

**Effect of Intrauterine Injection of Human  
Chorionic Gonadotropin at the Day of Ovum  
Pick-Up on Patients Undergoing ICSI for  
Recurrent Implantation Failure**

*Thesis*

Submitted for partial fulfillment of Master Degree  
in Obstetrics and Gynecology

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**2016**

بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

قالوا

سبناك لا علم لنا  
إلا ما علمتنا إنك أنت  
العليم الحكيم

صدق الله العظيم

سورة البقرة الآية: ٣٢



## Acknowledgment

First of all, I thank Almighty *Allah* for every blessing in my life, starting from the day I was born till this moment.

I would like to express my deepest gratitude to **Prof. Rowaa Abd El Azeem Mostafa**, professor of obstetrics and gynecology, faculty of medicine, Ain Shams University, for giving me privilege of working under his supervision and his helpful instructive guidance.

I feel so deeply indebted to **Prof. Mohamed Sayed Ali**, professor of obstetrics and gynecology, faculty of medicine, Ain Shams University, for his unlimited help, his continuous support and his exceptional attitude in encouraging me every step of the way.

I would like to express my deep thanks to **Dr. Hayam Fathy Mohammed**, Lecturer of Obstetrics and Gynecology, faculty of medicine, Ain Shams University, for her kind advices, constant and meticulous supervision and revision of the study.

I also wish to thank **Dr. Azza Awaad Abd El Razik**, director of embryology lab at IVF unit, Ain Shams University, for her great help in laboratory work.

Finally, I would like to thank my family “namely my **Mother**” and my friends for their indefinite prayers and support all through my way.



Shaimaa Shabaan

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## **List of Abbreviations**

<b>AFC</b>	: Antral follicle count
<b>ART</b>	: Assisted reproductive technology
<b>ASRM</b>	: American society for reproductive medicine
<b>E<sub>2</sub></b>	: Estradiol
<b>ESHRE</b>	: European society of human reproduction and embryology
<b>ET</b>	: Embryo transfer
<b>FET</b>	: Frozen embryo transfers
<b>FSH</b>	: Follicle stimulating hormone
<b>G-CSF</b>	: Granulocyte colony stimulating factor
<b>GnRH</b>	: Gonadotropin releasing hormone
<b>hCG</b>	: Human chorionic gonadotropin
<b>HLA</b>	: Human leukocyte antigen
<b>HMG</b>	: Human menopausal gonadotropin
<b>ICSI</b>	: Intracytoplasmic sperm injection
<b>IGFBP-1</b>	: Insulin like growth factor binding protein-1
<b>IL-1 R</b>	: Interleukin-1 receptor
<b>IMSI</b>	: Intracytoplasmic morphologically selected sperm injection
<b>IUI</b>	: Intrauterine insemination
<b>IVF</b>	: In vitro fertilization
<b>LH</b>	: Luteinizing hormone
<b>LIF</b>	: Leukemia inhibitory factor
<b>LMP</b>	: Last menstrual period
<b>M II</b>	: Metaphase II
<b>M-CSF</b>	: Macrophage colony stimulating factor
<b>MMP-9</b>	: Matrix metalloproteinase-9
<b>NK</b>	: Natural killer

## **List of Abbreviations** *(Cont.)*

<b>OPU</b>	: Ovum pick-up
<b>PB</b>	: Polar body
<b>PGD</b>	: Preimplantation genetic diagnosis
<b>PGS</b>	: Preimplantation genetic screening
<b>PN</b>	: Pronucleus
<b>PPARs</b>	: Peroxisome proliferator activated receptors
<b>PVP</b>	: Polyvinylirrolidone
<b>rFSH</b>	: Recombinant follicle stimulating hormone
<b>r-hLIF</b>	: Recombinant human leukemia inhibitory factor
<b>RIF</b>	: Recurrent implantation failure
<b>TH</b>	: T-helper cells
<b>TSH</b>	: Thyroid stimulating hormone
<b>TVUS</b>	: Trans-vaginal ultrasound
<b>VEGF</b>	: Vascular endothelial growth factor
<b>ZP</b>	: Zona pellucida

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## ABSTRACT

**Background:** Implantation is a complex initial step in the establishment of a successful pregnancy. Although embryo quality is an important determinant of implantation, temporally coordinated differentiation of endometrial cells to attain uterine receptivity and a synchronized dialog between maternal and embryonic tissues are crucial. **Aim:** The aim of the present study is to investigate the value of intrauterine injection of hCG at day of ovum pick-up in patients undergoing intracytoplasmic sperm injection in an attempt to improve implantation and pregnancy rates on patients with recurrent implantation failure. **Patients and Methods:** The recent study was designed as a prospective case-control, randomized, single blinded study, from February 2015 to January 2016, conducted at ART unit of Ain Shams University, maternity hospital. One hundred participants were assigned into the study, who were prepared to undergo an ICSI trial after a failed previous cycle or more. Patients were divided in to two groups: Group (1): The study group; included 50 patients who were injected by 500 IU of hCG intrauterine at the day of ovum pick-up. Group (2): The control group; included 50 patients, age matched, who went through the study without intrauterine injection. **Results:** The current study revealed that: **Conclusion:** it was found that this approach significantly improved pregnancy and implantation rates, with even improvement of the endometrial thickness at the day of embryo transfer; however miscarriage rates were non-significant between both groups.

**Key words:** Human chorionic gonadotropin, Ovum pick-up,  
Recurrent Implantation Failure

## Introduction

Implantation is a complex initial step in the establishment of a successful pregnancy. Although embryo quality is an important determinant of implantation, temporally coordinated differentiation of endometrial cells to attain uterine receptivity and a synchronized dialog between maternal and embryonic tissues are crucial. The exact mechanism of implantation failure is still poorly understood (*Cakman and Taylor, 2010*).

Successful implantation after in vitro fertilization (IVF) and embryo transfer (ET) depends on various factors related to the embryo quality and the endometrial receptivity. It is important that the embryo reaches the endometrial cavity during the period in which the endometrium is receptive, known as the “implantation window” (*Psychoyos, 1986*).

It is estimated that about 50% to 75% of lost pregnancies are due to failure of implantation (*Norwitz et al., 2001*).

Major advances have occurred in clinical and laboratory IVF techniques in recent years; however, the pregnancy rates remain around 30% per cycle (*de Mouzon et al., 2002*).

Implantation is a very intricate process that is regulated by many factors, one of the most important of which is human chorionic gonadotropin (hCG) (*Tsampalas et al., 2010*).

Human chorionic gonadotropin (hCG) is a heterodimeric placental glycoprotein hormone that is required to maintain pregnancy. The hCG is initially produced by the blastocyst 6–8 days after fertilization (*Lopata and Hay, 1989*).

It is one of the early embryonic signals in primates that is secreted by the embryo before its implantation (*Bonduelle et al., 1988*).

Different mechanisms have been described in which hCG can regulate implantation. An in vitro study demonstrated that hCG is a potent attractor of inflammatory cells, such as neutrophils, monocytes, and lymphocytes (*Reinisch et al., 1994*).

The hCG directly regulates endothelial cell responsiveness to interleukin 1 and amplifies the cytokine-mediated effect on cell proliferation, migration and release of angiogenic factors (*Bourdiec et al., 2013*).

Embryo implantation requires an extensive angiogenesis at the maternal-fetal interface. The hCG can modulate the receptivity of the endometrial stromal cells to interleukin-1 by upregulating its receptor (IL1R) during the implantation window. This function has an impact on angiogenesis, which is a pathway by which embryonic growth is promoted (*Bourdiec et al., 2012*).

Licht et al. developed an intrauterine microdialysis device to measure paracrine mediators. After the administration of 500 UI of hCG, they found a significant inhibition of intrauterine insulin-like growth factor binding protein 1 (IGFBP-1) and the macrophage colony-stimulating factor (M-CSF), while leukemia inhibitory factor (LIF), the vascular endothelial growth factor (VEGF) and the matrix metalloproteinase 9 (MMP-9) were significantly stimulated. These multiple effects appear to precede the classical endocrine role of the hCG and could be directly involved in the regulation of embryo implantation (*Licht et al., 2007*).

Mansour et al. reported the first time use of an intrauterine injection of hCG before the embryo transfers, and they found a significant improvement in the pregnancy rates of the IVF cycles (*Mansour et al., 2011*).

Another study was performed by Santibanez et al. which aimed to reproduce and confirm the benefits to the clinical effects of intrauterine injection of hCG before embryo transfer on the pregnancy rates in the IVF cycles (*Santibanez et al., 2014*).

## **Aim of the Work**

**T**he aim of the present study is to investigate the value of intrauterine injection of hCG at day of ovum pick-up in patients undergoing intracytoplasmic sperm injection in an attempt to improve implantation and pregnancy rates on patients with recurrent implantation failure.

## Human Chorionic Gonadotropin

**H**uman chorionic gonadotropin (hCG) is a hormone produced by the syncytiotrophoblast, a portion of the placenta following implantation (*Gregory and Finlay, 1999 and Cole, 2009*).

The presence of hCG is detected in pregnancy tests. Some cancerous tumors produce this hormone; therefore, elevated levels measured when the patient is not pregnant can lead to a cancer diagnosis. However, it is not known whether this production is a contributing cause or an effect of tumorigenesis (*Cole, 2009*).

The pituitary analog of hCG, known as luteinizing hormone (LH), is produced in the pituitary gland of males and females of all ages (*Cole 2009, and Hoermann et al., 1990*).

Both hCG and LH bind and function through a common hCG/LH receptor. The biggest difference between LH and hCG is that LH, has a circulating half-life of just 25-30 minutes (*Parlow et al., 1968*), while hCG, has a circulating half-life of approximately 37 hours (*Faiman et al., 1968*), or 80-fold longer than that of LH.

### Structure:

Human chorionic gonadotropin is a glycoprotein composed of 237 amino acids with a molecular mass of 25.7 kDa (*Lapthorn et al., 1994*).



**Figure (1):** Chemical formula of hCG

It is heterodimeric, with an  $\alpha$  (alpha) subunit identical to that of luteinizing hormone (LH), follicle stimulating hormone (FSH), thyroid stimulating hormone (TSH), and  $\beta$  (beta) subunit that is unique to hCG.

- The  $\alpha$  (alpha) subunit is 92 amino acids long.
- The  $\beta$ -subunit of hCG gonadotropin (**beta-hCG**) contains 145 amino acids, encoded by six highly homologous genes that are arranged in tandem and inverted pairs on chromosome **19q13.3 -CGB** (1, 2, 3, 5, 7, 8)

The two subunits create a small hydrophobic core surrounded by a high surface area-to-volume ratio 2:8 times that of a sphere. The vast majority of the outer amino acids are hydrophilic (*Lapthorn et al., 1994*).

Research shows that there are at least 4 independent variants of hCG, each produced by different cells with separate biological functions. All the molecules share a common hCG $\beta$ -subunit amino acid sequence (*Lapthorn et al., 1994*).

There is hCG, produced by differentiated syncytiotrophoblast cells or more specifically villous syncytiotrophoblast