### Diagnostic role of Pap-smear in patients with vaginal discharge

#### THESIS SUBMITTED FOR PARTIAL FULFILLED OF MASTER DEGREE IN OBSTETRICS AND GYNECOLOGY

#### By / MANAL AHMED FAIK SAMY

M.B.B.CH. 1990 Diploma Obstetrics& Gynecology1995 ALEXANDRIA UNIVERSITY

**Under Supervision of** 

#### MOHAMED BAYOMY SAMOUR

PROFESSOR OF OBSTETRICS AND GYNECOLOGY FACULTY OF MEDICINE AIN SHAMS UNIVERSITY

#### SHERIF HANAFI HUSSAIN

LECTURER in OBSTETRICS AND GYNECOLOGY FACULTY OF MEDICINE AIN SHAMS UNIVERSITY

AIN SHAMS UNIVERSITY

2010

#### **ACKNOWLEDGEMENT**

Thanks to God who has allowed and helped me to accomplish this work. This study has only been possible due to the continuous support and encouragement from many highly valued individuals.

Before presenting this study, I would like to express my deepest gratitude and great esteem to *Prof. Dr. Mohamed Bayomy Samour*, Professor of Obstetrics and Gynecology, Faculty of Medicine, Ain Shams University. He gave me the necessary guidance that has enabled me to complete this research.

My sincere thanks and gratefulness to *Dr. Sherif Hanafi Hussain*, Lecturer in Obstetrics and Gynecology, Faculty of Medicine, Ain Shams University. He provided me with the advice and support crucial for this work. I am grateful for his continuous help, time and endless co-operation.

Finally, I would like to thank all the members of the members of the Early Cancer Detection Unit at Ain Shams for their help and co-operation.

#### List of content

| List of abbreviations                           | I     |
|---|-------|
| List of tables                                  | II    |
| List of figures                                 | III   |
| Introduction                                    | IV    |
| Review of literature                            |       |
| Chapter (1): Microbiology of vagina             | (2)   |
| Chapter (2): Anatomy of the cervix              | (24)  |
| Chapter (3): Cervical cytology                  |       |
| Chapter (4): Human Papillomavirus               |       |
| Chapter (5): Cervical intraepithelial neoplasia | (80)  |
| Material and methods                            | (99)  |
| Results   | (105) |
| Discussion                                      | (115) |
| Summary   | 1     |
| Conclusion and recommendation                   | (135) |
| References                                      | ` '   |
| Arabic summary                                  | (160) |

#### List of abbreviations

| Abbreviation | Meaning   |
|--------------|---|
| ACOG         | American College of Obstetricians and Gynecologists   |
| ACS          | American Cancer Society                               |
| ALTS         | ASCUS/LSIL Triage study                               |
| ASC          | A typical squamous cells                              |
| ASCUS        | A typical squamous cells of undetermined significance |
| ATECC        | Atypical endo-cervical columnar cells                 |
| BV           | Bacterial vaginosis                                   |
| CIN          | Cervical intraepithelial neoplasia                    |
| CIS          | Carcinoma in situ                                     |
| CMV          | Cytomegalovirus                                       |
| DVI          | Direct visual inspection                              |
| FDA          | Food and Drug administration                          |
| G.V          | Gardnerella vaginalis                                 |
| HHV          | Human herpes virus                                    |
| HIV          | Human immunodeficiency virus                          |
| HPV          | Human Papillomavirus                                  |
| HSIL         | High grade Squamous intraepithelial lesion            |
| ICC          | Invasive cervical carcinoma                           |
| IUD          | Intrauterine device                                   |
| KOH          | Potassium hydroxide                                   |
| LBC          | Liquid-based cytology                                 |
| LCR          | Long control region                                   |
| LEEP         | Loop electrosurgical excision procedure               |
| LLETZ        | large loop excision of the transformation zone        |
| LSIL         | Low grade Squamous intraepithelial lesion             |
| NCI          | National Cancer Institute                             |
| OCP          | Oral contraceptive pills                              |
| PAP          | Pap smear Papanicolaou smear                          |
| PCR          | Polymerase chain reaction                             |
| PID          | Pelvic inflammatory disease                           |
| RVVC         | Recurrent vulvovaginal candidiasis                    |
| SCC          | Squamous cell carcinoma                               |
| SCJ          | Squamocolumnar junction                               |
| STDs         | Sexual transmitted diseases                           |
| TMI          | Time of first marriage                                |
| VVC          | Vulvovaginal candidiasis                              |

#### List of tables

| Tabel | Content   | Page |
|-------|---|------|
| 1     | Causes of vaginal discharge                               | 8    |
| 2     | Clinical manifestation and sequale of Trichomonas         | 15   |
| 3     | Classification of vulvovaginal candidiasis                | 18   |
| 4     | Sources of errors encountered in Pap smear                | 36   |
| 5     | The 2001 Bethesda system                                  | 43   |
| 6     | Risk factors known or postulated with HPV infection       | 63   |
| 7     | Comparision of cytology classification systems            | 85   |
| 8     | Acetowhite Changes in low and and high grade lesions      | 89   |
| 9     | Age distribution of the patients                          | 106  |
| 10    | Selected demographic and behavioral characteristics of    | 106  |
|       | patients with vaginal discharge.                          |      |
| 11    | Distribution of parity in relation to patient number      | 107  |
| 12    | Symptoms associated with vaginal discharge                | 109  |
| 13    | Pap smear results according to the type of infection      | 109  |
| 14    | The distribution of associated symptoms with vaginal      | 110  |
|       | Infection   |      |
| 15    | The distribution of cases with the duration of marriage   | 111  |
| 16    | The distribution of cases with the age of first marriage  | 111  |
| 17    | The distribution of patients with smoking                 | 112  |
| 18    | Distribution of cervical ectropion in patients with       | 112  |
|       | vaginal discharge.  |      |
| 10    |   | 110  |
| 19    | Distribution of cervical polyp in patients with vaginal   | 113  |
| 20    | discharge   | 110  |
| 20    | Cytological results of Pap smear                          | 113  |
| 21    | Characteristics of patients with no vaginal infection and | 114  |
|       | their cytological results                                 |      |
| 22    | Characteristics of patients with epithelial cell          | 114  |
|       | abnormalities   |      |

# Review of Literature

## Chapter 1

#### Microbiology of the Vagina

#### Vaginal microflora

The vagina and cervix of healthy females harbor an indigenous microflora. The normal vaginal flora is fairly homogeneous and consists of aerobic and anaerobic bacteria. The aerobic components include *lactobacilli*, Group B and D *streptococci*, *Staphylococcus epidermidis*, *Staphylococcus aureus*, and gram-negative enteric rods such as *Escherichia coli* (**Larsen and Galask**, **1980**).

Anaerobic cocci were reported in 7% to 57% of the cultures, predominantly *Peptostreptococcus asaccharolyticus* and *Peptostreptococcus anaerobius*, anaerobic gram-negative bacilli were isolated from most of the cultures; their isolation rates were between 57% and 65%. The predominant strains were *Prevotella bivia*, *Prevotella disiens*, *Bacteroides fragilis* group (Larsen and Galask, 1980).

Normal variations in cervico-vaginal flora are related to the effects of age, pregnancy, and menstrual cycle (Larsen and Galask, 1980).

The microflora in females before puberty, during the childbearing years, during pregnancy, and after menopause are not uniform. Colonization with lactobacilli is low in prepubertal females and postmenopausal females and high in pregnant females as well as those females in their reproductive years who are not pregnant (**Brook**, 2002).

At the time of puberty, *lactobacilli* become the predominant inhabitant of the vagina, presumably because of the effect of estrogens on the glycogen content of vaginal epithelial cells (**Paavonen**, **1983**).

When estrogen levels falls, such as in postpartum or menopausal women, the prevalence of *lactobacilli* declines (**Redonodo-lopez, 1990**).

A healthy vaginal ecosystem is dominated by certain species of *Lactobacillus*, which exert a significant influence on the microbiology of the vagina, some species of *Lactobacillus* suppress the growth of other endogenous bacteria in the vagina through the production of organic acids such as lactic acid,  $H_2$   $O_2$  and bacteriocins or lactocins (**Antonio**, **1999**).

The production of organic acids maintains the vaginal pH at < 4.5, thereby creating an inhospitable environment for the growth of most endogenous pathogenic bacteria. In addition to lactic acid, the combination of  $H_2O_2$  and lactocins further suppresses the endogenous pathogenic bacteria to maintain a healthy vaginal ecosystem (**Redondo-Lopez, 1990**).

Ninety-six percent of *Lactobacillus* species found in a healthy vaginal ecosystem produced  $H_2$   $O_2$ , whereas only 6% of the *lactobacilli* recovered from women with bacterial vaginosis (BV) produced  $H_2$   $O_2$  (Eschenbach, 1989).

**Hillier** (1992), has postulated that  $H_2O_2$  inhibits the growth of vaginal microorganisms either directly or through the enhancement of the enzyme peroxidase-halide.

H<sub>2</sub> O<sub>2</sub> produced by lactobacilli was found to be lethal to *Gardnerella* vaginalis, *Bacteroides bivia*, and *Escherichia coli* (**Klebanoff**, 1991).

**Hawes et al (1996)**, found that vaginal colonization with  $H_2$   $O_2$ -producing *lactobacilli* was associated with a decrease in the occurrence of BV.

A third component to this system is the production of bacteriocin or lactocin, which are proteins that have bactericidal activity, can inhibit adverse group of bacteria, including *G vaginalis* (Brink, 1994).

Bacteriocin produced by vaginal *lactobacilli* appears to have a broad spectrum of activity and inhibits a wide range of gram-positive and gramnegative bacteria (McGroaty,1993).

**McLean and McGroarty** (1996), suggested that lactic acid and low pH work synergistically with bacteriocin and may be more important than hydrogen peroxide in inhibiting the growth of *G vaginalis*.

Aroutcheva et al (2001), suggest that the number of bacteria (biocell mass) in the vaginal environment is responsible for pH changes. The biocell mass or numbers of bacteria are significant with regard to the quantity of organic acids produced. Thus, the mere presence of lactobacilli and their ability to produce lactic and other organic acids is not the only important factor in maintaining a healthy vaginal ecosystem—also important is the active growth and the increase in number of bacteria present.

(Newton et al, 2001) showed that several factors appear to have a major impact on the vaginal flora:

1) Hormones, especially estrogen, which support the growth of at least

Lactobacillus and Candida species.

- **2)** The menses, which appears to increase the growth of non-*Lactobacillus* flora.
- 3) STDs influence the growth of non-STD bacteria in the vagina
- **4**)Genetic differences, lead to variations in the attachment of bacteria to epithelial cells, which in turn influence the ability of the bacteria to compete with other microorganisms.
- **5**) The interactions among microbial species, exogenous STD microorganisms and non-STD microorganisms in the vaginal flora.

#### Vaginal cycle

The transient hormonal changes which occur each month over the course of the menstrual cycle affect vaginal flora.:

Estrogen increases the vaginal epithelial activity resulting in thickened epithelium with glycogen deposited in the superficial cells, the glycogen is metabolized by *lactobacilli* to lactic acid resulting in vaginal acidity of about PH 4.5 (Wilson, 2004).

Several studies have suggested that abnormal vaginal flora develop most frequently at the beginning of the menstrual cycle when the estrogen levels are lowest and the tendency for Gram- positive cocci to appear mostly at the beginning of the menstrual cycle was also noticed (**Keane**, 1997).

Bacterial vaginosis occurs most commonly at the beginning of the menstral cycle when estradiol levels are at their lowest (Wilson, 2007).

Estradiol levels increase during the follicular phase of the menstrual cycle which may account for mid- cycle spontaneous remission of bacterial

vaginosis and the high level of spontaneous remission of BV during pregnancy (Hav, 1997).

A microbiological study throughout the menstrual cycle found the rate of recovery of *lactobacilli* increased over the cycle and the concentration of non lactobacilli species was higher during menstruation (**Eschenbach**, **2000**).

Perimenopausal and postmenopausal women have low estrogen levels and they have been found to have high rates of abnormal vaginal flora at 35% and 70% (**Taylor-Robinson**, **2002**).

#### **Etiology of vaginal discharge:**

#### 1- Normal physiologic discharge

Normal vaginal secretions are composed of vulvar secretions from sebaceous, sweat, bartholin and skene glands; transudate from the vaginal wall; exfoliated vaginal and cervical cells; cervical mucous; endometrial and oviductal fluids and micro-organisms and their metabolic products. Normal vaginal secretions are floccular in consistency, white in colour and usually located in the dependent portion of the vagina (posterior fornix). The type and amount of vaginal secretions are influenced by hormonal levels, vaginal secretions may increase in the middle of the menstrual cycle because increase in the amount of cervical mucus (Huggins and Preti, 1981).

#### 2- Pathological vaginal discharge:

Causes of Vaginal discharge are many & can be summarized in this table:

Table (1): Causes of vaginal Discharge

#### Non-infective

- -Physiological
- -Cervical ectopy
- -Foreign bodies, such as retained tampon
- -Vulvual dermatisis

#### **Non-sexually transmitted infection**

- -Bacterial vaginosis
- -Candida infections

#### **Sexually transmitted infections**

- Chalamydia trachomatis
- -Neisseria gonorrhoeae
- -Trichomonas vaginalis

Quoted from: Spence and Melville 2007

#### **Bacterial vaginosis:**

#### **Epidemiology**

Bacterial vaginosis is the most prevalent cause of symptomatic vaginitis in sexually active women. At same time, 50% of the total number of bacterial vaginosis infections are asymptomatic (**Schwebke**, **2003**). Bacterial vaginosis is characterized by shift of vaginal flora from normal lactobacilli dominant to flora with greatly reduced numbers of lactobacilli and an overgrowth of *Gardnerella vaginalis*, *Mycoplasma hominis*, and anaerobic

bacteria such as *peptostretococci*, *Prevotella* spp and *Mobiluncus* spp (Wilson,2004).



Figure(1):Clue cells are epithelial cells with bacteria adhering to their surface and sometimes obscuring their borders. Clue cells indicate bacterial vaginosis (Library of the Centers for Disease Control and Prevention).

#### Risk factors of bacterial vaginosis:

- 1- Young age (Ralph, 1999).
- 2- Black ethnicity (Goldenberg, 1996).
- 3- Douching (Hawes, 1996).
- 4- IUD as contraception (Avonts, 1990).
- 5- Recent change in sexual partner (Hawes, 1996).
- 6-Multiple partners compared with one partner (Nillson, 1997).
- 7-Smoking (**Schwebke**, **1999**), cigarette smoke contains chemicals like nicotine which demonstrated in cervical mucus of smokers and may directly alter the vaginal microflora or may act by depleting Langerhans cells in cervical epithelium leading to local immunosuppression. This may be responsible for change of cervical flora causing bacterial vaginosis.

#### Etiology of Bacterial vaginosis:

Bacterial vagionosis is not a monoetiologic infection but rather a synergistic polymicrobial disequilibrium of the vaginal ecosystem. The exact mechanism for the onset of bacterial vaginosis remains a mystery. It is associated with a reduction in hydrogen peroxide producing lactobacilli, a rise in the vaginal pH, and the overgrowth of BV associated organisms. But which of these happen first, and which is the most is still a matter of debate (Wilson, 2004).

1) Reduction in the lactobacilli and hydrogen peroxide production (Klebanoff et al, 1990), showed in vitro that combining myeloperoxidases with hydrogen peroxide and a halide produced a potent oxidant, which was toxic to BV associated bacteria. Myeloperoxidases activity has been found in vaginal fluid and cervical mucus, and chloride is present in cervical mucus. The toxin effect of this LB+/myeloperoxidase/chloride system was rapid, with reduction in numbers of *G vaginalis* at 15 minutes and complete loss of viability at 60 minutes.

(Pavlova et al, 1997) suggest that associated organisms are sensitive to lactic acid and hydrogen peroxide, suppression of *lactobacilli* must come before overgrowth of BV associated bacteria.

#### 2) Change in pH:

The low pH of the vagina (3.2-4.8) is attributed to production of lactic acid by *lactobacilli* metabolism, and by the conversion of glycogen to lactic acid by estrogenished vaginal epithelial cells (**Wilson, 2004**).