# Rate of the detection of anti-hepatitis B core antibody (Anti-HBc) among blood donors negative for hepatitis B surface antigen (HBsAg)

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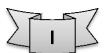
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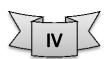
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### **List of Abbreviations**

ADV	ADEFOVIR DIPIVOXIL
AFP	
<i>ALT</i>	ALANINE AMINOTRANSFERASE
Anti-HBc	
Anti-HBe	Antibody to hepatitis B "e" antigen
Anti-HBs	ANTIBODY TO HEPATITIS B SURFACE ANTIGEN
Anti-HBx	
ASSLD	AMERICAN ASSOCIATION FOR THE STUDY OF LIVER DISEASE
<i>AST</i>	ASPARTATE AMINOTRANSFERASE
Au	
<i>bp</i>	BASE PAIR
cccDNA	
dATP	DEOXYADENOSINE TRIPHOSPHATE
DNA	DEOXYRIBONUCLEIC ACID
<i>EASL</i>	EUROPEAN ASSOCIATION FOR THE STUDY OF LIVER
ELISA	ENZYME-LINKED IMMUNOSORBENT ASSAY
<i>ETV</i>	ENTECAVER
GSHV	GROUND SQUIRREL HEPATITIS VIRUS
HAV	
HBcAg	
HBeAg	HEPATITIS B EARLY ANTIGEN
HBsAg	
HBV	HEPATITIS B VIRUS
HBxAg	
HCC	HEPATOCELLULAR CARCINOMA
HCV	HEPATITIS C VIRUS
HDV	HEPATITIS DELTA VIRUS

HIV	HUMAN IMMUNODEFICIENCY VIRUS
<i>IFN</i> - α	INTERFERON ALFA
<i>IFN-β</i>	INTERFERON BETA
<i>IFN-</i> ω	INTERFERON OMEGA
	INTERFERON GAMMA
<i>IFN- λ</i>	INTERFERON LAMBDA
	International Normalized Ratio
LAM	LAMIVUDINE
LdT	
	MILLION UNITS
ORFs	OPEN READING FRAME
PCR	POLYMERASE CHAIN REACTION
PEG-IFN	PEGYLATED INTERFERON
POL	POLYMERASE
	PROTHROMBIN TIME
RT	
RT-PCRRE	VERSE TRANSCRIPTION POLYMERASE CHAIN REACTION
TDF	TENOFOVIR DISOPROXIL FUMARATE
ULN	UPPER LIMIT OF NORMAL
	World Health Organization
WHV	WOODCHUCK HEPATITIS VIRUS

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### Introduction

Hepatitis B virus (HBV) is a hepatotropic Deoxynucleic acid (DNA) virus belonging to the genus Orthohepadnavirus, family Hepadnaviridae (*Tiollais et al.*, 1985).

The virus infects only human and some other non-human primates. It is transmitted mainly by infectious blood, serum or plasma. Once inside the host, HBV is transported through the blood stream to the liver, which is the primary site of infection (Korba et al., 1989; Uemoto et al., 1998).

Hepatitis B surface antigen (HBsAg) is the first marker to appear in the circulation prior to biochemical evidence of liver damage or the onset of jaundice. Hepatitis B core antibody (anti-HBc) is detectable 2-4 weeks after the appearance of the surface antigen; it persists throughout the infection and after recovery (Harrison et al., 2000).

It was demonstrated that in some individuals negative for HBsAg and positive for anti-HBc, HBV continue to replicate which may explain why some blood derivatives, negative for HBsAg but positive anti-HBc, are able to transmit the infection, both after transfusion and after transplantation of organs (Ben-Ayed et al., 2001).

Isolated anti hepatitis B core (anti-HBc), which is defined as positive Anti-HBc with undetectable HBsAg and Hepatitis B Surface antibody (anti-HBs), may occur in any of several situations: (i) a false-positive Anti hepatitis B core antigen (anti-HBc) result; (ii) low levels of HBV replication inside the hepatocyte, without detectable production of HBsAg; (iii) the window phase of acute HBV infection; (iv) the loss of anti-HBs with time or failure to develope antibody response against the antigen after infection; or (v) the presence of a vaccine escape mutant, not detected by most of the currently available HBsAg detection tests (Al-Mekhaizeem et al.,2001; Behzad-Behbahani et al.,2006).

### AIM OF THE WORK

The aim of this work is to detect Anti-HBc among blood donors negative for HBsAg in order to evaluate the safety of this blood.

# REVIEW OF LITERATURE

### HISTORY OF HEPATITIS B VIRUS

Hepatitis is a general term meaning inflammation of the liver and can be caused by a variety of different viruses such as hepatitis A, B, C, D and E. Since the development of jaundice as a characteristic feature of liver disease, a correct diagnosis can only be made by testing patients' sera for the presence of specific antiviral antigens or antibodies (Robinson, 1995; Mahoney and Kane, 1999; Hollinger and Liang, 2001).

The existence of a parenterally transmitted form of hepatitis was documented by Lurman (MacCallum, 1947). He reported the development of jaundice in 15% of 1,289 shipyard workers in Bremen, 2 to 8 months after they had received smallpox vaccine prepared from human "lymph". Subsequent outbreaks of hepatitis were eventually traced to the use of improperly sterilized syringes and needles among patients who required parenteral inoculations and among those receiving tattoos (Hasegawa et al., 1991; Fourrier et al., 1999).

In 1937, it was reported from England that approximately 40% of 109 individuals receiving inoculations of a single batch of human measles convalescent serum developed jaundice after incubation periods of up to 114 days. This was followed by an outbreak in British troops given filtered human mumps convalescent plasma (Beeson et al., 1944). These clinical entities were grouped together as "homologous serum jaundice" (Merril et al., 1972). In 1937, Findlay and MacCallum described cases of jaundice occurring 2 to 7 months after inoculation of volunteers with an attenuated strain of yellow fever vaccine prepared with the addition of human serum to stabilize the product (Findlay and MacCallum, 1938). Human serum was subsequently implicated as the vehicle of transmission. It was also observed that the incubation period of parenterally transmitted hepatitis was much longer than that described for infectious hepatitis. It is assumed that these outbreaks were caused by HBV infection, but a hepatitis C virus (HCV) etiology also cannot be excluded (Franco et al., 1992).