Serum Ficolin- 2 Concentration and its Relation to Severity of Liver Inflammation and Response to Antiviral Therapy in Egyptian Chronic Hepatitis C Patients

Thesis Submitted for Partial Fulfillment of M.D. Degree in Internal Medicine

Submitted by

Abd Elbadea MohamedAllakany M, B, B.Ch, M.Sc.

Under supervision of

Prof. Dr/ Sayed Mohamed Shalaby

Professor of Internal Medicine Faculty of Medicine, Ain Shams University

Prof. Dr. /Essam Mohamed Bayomy Helal

Professor of Internal Medicine Faculty of Medicine, Ain Shams University

Prof. Dr. /Wesam Ahmed Ibrahim Mohamed

Professor of Internal Medicine Faculty of Medicine, Ain Shams University

Dr. /Angie Mohamed Ali Rund

Lecturer of Internal Medicine Faculty of Medicine, Ain Shams University

> Faculty of Medicine Ain ShamsUniversity 2017



سورة طه الآيه رقم ۱۱۶



Acknowledgement

First of all, all gratitude is due to **God**almighty for blessing this work, until it has reached its end, as a part of his generous help, throughout my life.

Really I can hardly find the words to express my gratitude to **Professor Doctor/ Sayed Mohamed Shalaby,** Professor of Internal Medicine, faculty of medicine, Ain Shams University, for his supervision, continuous help, encouragement throughout this work and tremendous effort he has done in the meticulous revision of the whole work. It is a great honor to work under his guidance and supervision.

I would like also to express my sincere appreciation and gratitude to **Prof. Dr. /Essam Mohamed Bayomi Helal** Professor of Internal Medicine, faculty of medicine, Ain Shams University, for his continuous directions and support throughout the whole work. I really appreciate his patience and support.

Really I can hardly find the words to express my gratitude to **Prof. Dr. /Wesam Ahmed Ibrahim Mohamed** Professor of Internal Medicine, Faculty of Medicine, Ain Shams University for her continuous directions and meticulous revision throughout the whole work.

I would like to express my deepest gratitude and sincere thanks to **Dr./Angie Mohamed Ali Rund** Lecturer of Internal Medicine, Faculty of Medicine, Ain Shams University, for suggesting and planning this work, her instructive supervision, continuous guidance, unlimited help and unfailing support, valuable instructions throughout the work and final revision of the manuscript.

I am deeply grateful for **Prof.Dr. /Khaled AbdelKader Farrag**, consultant of Hepatology and Gastroentrology Damanhour National Medical Institute, for his valuable help and guidance.

I also extend my sincere thanks and gratitude to **Prof. Dr.**/Ashraf Basiony Shalaby, Consultant of clinical pathology and Blood and Virus Research.

With sincere appreciation to **Dr. /Marwan Ashraf Shalaby** and the staff of Shalaby Laboratories for their efforts in producing the results of analysis related to the subject of the message.

This work was also dedicated to the spirit of my father and my mother, may Allah have mercy on them.

Last but not least, I dedicate this work to my family, whom without their sincere emotional support, pushing me forward this work would not have ever been completed.

Abd Elbadea Mohamed Allakany

Contents

List of Abbreviations	i
List of Tables	ii
List of Figurs	
Introduction and Aim of the Work	
Hepatitis C Virus	4
Ficolins	40
Ficolin 2 in Hepatitis C Infection	70
Patients and Methods	75
Results	79
Discussion	1.3
Summary and Conclusion	١٠8
Recommendations	110
References	111
Arabic Summary	

List of Abbreviations

AASLD/IDSA/IAS-USA: American Association for the

Study of Liver Diseases/ Infectious Diseases Society of America / International

Antiviral Society – USA

ac-LDL : Acetylated low-density lipoprotein

ARF : Alternate reading frame

BOC : Boceprevir

CDC : Centers for Disease Control and Prevention

CLDN1 : Claudin-1

CRHD : Rheumatic heart disease

CRP : C-reactive protein

DAA : Directly acting antivirals

DCV : Daclatasvir

EASL : European Association for the Study of the

Liver

EIAs : Enzyme-linked immunoassays

FCN2 : Ficolin 2

GAG : Glycosaminoglycans GlcNAc : N-acetyl glucosamine

GT : Genotype

HAE-C1-INH: Hereditary angioedema due to C1-inhibitor

deficiency

HCC : Hepatocellular carcinoma

HCV : Hepatitis C virus

H-ficolin : Hakata antigen or ficolin-3

HIV : Human immunodeficiency virus

HLA : Human leukocyte antigen

IL28B : Interleukin 28B

IRES : An internal ribosome entry site

IRRDR : Interferon RBV resistance- determining

region

List of Abbreviations(Cont.)

IVDU : Intravenous drug usersLDL : Low-density lipoproteins

LDV : Ledipasvir

L-ficolin : Liver ficolin or ficolin-2

MASPs : Mannan associated serine proteases

MBL : Mannose-binding lectin

MELD : Model for End-Stage Liver Disease

M-ficolin : Monocyte ficolin or ficolin-1

mRNA : Messenger RNA

MSM : Men who have sex with menNPC1L1 : Niemann–Pick C1-like 1NTR : Nontranslated regions

OCLN : Occludin

ORF : Open reading frame

PCR : Polymerase chain reaction

PEG-IFN : Pegylated Interferon

PTX3 : Pentraxin-3 RBV : Ribavirin

RTKs : Tyrosine kinases

SJS : Stevens - Johnson syndrome

SMV : Simeprevir

SOC : Therapy standard of care therapy

SOF : Sofosbuvir

SR-BI : Scavenger receptor B type I SVR : Sustained virological response

TE: Treatment-experienced

TMA : Transcription-mediated amplification

TN : Treatment-naïve

TPV : Telaprevir

WHO : World Health Organization

List of tables

Table	Title	Page
1	Child Pugh score for disease staging and	9
	prognosis	
2	The human ficolins	42
3	Some disease associations of L-ficolin insufficiency	67
4	Demographic data of the patient's gender	79
5	Laboratory data of patients group	82
6	Comparison between the two groups as regard to liver enzymes	83
7	Distribution of studied group according to patient's liver enzymes (After TTT)	84
8	Comparison between the two groups as regard to FCN2 (student t-test) (pre treatment)	85
9	Abdominal ultrasonography data of the patients group.	86
10 a	Liver biopsy data of the patients group.	87
10b	Comparison of FCN2 in A1F1 and A2F2 patients in liver biopsy	88
11	Anti-schisto.Abs. data of the patients group.	88
12	FCN2 before and after treatment in patients group.	89
13	Comparison of ALT before and after treatment in patients group.	90
14	Comparison of FCN2 in males and females patients before and after treatment.	91
15	Comparison of AST in high AST patients and normal AST paints before and after treatment	93
16	Comparison of FCN2 before and after treatment in high ALT patients and normal ALT paints.	94
17	Comparison of FCN2 levels in responders and nonresponders before and after treatment.	96
18a	Comparison between FCN2 and Other parameters in patients group.	97
18b	Comparison between FCN2 and PCR (viral load) in responders and nonresponders	98

List of figures

Fig.	Title	Page
1	Genome organisation of Hepatitis C virus	11
2	A simplified diagram of the HCV replication cycle	12
3	A simplified diagram of the HCV replication cycle	15
4	Detection limits and linear dynamic ranges of commercially available HCV RNA detection assays	19
5	The human FCN2 gene.	54
6	L-ficolin structure	55
7	Demographic data of the patient's gender	79
8	Comparison between the two groups as regard to AST (SGOT)	83
9	Comparison between the two groups as regard to ALT (SGPT)	84
10	Comparison between the two groups as regard to FCN2	85
11	Abdominal ultrasonography data of the patients group	86
12	Liver biopsy data of the patients group.	87
13	Anti-schisto.Abs. data of the patients group.	88
14	Comparison of FCN2 before and after treatment in patients group	89
15	Comparison of ALT before and after treatment in patients group	90
16	Comparison of FCN2 in males and females patients before and after treatment.	92
17	Comparison of AST in male and female patients before and after treatment.	93
18	Comparison of FCN2 before and after treatment in high ALT patients and normal ALT patients	95

List of figures(Cont.)

Fig.	Title	Page
19	Comparison of FCN2 levels in responders and	96
	nonresponders before and after	
	treatment.	
20	Correlation between FCN2 and S. Albumin	99
21	Correlation between FCN2 and ALK.	99
	Phosphates	
22	Correlation between FCN2 and AST	100
23	Correlation between FCN2 and ALT	100
24	Correlation between FCN2 and Total bilirubin	101
25	Correlation between FCN2 and PCR	101
26	Correlation between FCN2 and Liver Biopsy	102

Introduction

Chronic hepatitis C (HCV) infection is a major worldwide public health problem. Worldwide, up to 170 million patients are chronically infected and are at risk of developing liver cirrhosis and hepatocellular carcinoma. (*Shepard et al.*, 2005).

Egypt has the highest prevalence of adult HCV infection in the world, averaging 15%-25% in rural communities. (*Esmat et al.*, 2012).

Infection with HCV causes chronic disease in 80% of cases and current treatments are not completely effective. (*Garcia-Samaniego et al.*, 2013).

The innate and adaptive immunity are both involved in defense against HCV infection. The innate immune system provides an immediate line of defense, triggering inflammation and playing a critical role in activating adaptive immunity. (*Tarr et al.*, 2012).

Innate immunity comprises both cellular and humoral components, including complement activation and involving complement C1q, Lectins, Collectins and Ficolins. These molecules activate the complement cascade, neutralize pathogens and recruit antigen presenting cells. Some studies reported that serum mannan-binding lectin (MBL) concentrations are increased in Egyptian chronic hepatitis C patients and related to disease progression and response to interferon-based antiviral therapy. (Esmat et al., 2012).

Ficolin -2 is a kind of human serum complement lectins, which is implicated in innate immunity. Recent studies reported that early increased Ficolin-2

concentrations are associated with severity of liver inflammation and efficacy of antiviral therapy in chronic hepatitis C patients (non Genotype 4). Little is known about the relation of Ficolin-2 to severity of disease and response to interferon-based therapy in Egyptian chronic hepatitis C patients (Genotype 4). (*Hu et al.*, 2013).

Aim of the Work

This work aims to determine the dynamics of Ficolin-2 in Egyptian chronic hepatitis C patients: relation of serum concentration to severity of disease and response to antiviral therapy.

Chapter (1):

Hepatitis C Virus

Epidemiology

Hepatitis C is a disease of significant global impact. According to the World Health Organization there are about 150 million people chronically infected with hepatitis There considerable (HCV). are differences: in some countries, e.g., Egypt, the prevalence is >10% (WHO, 2013). In Africa and the Western Pacific prevalence is significantly higher than in North America and Europe (CDC, 2013). It is estimated that there are 2-5million HCV-positive persons in Europe. Certain groups are preferentially affected, like injection drug users. In Europe and the United States chronic hepatitis C is the most common chronic liver disease. The majority of liver transplants performed in these regions are for chronic HCV. It is difficult to determine the number of new HCV infections, as most acute cases are not noticed clinically. Recent numbers from Europe still show an ongoing epidemic of acute HCV especially among intravenous drug users (IVDU) and men who have sex with men (MSM) (Rockstroh et al., 2012).

Transmission

Parenteral exposure to hepatitis C is the most efficient means of transmission. The majority of patients infected with HCV in Europe and the United States acquired the disease through intravenous drug use or blood transfusion, which has become rare since routine testing of the blood supply for HCV began. The following possible

routes of infection have been identified in blood donors (in descending order of transmission risk):

- 1. Injection drug use
- 2. Blood transfusion
- 3. Sex with an intravenous drug user
- 4. Having been in jail more than three days
- 5. Religious scarification
- 6. Having been struck or cut with a bloody object
- 7. Pierced ears or body parts
- 8. Immunoglobulin injection

Very often in patients with newly diagnosed HCV infection no clear risk factor can be identified. Factors that may increase the risk of HCV infection include greater numbers of sex partners, history of sexually transmitted diseases, and failure to use a condom. Whether underlying HIV infection increases the risk of heterosexual HCV transmission to an uninfected partner is unclear. The seroprevalence of HCV in MSM (men who have sex with men) ranges from about 4 to 8%, which is higher than the for prevalence reported general European populations, increasing globally over the last decade (Rockstroh et al., 2012).

The risk of perinatal transmission of HCV in HCV RNA-positive mothers is estimated to be 5% or less (*Ohto* et al., 1994). Cesarean section has not been shown to reduce transmission. There is no evidence that breast feeding is a risk factor. Hemodialysis risk factors include blood transfusions, the duration of hemodialysis, and the prevalence of HCV infection in the dialysis unit, and the type of dialysis. The risk is higher with in hospital hemodialysis vs. peritoneal dialysis. Contaminated medical equipment, traditional medicine rites, tattooing, and body piercing are considered rare transmission routes. There is