

**EARLY DIAGNOSIS OF PLANT DISEASES
CAUSED BY *Ganoderma* spp.**

By

LABIBA AHMED REDA

B.Sc. Agric. Sc. (Plant pathology), Ain Shams University, 2001

M.Sc. Agric. Sc. (Plant pathology), Ain Shams University, 2008

**A thesis submitted in partial fulfillment
of
the requirements for the degree of**

DOCTOR OF PHILOSOPHY

in

**Agricultural Science
(Plant Pathology)**

**Department of Plant Pathology
Faculty of Agriculture
Ain Shams University**

2013

Approval Sheet

EARLY DIAGNOSIS OF PLANT DISEASES CAUSED BY *Ganoderma* spp.

By

LABIBA AHMED REDA

B.Sc. Agric. Sc. (Plant pathology), Ain Shams University, 2001

M.Sc. Agric. Sc. (Plant pathology), Ain Shams University, 2008

This thesis for Ph.D. degree has been approved by:

Dr. Mohamed Anwar Abd-Elsattar

Prof. Emeritus of Plant Pathology, Faculty of Agriculture, Suez
Canal University

Dr. Ibrahim Sadek Elewa

Prof. Emeritus of Plant Pathology, Faculty of Agriculture, Ain
Shams University

Dr. Mostafa Helmy Mostafa

Prof. Emeritus of Plant Pathology, Faculty of Agriculture, Ain
Shams University

Dr. Mohamed Aly Ahmed

Prof. Emeritus of Plant Pathology, Faculty of Agriculture, Ain
Shams University

Date of Examination: 22 / 9 / 2013

EARLY DIAGNOSIS OF PLANT DISEASES CAUSED BY *Ganoderma* spp.

By

LABIBA AHMED REDA

B.Sc. Agric. Sc. (Plant pathology), Ain Shams University, 2001

M.Sc. Agric. Sc. (Plant pathology), Ain Shams University, 2008

Under the supervision of:

Dr. Mohamed Aly Ahmed

Prof. Emeritus of Plant Pathology, Department of Plant Pathology, Faculty of Agriculture, Ain Shams University
(Principal supervisor)

Dr. Mostafa Helmy Mostafa

Prof. Emeritus of Plant Pathology, Department of Plant Pathology, Faculty of Agriculture, Ain Shams University

Dr. Magdy Gad-Elrab Mohamed El - Samman

Prof. of Plant Pathology, Department of Plant Pathology, Faculty of Agriculture, Ain Shams University

التشخيص المبكر للأمراض النباتية المتسببه عن أنواع
Ganoderma فطر

رسالة مقدمة من

لبيبة أحمد رضا

بكالوريوس علوم زراعية (أمراض النبات)، جامعة عين شمس، 2001
ماجستير علوم زراعية (أمراض النبات)، جامعة عين شمس، 2008

للحصول على

درجة دكتور فلسفة في العلوم الزراعية
(أمراض النبات)

قسم أمراض النبات
كلية الزراعة
جامعة عين شمس

صفحة الموافقة على الرسالة

التشخيص المبكر للأمراض النباتية المتسببه عن أنواع

Ganoderma فطر

رسالة مقدمة من

لبيبة أحمد رضا

بكالوريوس علوم زراعية (أمراض النبات)، جامعة عين شمس، 2001
ماجستير علوم زراعية (أمراض النبات)، جامعة عين شمس، 2008

للحصول على
درجة دكتور فلسفة في العلوم الزراعية
(أمراض النبات)

وقد تمت مناقشة الرسالة والموافقة عليها
الجنة :

د. محمد أنور عبد الستار

أستاذ أمراض النبات المتفرغ ، كلية الزراعة، جامعة قناة السويس

د. إبراهيم صادق عليوة

أستاذ أمراض النبات المتفرغ ، كلية الزراعة، جامعة عين شمس

د. مصطفى حلمي مصطفى

أستاذ أمراض النبات المتفرغ ، كلية الزراعة، جامعة عين شمس

د. محمد علي أحمد

أستاذ أمراض النبات المتفرغ ، كلية الزراعة، جامعة عين شمس

تاريخ المناقشة: 22 / 9 / 2013

جامعة عين شمس
كلية الزراعة

رسالة دكتوراه

اسم الطالب : لبيبة أحمد رضا
عنوان الرسالة : التشخيص المبكر للأمراض النباتية المتسببه عن
أنواع فطر *Ganoderma*
اسم الدرجة : دكتور فلسفة في العلوم الزراعية (أمراض النبات)

لجنة الإشراف:

د. محمد علي أحمد

أستاذ أمراض النبات المتفرغ ، قسم أمراض النبات ، كلية الزراعة ، جامعة عين شمس
(المشرف الرئيسي).

د. مصطفى حلمي مصطفى

أستاذ أمراض النبات المتفرغ ، قسم أمراض النبات ، كلية الزراعة ، جامعة عين شمس

د. مجدي جاد الرب محمد السمان

أستاذ أمراض النبات ، قسم أمراض النبات ، كلية الزراعة ، جامعة عين شمس

تاريخ التسجيل: 13 / 10 / 2008

الدراسات العليا

أجيزت الرسالة بتاريخ

2013 / 9 / 22

موافقة مجلس الجامعة

2013 / /

ختم الإجازة

موافقة مجلس الكلية

2013 / /

ABSTRACT

Labiba Ahmed Reda: Early Diagnosis of Plant Diseases Caused by *Ganoderma* spp. Unpublished Ph.D. Thesis, Department of Plant Pathology, Faculty of Agriculture, Ain Shams University, 2013.

Fifteen *Ganoderma* isolates were collected from five naturally diseased host plants i.e. navel orange, morus, casuarina, date palm and fan palm from two governorates in Egypt i.e. Kalyubia and Giza.

Morphological observation, cultural characteristics and chemical test revealed that all fifteen isolates of *Ganoderma* associated with *Ganoderma resinaceum* and *Ganoderma lucidum*.

Phylogenetic analysis of ITS1 sequences revealed that the selective five isolates (1, 4, 7, 10, 13) belong to a single species i.e. *Ganoderma resinaceum*.

Pathogenicity test of five *Ganoderma* isolates, obtained from naturally diseased navel orange, morus, casuarina, date palm and fan palm plants was carried out under greenhouse conditions. Pathogenicity test revealed that all fungal isolates were pathogenic to their host plants. These isolates caused root-rot symptoms, suddenly wilt without any yellowing symptoms, dieback of twigs and branches on artificially inoculated navel orange grafted on sour orange rootstocks. Visible symptoms were distinguished 12 months after inoculation of young trees.

The same symptoms were observed on navel orange grafted on volkamer lemon rootstocks artificially inoculated with two *Ganoderma* spp. (isolate1 and 13). Meanwhile root-rot symptoms, a slight distinct yellowing and desiccation of the lower leaves were showed on fan palm plants artificially inoculated with *Ganoderma* spp. (isolate1 and 13) 6 months after inoculation.

Polymerase chain reactions (PCR) were used for the confirmation of pathogenicity in inoculated plants using primers Gan1 and Gan2 generated from internal transcribed spacer region of rDNA. Primers Gan1 and Gan2 amplified a DNA fragment of the size of 167 bp when DNA of *Ganoderma* isolates were used for amplification. The amplification of a 167-bp PCR product was achieved with inoculated roots of sour orange rootstocks and volkamer lemon rootstocks 6 months after inoculation with tested *Ganoderma* isolates before disease symptoms had been expressed. Meanwhile the amplification of a 167-bp PCR product was achieved with inoculated roots of fan palm plants 5 months after inoculation with *Ganoderma* isolates before disease symptoms had been expressed. No PCR signal of *Ganoderma* DNA was observed in non-inoculated (healthy) roots. The DNA extract could be diluted up to 1:10 to get a positive reaction.

Physiological studies revealed that the tested fungal isolates produced cellulase, pectin degrading enzymes either in cultural filtrates or of plant tissues.

Data showed an ability of fungal isolates for degradation of lignin on wood chips.

Severe degradation of lignin was detected in young trees of navel orange grafted on sour orange and navel orange grafted on volkamer lemon artificially inoculated with tested fungal isolates. Degradation of lignin in inoculated fan palm seedlings was not significant.

Results showed that no toxins were found in *G. resinaceum* liquid medium.

Results indicated that the temperature of incubation has greeted effect of sour orange root pieces colonization by both *G. resinaceum* isolates. At 15 °C, rate of colonization of sour orange by both tested isolates was very poor. The fungus attracted to wound area indicated

chemotaxis effect of root pieces on fungal growth. It could be conclude that 20-30 °C is the optimum for colonization of root pieces by both isolates. Isolate No. 13 tend to be more aggressive at 35 °C, and isolate No. 1 was more aggressive at 25 °C.

Scanning electron microscopy revealed that both isolates had great effect on phloem elements and xylem vessels. Fungal hyphae were shown on xylem vessels at different temperature degrees of incubation. Moreover, tylosis was evident in many cases.

Key words: *Ganoderma resinaceum*, pathogenicity, isolation, early diagnosis, PCR, internal transcribed spacer, ITS1, Isolation, navel orange, morus, casuarina, date palm, fan palm, root- rot, taxonomy, Morphological characteristics, cultural characteristics, Laccate fungi, basidiospores, chlamydospores, hyphal system, cutis, Phylogeny, Wood-decay fungi, ligninolytic enzymes, pectintransesterase, cellulolytic enzyme, cellulase, toxin, Scanning electron microscopy.

ACKNOWLEDGEMENT

I wish to express my deep thanks to **ALLAH** who fulfilled my hopes and promise to offer every possible aid for any one in need to it.

I am deeply indebted to **Prof. Dr. Mohamed Aly Ahmed**, Prof. of Plant Pathology, Faculty of Agric., Ain Shams Univ., for suggesting the research work, kind supervision, his encouragement, valuable advice and guidance during this work and the preparation of this manuscript.

I am particularly grateful to **Prof. Dr. Mostafa Helmy Mostafa**, Prof. of Plant Pathology, Faculty of Agric., Ain Shams Univ., for suggesting the current study, his kind supervision, assistance sincere advice and guidance during this work and the preparation of this manuscript.

I would like to thank **Prof. Dr. Magdy Gad-Elrab Mohamed El – Samman** , Prof. of Plant Pathology, Faculty of Agric., Ain Shams Univ., for his supervision

I would like to thank **Dr. Naglaa M. Ebeed**, Associate Prof. of genetics, Faculty of Agric., Ain Shams Univ., for suggesting the current study, her kind supervision, assistance sincere advice and guidance during this work and the preparation of this manuscript.

Special thanks to all staff members of genetics Dept. Faculty of Agric., Ain Shams Univ., for their helping, fruitful advising and faithful encouragement during the progresses.

Finally, I am indebted to thank my family for their continuous encouragement.

CONTENTS

Title	Page
List of Tables.....	IV
List of Figures.....	VII
I. Introduction.....	1
II. Review of Literature.....	5
1. Host range.....	5
2. The behavior of <i>Ganoderma</i> on their hosts.....	6
3. Isolation.....	8
4. Pathogenicity.....	9
5. Identification.....	10
5.1. Traditional method.....	10
5.1.1. Morphological features.....	10
5.2. Cultural features.....	13
5.3. Molecular identification, sequence and phylogenetic analysis.....	14
6. Early diagnosis.....	16
7. Extracellular enzymes.....	18
8. Toxin production.....	22
III. Materials and Methods.....	23
1. Source of fungal isolates and diseased plant samples.....	23
2. Isolation, Purification and Maintenance.....	23
3. Identification of causal organism.....	23
3.1. Morphological identification.....	24
3.1.1. External morphological features.....	24
3.1. 2. Internal morphological features.....	24
3.1. 3. Scanning electron microscopy.....	24
3.2. Cultural characteristics and chemical tests.....	25
3. 3. Molecular identification.....	25
3.3.1.DNA extraction, polymerase chain reaction (amplification), sequence and phylogenetic analysis.....	25

3.3.1.1. DNA extraction.....	25
3.3.1.2. Polymerase chain reaction (amplification).....	26
3.3.1.3. Sequence and phylogenetic analysis.....	26
4. Artificial inoculation.....	27
4.1. Preparation of inoculum and inoculation.....	27
5. Early diagnosis.....	28
5.1. Root sampling for early diagnostic tests.....	28
5.2. DNA extraction from root samples and polymerase chain reaction.....	29
6. Physiological studies.....	29
6. 1. Cellulase assay.....	29
6.1.1. Extraction of cellulase enzyme from the cultural filtrate of pathogenic fungus.....	29
6.1.2. Extraction of cellulase enzyme from diseased plants.....	29
6.1.3. Determination of cellulase [β (1 - 4) glucanases] activity...	30
6.2. Pectin transeliminase (s) assay.....	31
6.2.1. Extraction of pectin transeliminase (s) (pectinlyase) from the cultural filtrate of pathogenic fungus.....	31
6.2.2. Extraction of pectin transeliminase (s) enzymes from diseased plants.....	31
6.2.3. Determination of pectin transeliminase (s) enzyme activity.....	31
6.3. Determination of lignin content.....	32
6.3.1. Extraction of lignin from the pathogen and diseased plants.....	32
7. Extraction of toxin from the pathogenic fungus.....	32
7.1. Isolation of toxin.....	32
7.2. Bioassay (Detached leaf assay).....	33
8. Colonization of sour orange (<i>Citrus aurantium</i>) root pieces by <i>Ganderma resinaceum</i> at different temperature degrees of incubation and Scanning electron microscopy studies.....	33

8.1. Colonization of sour orange (<i>Citrus aurantium</i>) root pieces by <i>Ganderma resinaceum</i> at different temperature degrees of incubation	33
8.2. Scanning electron microscopy studies.....	34
IV. Results.....	35
1. Source of fungal isolates and diseased plant samples.....	35
2. Isolation, purification and maintenance.....	38
3. Identification of the causal organisms.....	38
3.1.Morphological features, cultural characteristics and chemical test.....	38
3.2.Molecular identification (amplification, sequence and phylogenetic analysis).....	65
4. Artificial inoculation.....	75
5. Early diagnosis.....	81
6. Physiological studies.....	84
6.1. Cellulase activity.....	84
6.2. Pectin transesterinase (s) activity.....	93
6.3. Lignin degradation.....	101
7. Bioassay of toxin (detached leaf assay).....	108
8. Colonization of sour orange (<i>Citrus aurantium</i>) root pieces by <i>Ganderma resinaceum</i> at different temperature degrees of incubation and Scanning electron microscopy studies.....	109
8.1. Colonization of sour orange (<i>Citrus aurantium</i>) root pieces by <i>Ganderma resinaceum</i> at different temperature degrees of incubation.....	109
8.2 Scanning electron microscopy studies.....	113
V. Discussion.....	120
VI. Summary.....	134
VII. References.....	145
Arabic Summary	

LIST OF TABLES

Table	Title	Page
Table 1	Morphological features, cultural characteristics and chemical test of <i>Ganoderma</i> isolates.....	64
Table 2	List of five <i>Ganoderma</i> isolates.....	66
Table 3	Determination of cellulase [β (1-4) glucanase] activity (as μg glucose min^{-1} g^{-1} sample) in Czapek's medium 10 days after inoculation with the tested pathogenic <i>Ganoderma resinaceum</i> isolates.....	87
Table 4	Determination of cellulase [β (1-4) glucanase] activity (as μg glucose min^{-1} g^{-1} sample) extracted from sour orange rootstock roots (<i>Citrus aurantium</i>) inoculated with <i>Ganoderma resinaceum</i> isolates 6, 9 and 12 months after inoculation, respectively.....	87
Table 5	Determination of cellulase [β (1-4) glucanase] activity (as μg glucose min^{-1} g^{-1} sample) extracted from volkamer lemon rootstock roots (<i>Citrus volkameriana</i>) inoculated with <i>Ganoderma resinaceum</i> isolates 6, 9 and 12 months after inoculation, respectively.....	88
Table 6	Determination of cellulase [β (1 - 4) glucanase] activity (as μg glucose min^{-1} g^{-1} sample) extracted from fan Palm roots (<i>Washingtonia filifera</i>) inoculated with pathogenic <i>Ganoderma resinaceum</i> isolates 6 months after inoculation, respectively.....	88
Table 7	Cellulase [β (1-4) glucanase] activity in inoculated sour orange rootstock roots (<i>Citrus</i>	