warts in Turkey, taking the advantage of vaccination schedule in this country (*Kus et al., 2000*).

BCG is given routinely in vaccination in Egypt (*Kotb and Azza, 1997*). No studies have been done to evaluate the efficacy of tuberculin use in treatment of warts in Egypt.



Aim of the Work

The aim of this study is to evaluate the effectiveness and adverse reactions, if present, of intralesional injection of tuberculin antigen (Purified protein Derivative, PPD) in treatment of warts.

1.1.1. Human Papilloma Virus (HPV)

1.1.1. Virology of HPV:

HPV represents a complex group of small DNA tumor viruses that belong to genus A of the family Papovaviridae (*Melnick et al., 1974*).

Papillomaviruses are non-enveloped, double-stranded DNA viruses approximately 50 nm in diameter. They are spherically shaped viral particles (virions) which consist of an outer protein shell (capsid) surrounding a single closed super coiled circle of double stranded DNA of about 8000 nucleotide base pairs (8 kilobases kb) that comprises the HPV genome (*Van Ranst et al., 1997*).

The HPV genome is composed of three domains: the upstream regulatory region (URR), the early region (E), and the late region (L). The URR is approximately 1 kb in length, and lacks open reading frames (ORFs). It contains the origin of replication and many of the control elements for transcription and replication. The early region is approximately 4 kb in length, and contains the ORFs for the viral genes that are principally expressed early in the papillomavirus life cycle. The late region is approximately 3 kb in length, and encodes the viral capsid proteins (*Howley and Lowy, 2001*).

The HPVs have two genera, the genus α -papillomavirus containing mucosal and cutaneous HPV and the genus

β-papillomavirus containing the types associated with epidermodysplasia verruciformis (EV). Within each genus, HPV types are grouped according to the DNA sequence homology into species that often share similar biologic and pathologic properties (e.g. HPV-16 and -31) (*DeVilliers et al., 2004*).

1.1.1. Early genes:

The early gene includes E1, E2, E4, E5, E6 and E7. The E genes are transcribed at low levels in the basal cell layer and stratum malpighii. The first genes to be expressed following infection are the E1 and E2 genes, which are viral regulatory proteins that are responsible for controlling the transcription of the viral genes and replication of the viral genome. The E4 protein is expressed in the terminally differentiated keratinocytes where it causes cytokeratin collapse, thus facilitating the assembly and maturation of the virus. E4 is probably a late gene because it is expressed late in the cycle of virus replication (*Tyring, 2000*).

One major effect of HPV proteins E5, E6 and E7 is that the epidermal cell cycle, which is normally blocked for cells that are supra-basal, continues so that HPV genome copy number can be amplified to high levels during viral replication for assembly into virions. The E6 and E7 proteins of the high-risk mucosal HPV types act as viral oncoproteins, but no such functions are associated with the corresponding proteins of the low-risk mucosal and EV types. There is limited understanding

of possible differences between the oncogenic EV HPV types (HPV- α , - β) and the non-oncogenic EV HPV types (*Pfister, 2003*).

In combination with other cellular proteins, E β from high-risk mucosal HPV causes degradation of the cellular protein p 53 , so that E β -facilitated destruction of p 53 removes a brake on supra-basal cell cycling (*Mantovani and Banks, 1999*).

The E γ protein binds the retinoblastoma tumor suppressor protein (pRB) leading to induction of DNA replication of HPV genome. Both E β and E γ are multifunctional proteins and while their effects on p 53 and pRB are critical ones, they do have additional targets important to the oncogenic potential of the virus (*Duensing and Munger, 2003*).

1.1.2. Late genes:

The two late genes, L β and L γ encode the major and minor capsid protein respectively, these are the structural proteins of the virion. L β is the major capsid protein which represents about 90% of the total virion protein, and L γ is a minor capsid protein which accounts for only a small portion of the virion mass. The function of L γ protein is not yet clear (*Kimbauer, 1996*).

During virion assembly within cells, L β multimerizes into pentamers called capsomeres, and $\gamma\gamma$ capsomeres multimerize to

form the viral capsid with an icosahedral symmetry. The capsid surrounds the viral DNA, thereby protecting it from degradation, and it also enables the virus to bind efficiently to target cells (*Kimbauer et al., 1992*).

When expressed in cell culture, the L1 major capsid proteins of all papillomavirus types tested self-assemble into virus-like particles (VLP) that are morphologically similar to native virions. In addition, papillomavirus VLP display type-specific and neutralizing surface epitopes and are therefore useful as an antigen to detect serum antibodies and as a subunit prophylactic vaccine (*Lowy and Frazer, 2003*).

1.2.1. Pathogenesis:

The papillomaviruses are highly species-specific, and productive infection has never been observed outside their natural host tissue (no known infection of man by animal HPV). HPVs are also tissue specific, i.e. the papillomavirus life cycle is completed only in fully differentiated squamous epithelia. This has impeded its study in monolayer tissue culture cells, where late gene expression and virion production do not occur (*Howley and Lowy, 2001*).

Infection begins with viral entry followed by one of three paths: Latent infection, in which there is no gross or microscopic evidence of disease, subclinical infection, in which microscopy reveals evidence of infection in the absence of