

Department of Virology

Isolation, Molecular Characterization and Pathogenicity of IBDV associated with Mortalities in Broiler Chickens

A Thesis presented by

Reem Ahmed Said Ahmed Soliman

(B. V.Sc, Cairo University, 2005)

(MSc.D., Cairo University, 2010)

For PhD Degree

In Veterinary Medical Science. (Virology)

Under Supervision of

Prof. Dr. Hussein Aly Hussein Ahmed

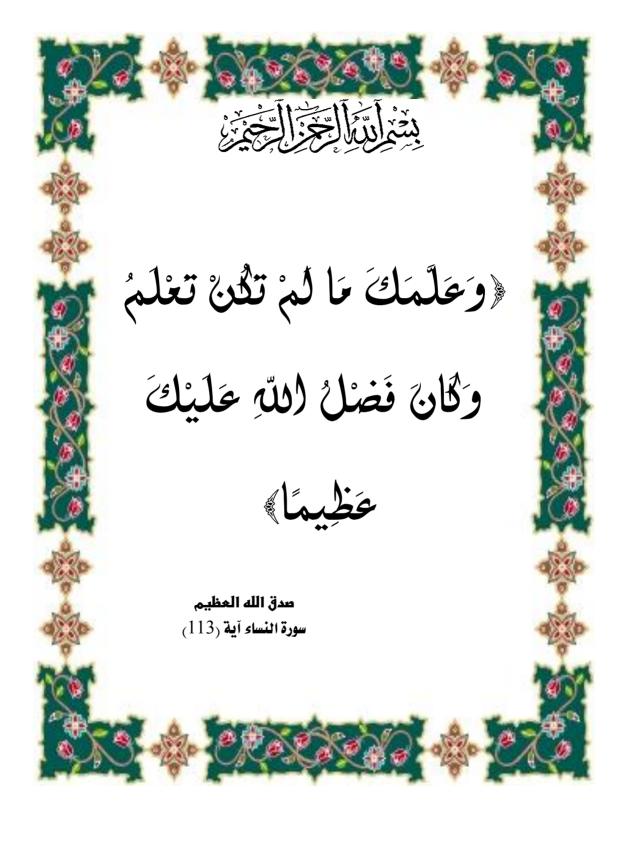
Professor of Virology Department
Faculty of Veterinary Medicine
Cairo University, Giza

Prof. Dr. Ismaiel Mohamed Reda

Professor of Virology
Faculty of Veterinary Medicine
Cairo University

Prof. Dr. Mounir Mohamed El-Safty

Chief Researcher, Central
Laboratory for Evaluation of Vet.
Biol., Director of national project for
production of SPF Eggs
2017



Cairo University Faculty of Veterinary Medicine Department of Virology

Name: Reem Ahmed Said Ahmed Soliman Nationality: Egyptian

Date and place of birth: 1/12/1983, Saudi Arabia Speciality: Virology

Supervisors: Prof. Dr. Hussein Aly Hussein Ahmed

Prof. Dr. Ismail Mohamed Reda

Prof. Dr. Mounir Mohamed El-Safty

Abstract

The present study was performed to detect, isolate and identify the causative agent associated with mortalities in broiler flocks in Beheira Governorate in EGYPT. Field samples were collected from broiler chickens demonstrating mortalities. The samples were inoculated in one-day old SPF chicks via intranasal and intraocular route. Chicks were kept in isolators with daily observation for five weeks post infection. Blood samples were collected and tested for infectious bursal disease virus (IBDV), chicken anemia virus (CAV), infectious bronchitis virus (IBV), Newcastle disease virus (NDV) and Avian influenza H5 and H9 subtypes by ELISA and HI tests. The results indicated that the samples were positive for IBDV. Histopathological examination of the collected bursae and kidneys, four and seven days post inoculation (DPI) revealed characteristic lesions for IBDV including inter-follicular connective tissue proliferation, compressed follicles, depleted lymphocytes in the bursae, congested blood vessels and hemorrhage in the kidneys. Conventional Reverse transcriptase-polymerase chain reaction (RT-PCR) and Real-time RT-PCR (rtRT-PCR) assays for VP2 gene of IBDV confirmed the presence of IBDV. Sequence analysis of amplified PCR product indicated continuous circulation of virulent IBDV strains in Egypt and it was closely related to previously isolated very virulent strain Giza 2008. Respective nucleotide sequence was submitted to NCBI GenBank with the access number KY200662. The pathogenic effect of the isolated (Egypt/ IBDV/Behera 2011) strain was studied. One-day old SPF chicks were inoculated by intra ocular and intra nasal route with the bursal homogenate of the infected farm (10³ID₅₀/ml). At 3rd, 7th, 10th, 14th, 21th and 28th days post inoculation (DPI), 10 chicks were randomly selected, weighted, bled, humanely euthanized and then necropsied. Bursa, spleen, and kidney were taken out and weighted. Isolated IBD strain Pathogenicity was evaluated by mortality percent, bursa, spleen, kidney /body weight ratio. Five bursae were collected at 24 hours interval up to 96 hours, then at 7th and 10th DPI. Collected bursae were either fixed in 10% formol saline or in a mixture of 2.5% glutaraldehyde for histopathology and transmission electron microscope (TEM) respectively. Histopathological lesions were scored from 0 to 4 based on the presence of degenerative alterations, necrosis, follicular atrophy in the bursa of Fabricius and the percentage of altered lymphoid follicles. IBD antigen demonstrated by Agar gel diffusion test (AGDT), Antibodies against IBDV were measured by Enzyme-linked Immunosorbent assay (ELISA). Twenty infected chicks died (25%), mortalities started 3 DPI and ceased 7 DPI with the highest mortality of 3 chicks at 5 DPI (7.5%) compared with negative control which showed no deaths at all over experimental time. The highest bursa/body weight ratio was observed at 3rd, 7th, 10th DPI then decreased at 14th, 21th, 28th DPI; respectively. Histopathologically, severe tissue reaction in the bursa of Fabricius of inoculated chicks, and the degree of damage varies including hyperemia, hemorrhage and edema, degeneration, necrosis. The Mean Severity (MSI) ranged from 0.5 to 4. IBD viral particles were visualized by Transmission Electron microscope (TEM). Agar gel diffusion test (AGDT) confirmed the presence of IBDV. ELISA were positive for IBDV antibodies from 7 DPI, increased at 21 DPI then declined at 28 DPI. Indeed the study reports the isolation, molecular characterization and pathogenicity of IBDV strain (Egypt/ IBDV/Behera 2011).

Key Words: Infectious Bursal Disease Virus (IBDV); Vp2 Gene Hypervariable Region; Real-time RT-PCR; SPF chickens; immunosuppression; pathogenicity.

DEDICATION

To my beloved Father whom I carry his name proudly,

My great Mother the secret of my life and success,

My twin soul Hatem for his encouragement and support,

My husband Mostafa for faithful efforts & encouragement

My lovely children JANA LASSER, I see optimism in their eyes Lhappiness in their laughter.

ACKNOWLEDGMENT

First and always, the prayerful thanks to our **Superb Bestower** (Al-wahhab) **ALLAH** who gives me the powerful to begins and complete this thesis by his guidance and care. He gives me everything I have and I need.

I would like to express all my respects to my supervisor **Prof. Dr. Ismail Mohamed Reda** – Professor of Virology – Faculty of Veterinary Medicine, Cairo University, and God bless his soul and makes the rest of Paradise.

I would like to express my deep thanks and gratitude to **Prof. Dr. Hussein Aly Hussein Ahmed**-Professor of Virology Department, Faculty of Veterinary Medicine, Cairo University for his stimulating supervision, guidance, great effort, useful discussions and support in every step of my work. I heartily thank him very much for his valuable helps and advice.

I would like to express my deep thanks and respect to my general manager **Prof. Dr.**Mounir Mohamed El-Safty (Chief Researcher, Head of SPF Eggs Dep. In CLEVB and Director of National Project for Production of SPF Eggs) for his kind and valuable supervision, advice, experience, teaching me the practical works, continuous encouragement, and the facilitation he offered during carrying out this thesis.

Best regards and thanks to **Prof Dr. Laiyla Tantawy** for her help throughout this study.

Best regards and thanks to **Dr. Bassem Abd El-hamid** for his help and support throughout this study.

And very great thanks to **Dr. Hala Mahmoud Attya** (Researcher and head of unit of Quality Control of SPF Eggs Dep. in CLEVB) for her attributes and great help during my work and in writing the thesis.

I would like to express my deep thanks to **Dr. Ahmed Maher** (Researcher in CLEVB) for his great helps in my work.

I would like to express my best regards and great thanks to **Dr. Marwa Fathy** for her support and encouragement in my work, writing the thesis and every things in my life.

I wish to thank **Dr. Wahid Hussein** for his help throughout this study.

At last but not least I would like to thank all my colleagues and personnel by their names (Doctors, Technicians and workers) of CLEVB, who give a hand whenever needed.

List of abbreviations

AGPT	Agar gel precipitation test
bp	Base pair
BF	Bursa of Fabricius
CMI	Cell-mediated immunity
CLVB	Central Laboratory for Evaluation of Veterinary Biologics
CIAV	chicken infectious anemia virus
CAM	Chorioallantoic membrane
DPI	day post infection
DNA	Deoxy-ribonucleic acid
EU	ELISA unit
ECE	Embryonated chicken egg
ELISA	Enzyme linked immunosorbant assay
GALT	Gut-associated lymphoid tissues
HI	Heamagglutination inhibition
НА	Hemagglutination
HVR	Hyper Variable Region
IBA	Infectious bursal agent
IBDV	Infectious bursal disease virus

microliter
Newcastle disease virus
Nitric oxide
Office of international Epizootics
open reading frame
Phosphate buffer saline
Polymerase chain reaction
Postmortem
Power of hydrogen
Real time Reverse transcriptase polymerase chain reaction
Red Blood Corpuscles
Reverse transcriptase polymerase chain reaction
ribonucleic Acid
Specific pathogen free
surface immunoglobulin M
Transmission Electron microscope
virus neutralization test
Volume
Weight

CONTENTS

Sr.	Content	Page No.
	INTRODUCTION	1
	REVIEW OF LITRATURE	4
2.1.	History of Infectious Bursal Disease Virus	4
2.1.1	History of IBDV in the world	4
2.1.2.	History of IBD in Egypt	6
2.2.	Etiology of IBDV	11
2.3.	Resistance to physical and chemical agents	12
2.4.	Virus structure and chemical Composition	13
2.4.1.	IBDV viral proteins	15
2.5.	Pathogenesis and Virus Replication of IBDV	17
2.6.	Transmission of IBDV	25
2.7.	Incubation Period, Clinical Signs, and Mortality	26
2.8.	Gross Lesions	28
2.9.	Histopathological and microscopic changes	30
2.10	Bursa, spleen and kidneys / body weight ratios	31
2.11.	Antigenic Variation of IBDV Strains	32
2.12.	Isolation of IBDV	35
2.12.1.	Embryonated chicken eggs	35
2.12.2.	Isolation of IBDV on tissue culture	36
2.13.	Serological tests	36
2.13.1.	Agar gel precipitation test(AGPT)	36
2.13.2.	Enzyme Linked Immuno Sorbant Assay (ELISA)	39
2.14.	Detection of IBDV nucleic acid using molecular techniques.	41
2.14.1.	Reverse Transcription Polymerase Chain Reaction (RT-PCR)	41
2.15.	Differential Diagnosis of IBDV	44
	MATERIALS AND METHODS	46

3.1.	Materials	46
3.1.1	Experiment number No. 1 : (Materials used in trails for detection, isolation and identification of the causative agent)	46
3.1.1.1.	Materials used in Enzyme Linked ImmunoSorbant Assay kits (ELISA)	47
3.1.1.2.	Materials used in Haemagglutination inhibition (HI) test	49
3.1.1.3.	Materials used in histopathology	50
3.1.1.4.	Materials used in real time RT- PCR (rt RT –PCR) for IBD	51
3.1.1.5.	Materials used in Conventional Reverse Transcription- Polymerase Chain (RT-PCR)	52
3.1.1.6.	Materials used in IBD Sequence analysis and phylogenic tree	54
3.1.2.	Materials used in (Experiment number 2): Studying of Pathogenicity of IBDV (Egypt /IBDV/ Behera 2011) strain	55
3.1.2.1.	Materials used for record of organs to body weight index	56
3.1.2.2	Estimation of bursal score lesions, mean severity index and histopathological study	56
3.1.2.3.	Materials for Transmission Electron microscope (TEM)	56
3.1.2.4.	Materials used in Agar gel precipitation test (AGPT)	57
3.1.2.5.	Materials for Detection of antibodies against the IBD virus by Enzyme-linked ImmunoSorbant assay (ELISA)	57
3.2.	Methods	59
3.2.1.	Experiment number. 1: (Trails for detection, isolation and identification of the causative agent)	59
3.2.1.1.	Enzyme Linked ImmunoSorbant Assay (ELISA)	60
3.2.1.2.	Haemagglutination inhibition (HI) test	66
3.2.1.3.	Histopathological examination	68
3.2.1.4.	Molecular characterization of the isolated virus by rt RT-PCR	68
3.2.1.5.	Conventional Reverse Transcription- Polymerase Chain Reaction (RT-PCR)	72

3.2.1.6.	Methods of IBD Sequence analysis and phylogenic tree	74
3.2.1.7.	Submission in GenBank	78
3.2.2.	Methods used in Experiment number 2: Studying of Pathogenicity of IBDV (Egypt /IBDV/ Behera 2011) strain	79
3.2.2.1.	Methods of determination of organs (bursa /spleen / kidney) to body weight index	81
3.2.2.2.	Estimation of bursal score lesions, mean severity index and histopathological study	81
3.2.2.3.	Methods of Transmission Electron microscope (TEM)	82
3.2.2.4.	Methods of Agar gel precipitation test (AGPT)	83
3.2.2.5.	Detection of antibodies against the IBD virus by Enzyme-linked ImmunoSorbant assay (ELISA)	83
	RESULTS	86
4.1.	Experiment (1): Results of detection, isolation and identification of the causative agent	86
4.1.1.	Results of Enzyme Linked ImmunoSorbant Assay (ELISA)	86
4.1.2.	Results for Haemagglutination inhibition (HI) test	87
4.1.3.	Histopathological examination of bursa of Fabricius and kidney collected from inoculated chicks 4 th and 7 th PI	89
4.1.4.	Results of Molecular characterization of the isolated virus by real time PCR(rt RT-PCR) in bursal samples	91
4.1.5.	Results of molecular characterization of the isolated virus by Conventional Reverse Transcription-Polymerase Chain Reaction (RT-PCR)	92
4.1.6.	Results of sequence analysis of VP2 gene of the local isolated IBDV	94
4.1.6.1.	Nucleotide sequence identity	94
4.1.6.2.	Amino acid sequence identity	97
4.2.	Results of Experiment number (2): Results of Propagation of the virus In Embryonated Chicken Eggs	101
4.2.1.	Results of virus infectivity titration	102
4.2.2.	Results of Mortality percent	102
4.2.3.	organs to body weight index	102
4.2.4.	The result of histopathological study of bursa of	105