# Relation between blood CD49d and muscle ultrasonography in children with Duchenne Muscular Dystrophy

#### **Ehesis**

Submitted for partial fulfillment of Master degree in Pediatrics

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# **List of Contents**

Subject	age No.
List of Abbreviations	i
List of Tables	ii
List of Figures	iii
Introduction	1
Aim of the Study	4
Review of Literature	
Duchenne Muscle Dystrophy	5
Diagnostic Ultrasound in Duchenne Muscular Dystrophy	58
Role of VLA-4 (CD49d/CD29) on T cell traffick Dnchenne Muscular Dystrophy	
Patients and Methods	73
Results	87
Discussion	111
Summary	123
Conclusion	126
Recommendations	127
References	128
Arabic Summary	

## **List of Abbreviations**

Abbr. Full-term

**DMD** : Duchenne Muscle Dystrophy

**DGC**: Dystrophin-associated glycoprotein complex

MUS : Muscle ultrasound

**QMUS** : Quantitative muscle ultrasound

**KD** : Kilodaltons

**n NOS** : Neuronal nitric oxide synthase

**I NOS** : Inducible nitric oxide synthase

**SDB** : Sleep-disordered breathing

**REM** : Rrapid eye movement sleep

**WAIS** : Wechsler Adult Intelligence Scale

**IQ** : Intelligence Quotient

**CK** : Creatine kinase

**EMG** : Electromyography

**ATPase** : Adenosine triphosphatase

**MLPA** : Multiplex ligation-dependent probe amplification

**PCR** : Polymerase chain reaction

#### List of Abbreviations

**FISH** : Fluorescence in situ hybridization

DHPLC : Denaturing high performance liquid chromatographySCAIP : Single-condition amplification internal primer sequencing

**DGGE** : Denaturing gradient gel electrophoresis

**DOVAMK-S**: Detection of virtually all mutations - single-strand

**DCM** : Dilated cardiomyopathy

**NIPPV**: Intermittent positive pressure ventilation

**DXA** : Dual energy x-ray absorptiometry

**25OHD** : 25-hydroxyvitamin D

**AAN** : American Academy of Neurology **rAAV** : Recombinant adeno-associated viral

**FDA** : Food and Drug Administration

**EMG** : Electromyography

**CT** : Computed tomography

**MRI** : Magnetic resonance imaging

**ECM** : Extracellular matrix

**VCAM-1**: Vascular cell adhesion molecule-1

**FN**: Fibronectin

**CBC** : Complete blood Count

**ELISA** : Enzyme-linked immunosorbent assay

**ROM** : Range of motion

**10MWT**: Ten Meters Walk Test

**QBA** : Quantitative backscatter analysis

**GSL** : Grayscale levels

# **List of Tables**

Table No.	Title	Page No.
<b>Table (1):</b>	Muscle power grading	85
<b>Table (2):</b>	Demographic and anthropometre patients and controls.	
<b>Table (3):</b>	Laboratory data of studied groups.	90
<b>Table (4):</b>	Descriptive statistics of complication DMD (cardiac involvement, pulm function, ten meter test) and stherapy among the studied patients	nonary steroid
<b>Table (5):</b>	Descriptive data regarding in mean value of examined musclultrasound among the patients ground	les by
<b>Table (6):</b>	Comparison between patients controls as regards demographi anthropometric data.	c and
<b>Table (7):</b>	Comparison between patients controls as regards laboratory data.	
<b>Table (8):</b>	Comparison of CD 49 according disease duration	
<b>Table (9):</b>	Comparison of CD 49 according group	
<b>Table (10):</b>	Comparison of CD 49 according meter test category	
<b>Table (11):</b>	Comparison of CD 49 according progression in non ambulant patier	_

#### List of Tables

<b>Table (12):</b>	Comparison of CD 49 according to steroid therapy	. 102
<b>Table (13):</b>	Comparison of CD 49 according to cardiac involvement	. 103
<b>Table (14):</b>	Comparison of CD 49 according to pulmonary function	. 104
Table (15):	Diagnostic performance of CD 49 for the identification of established Duchenne muscular dystrophy.	. 105
<b>Table (16):</b>	Correlation between CD49d and TLC	. 106

# **List of Figures**

Figure No	o. Title	Page No.
Figure (1):	The dystrophin-associated properties (DPC) in skeletal muscle.	
Figure (2):	The Characteristic traits of Ducher	nne 14
Figure (3):	Typical DMD disease progression.	17
Figure (4):	Muscle biopsy of DMD	24
Figure (5):	Normal dystrophin immunostain muscle	_
Figure (6):	Algorithm for the diagnosis of Ducand Becker muscular dystrophy	
Figure (7):	Example of exon skipping in Duc muscular dystrophy (DMD) patien	
Figure (8):	Summary of the wide rang approaches being used to treat Duc muscular dystrophy	chenne
Figure (9):	Interactions between ultrasound and normal and pathological tissue	
<b>Figure (10):</b>	Activated autoreactive T cells expr VLA-4 adhere to endothelium interactions with VCAM and entothe the tissue	n via er into
Figure (11):	Pie chart showing frequency category among patients group	_
<b>Figure (12):</b>	Pie chart showing frequency of duration category in patients	

Figure (13):	Pie chart showing cardiac involvement among patients group
<b>Figure (14):</b>	Pie chart showing frequency of steroid therapy among patients group92
<b>Figure (15):</b>	Pie chart showing frequency of pulmonary function among patients group
<b>Figure (16):</b>	Pie chart showing ten meter test among patients group
<b>Figure (17):</b>	Pie chart showing progression in non ambulant patients
<b>Figure (18):</b>	Mean Hb of patients and controls97
<b>Figure (19):</b>	Mean TLC of patients and controls97
<b>Figure (20):</b>	Mean CD 49 of patients and controls 98
<b>Figure (21):</b>	Mean CD 49 according to disease duration
<b>Figure (22):</b>	Mean CD 49 according to age group 100
<b>Figure (23):</b>	Mean CD 49 of ambulant and non ambulant patients
<b>Figure (24):</b>	Mean CD 49 of slow and rapid progressor in non ambulant patients 102
<b>Figure (25):</b>	Mean CD 49 according to steroid therapy
<b>Figure (26):</b>	Mean CD 49 of normal Echo and dilated cardiomyopathy patients
<b>Figure (27):</b>	Mean CD 49 according to pulmonary function

#### List of Figures

Figure (28):	Diagnostic performance of CD 49 for the identification of established Duchenne muscular dystrophy	106
Figure (29):	Correlation between CD49d and CPK	107
<b>Figure (30):</b>	Correlation between age and ten meter test of the patients	108
Figure (31):	Correlation between disease duration and ten meter test	109
<b>Figure (32):</b>	Correlation between ten meter test in ambulant patient and total muscle ultra sound	110

#### **Abstract**

**Background:** Duchenne Muscular Dystrophy (DMD) an X-linked recessive disorder affecting 1 in 3500-6000 male births. It is caused by the absence of functional dystrophin, due to mutations in the dystrophin gene. Although the cause of DMD is genetic, there is accumulating data suggesting that an immune response may play a role in pathophysiology of this disease and also contributes to disease progression in DMD patients. Alpha integrins (CD49d) can drive T cells to the site of inflammation favoring migration and adhesion to the muscle tissue and muscle damage by interacting more strongly with the fiber and extracellular matrix proteins. DMD patients are clinically heterogeneous and the functional defect cannot be correlated with genotype. Therefore, it is important to be able to define reliable noninvasive inflammatory biomarkers. Methods: A case control study was carried out at neurology outpatient clinic, Children's Hospital, Ain Shams University. The study included 20 male patient (mean age of 10.35 years), compared to 20 healthy children age, sex and socioeconomic matched served as control group. Enrolled subjects underwent peripheral blood sampling (for measuring CD49d, CBC and CPK ), motor grading and muscle ultrasound. Results: Patients had a significant higher CD 49 levels than controls. Cutoff values was 0.250 with 90% sensitivity and 70% specificity. Conclusions: Our data indicates that CD49d expression in high levels involved in the inflammatory process seen in DMD patients.

**Key words:** blood CD49d, muscle ultrasonography, children, Duchenne Muscular Dystrophy

#### Introduction

uchenne Muscular Dystrophy (DMD) is an X-linked recessive disease of muscle characterized by a progressive loss of functional muscle mass and replacement with fibrofatty tissue (*Sussman and Michael*, 2002).

The incidence of DMD is approximately 1 in 3500 live male births (Gulati et al., 2005). It is caused by mutations in the DMD gene. This gene encodes a protein called dystrophin (Blake et al., 2002), which localizes to the cytoplasmic face of the sarcolemma (plasma membrane) of the skeletal muscle (Ervasti, 2007), forming one component of a large glycoprotein complex (dystrophin-associated glycoprotein complex). The majority of mutations are intragenic deletions, which account for 65-72% of all DMD patients (Aartsma-Rus et al., 2006). Point mutations, small deletions or insertions account for 20% of duplications patients without deletions or (Prior and Bridgeman, 2005).

The precise mechanism of how dystrophin deficiency leads to degeneration of muscle fibers remains unclear. Absence of dystrophin at the plasma membrane leads to delocalisation of dystrophin-associated proteins from the membrane, disruption of the cytoskeleton with resultant membrane instability and increased susceptibility to mechanical stress (*Deconinck et al.*, 2007).

Duchenne muscular dystrophy is the most common fatal genetic disorder diagnosed in childhood, and it does shorten life. Because the muscle weakness increases gradually over the years, complications eventually develop. The breathing or heart problems usually become more serious for older teenagers or people in their 20s. In the past, most people with DMD did not live beyond their early 20s. Improvements in treatment have meant that life expectancy has increased. At present, average life expectancy for people with DMD is 27 years. However, there is a lot of individual variation in the severity of DMD and the individual life expectancy (*Kliegman et al.*, 2011).

There is no cure for DMD at this time, but there continues to be a tremendous amount of research taking place across the world (*Jefferies et al.*, 2005). Treatment aims to control symptoms to improve quality of life. Steroid drugs can slow the loss of muscle strength. They may be started when the child is diagnosed or when muscle strength begins to decline. Physical therapy is used to promote mobility and prevent contractures (*Finder et al.*, 2004).

Ultrasound imaging has proved to be a useful, non invasive screening tool in the investigation of children with neuromuscular disease. Muscular dystrophies were associated with an increase in the intensity of echo reflected from the muscle substance, with corresponding loss of bone echo. Severity of changes on the scan did not relate to

functional disability, and some children had good function yet strikingly abnormal scan. the degree on the scan correlated with the degree of disruption of muscle architecture on biopsy (*Wattjes et al.*, 2010). Muscle ultrasound (MUS) is potentially an attractive follow up tool for DMD because it reflects the severity of the dystrophic process without the need for invasive procedures, by quantifying echo intensity (i.e. mean grey level of muscle images) and muscle thickness (*Jansen et al.*, 2012).

DMD patients are clinically heterogeneous and the functional defect cannot be correlated with genotype. Therefore, it is important to be able to define reliable noninvasive biomarkers to better define the disease progression (*Pinto-Mariz et al., 2015*). The non-invasive biomarkers obtained at early stages of the disease are found to be highly predictive for the loss of ambulation before 6 months of age. An elevation in the number of circulating CD49dhi T cells is found to be strongly associated with the severe clinical form of the disease. This factor can be used as predictive tests for screaning to separate them into groups with slow or fast disease progression before their inclusion into a therapy (*Barthélémy et al., 2014*).

Moreover, anti-CD49d peptides or antibodies can be used as a therapeutic approach to decrease inflammation-mediated tissue damage in DMD.