

**Study of cytotoxic- T lymphocyte antigen 4 gene
polymorphisms in immune thrombocytopenic
purpura**

Thesis

Submitted for fulfillment of the
M.Sc Degree in
Clinical and Chemical pathology

By

Heba Mostafa Ahmed Mohammed
(M.B., B.Ch)

Supervised by

Prof. Dr Shahira Amen Zayed

Professor of Clinical and Chemical Pathology
Faculty of Medicine, Cairo University

Prof.Dr Somaya Mohammad ELgawhary

Professor of Clinical and Chemical Pathology
Faculty of Medicine, Fayoum University

Dr. Hanan AL-Husseini Mohamed

Lecturer of Clinical and Chemical Pathology
Faculty of Medicine, Cairo University

Faculty of Medicine
Cairo University
2012

Table of contents

Contents	Page
List of abbreviations	i-iv
List of tables	v
List of figures	vi
Acknowledgment	vii
Abstract	viii
Introduction Aim of the study	1-3
Review of literature	
Chapter (1) <u>Immune Thrombocytopenic Purpura</u>	4-19
Chapter (2) <u>Pathogenesis of Immune Thrombocytopenic purpura</u>	20-31
Chapter (3) <u>Cytotoxic T lymphocyte antigen -4</u>	32-43
Subject and methods	44-56
Results	57-70
Discussion	71-75
Summary&Conclusion	76-77
Recommendations	78
References	79-95
Arabic summary	

List Of Abbreviations

• aCL	Anti cardiolipin
• AIHA	Autoimmune haemolytic anaemia
• Ala	Alanine
• ANA	Anti nuclear antibody
• APCs	Antigen presenting cells
• CBC	Complete blood count
• CD	Cluster of differentiation
• CI	Confidence interval
• CMV	Cytomegalovirus
• CO	Carbon monoxide
• CT	Computed tomography
• CTLA-4	Cytotoxic T lymphocyte antigen -4
• CX3CR1	CX3chemokine receptor 1
• DCs	Dendritic cells
• dsDNA	Double stranded DNA
• EDTA	Ethylene diaminetetraacetic acid
• Foxp3	Forkhead family transcription factor p3
• GD	Graves' disease
• GTR	Glucocorticoids induced tumor necrosis receptor

• GM-CSF	Granulocyte macrophage colony stimulating factor
• GO	Graves ophthalmology
• GP	Glycoprotein
• H pylori	Helicobacter pylori
• HCV	Hepatitis C virus
• HDMP	High dose methylprednisolone
• HIV	Human immune virus
• HLA	Human leucocytic antigen
• HO-1	Hemeoxygenase -1
• HSCT	Haematopoietic stem cell transplantation
• ICH	Intracranial haemorrhage
• IDO	Indoleamine dioxygenase
• IFN γ	Interferon gamma
• IgA	Immunoglobulin A
• IgG	Immunoglobulin G
• IgM	Immunoglobulin M
• IL	Interleukin
• ITP	Immune thrombocytopenic purpura
• IvIg	Intravenous immunoglobulin
• KIR	Killer cell immunoglobulin like receptor
• LA	Lupus anticoagulant

- LEF1 Lymphoid enhancer factor-1
- M-CSF Macrophage colony stimulating factor
- MHC Major histocompatibility complex
- MMR Measles, Mumps, and Rubella
- MRI Magnetic resonance imaging
- MS Multiple sclerosis
- NFAT Nuclear factor of activated T cells
- NK Natural killer
- OR Odds ratio
- PaIgG Platelet associated immunoglobulin G
- PCR Polymerase chain reaction
- PEG-rhMGDF growth Pegylated recombinant megakaryocyte factor
- RA Rheumatoid arthritis
- RBCs Red blood cells
- REs Restriction enzymes
- RFLP Restriction fragment length polymorphism
- rhTPO Recombinant thrombopoietin
- sCTLA-4 Soluble cytotoxic T lymphocyte antigen -4
- SD Standard deviation
- SLE Systemic lupus erytheromatosus
- SNP Single nucleotide polymorphism
- SNPs Single nucleotide polymorphisms

- Ss Systemic sclerosis
- TCR T cell receptor
- Th T helper
- Thr Threonine
- TNF- α Tumor necrosis factor alpha
- TPO Thrombopoietin
- Treg T regulatory
- UC Ulcerative colitis
- UTR Untranslated region
- UV Ultra violet
- VLA-4 Very late antigen -4
- WBCs White blood cells

List Of Tables

		Page
1	Clinical features of acute and chronic ITP	7
2	The currently available treatment for ITP	19
3	Reaction composition using PCR master mix	50
4	Age and sex distribution in both ITP and control groups	60
5	Statistical analysis of peripheral blood pictures of ITP and control groups	61
6	Statistical analysis of genotype frequencies of CTLA-4 A49G polymorphism In both ITP and control groups	62
7	Statistical analysis of allele frequencies in both ITP and control groups	63
8	Comparison of genotype frequencies in relation to gender in ITP group	64
9	Comparison of allele frequencies in relation to gender in ITP group	64
10	Statistical analysis of different laboratory parameters in relation to the genotypic frequencies in the ITP group	65
11	Statistical analysis of different laboratory parameters in relation to the allelic frequency in the ITP group	67
12	Clinical and laboratory data of the control group	69
13	Clinical and laboratory data of the ITP group	70

List Of Figures

Page

1	Pathogenesis of autoantibody production in ITP	22
2	Summary of the complex interactions between immune cells in patients with ITP	28
3	Schematic structure of the human CTLA-4 gene	34
4	Function of CTLA-4 protein on Tcells	37
5	Sex distribution in both ITP and control groups	60
6	Genotype frequency of CTLA-4 A49G in ITP and control groups	62
7	Allele frequency of CTLA4 A49G polymorphism in ITP and control groups.	63
8	Comparison of the median platelet counts among different genotypes in ITP group	66
9	Comparison of the median absolute lymphocytic counts among different genotypes in ITP group	66
10	PCR-RFLP results of CTLA4 A49G polymorphism	68

Acknowledgment

It is great honor to express my deep gratitude and appreciation to Prof. Dr. Shahira Amen Zayed, Professor of Clinical pathology –Faculty of medicine –Cairo University , for her cooperation and continuous guidance. She has sacrificed a lot of her busy time to teach me and revise over step of this thesis.

I would like to express my sincere thanks to Prof .Dr. Somaya Mohammad Elgawhary, Professor of Clinical pathology –Faculty of medicine –Fayoum University, who offered much of her time for providing me with close supervision ,guidance and support. She spared neither time nor knowledge until the end of this work,

I would like to express my sincere thanks to Dr. Hanan AL-Husseini Mohamed , Lecture of Clinical pathology –Faculty of medicine –Cairo University , who sacrificed a lot of her busy time to teach me and revise over step of this thesis .Also she kindly offered me much of her time and experience in the laboratory aspect of this work .

I would like to express my sincere thanks to Dr. Rania Ismail , Lecture of Pediatrics–Faculty of medicine –Cairo University , She offered me much of her time and experience in diagnosis and collection of cases.

I would also like to thank all my patients and their families for their cooperation .

Finally, I acknowledge my family , colleagues, and all those who helped me in this work.

Abstract

Idiopathic thrombocytopenic purpura (ITP) is an acquired autoimmune disease with many immune dysfunctions. Cytotoxic T lymphocyte antigen 4 (CTLA-4) is a T-lymphocyte surface molecule that can down modulate and terminate immune responses. Recently, several studies have confirmed that some polymorphisms of this gene can influence its expression level, therefore speculating that they might be associated with autoimmune diseases. In order to investigate the role of the *CTLA-4* gene in ITP, we investigated A49G polymorphism of the *CTLA-4* gene in 30 ITP patients and 20 healthy controls through polymerase chain reaction (PCR)-restriction fragment length polymorphism. No significant differences were revealed in genotypes and allele distributions between the patients with ITP and the controls.

Keywords: CTLA-4, ITP, T lymphocyte, PCR-RFLP, Polymorphism .

Introduction

Idiopathic thrombocytopenic purpura (ITP) is a **bleeding disorder** in which the immune system destroys platelets, which are necessary for normal blood clotting. ITP is sometimes called immune thrombocytopenic purpura. ITP occurs when certain immune system cells produce **antibodies** against platelets. The antibodies attach to the platelets. The spleen destroys the platelets that carry the antibodies. In children, the disease sometimes follows a viral infection. In adults, it is more often a chronic (long-term) disease and can occur after a viral infection, with use of certain drugs, during pregnancy, or as part of an immune disorder. ITP affects women more frequently than men, and is more common in children than adults. The disease affects boys and girls equally (***McMillan , 2007**).

Autoimmune disorders are the result of disturbed immune tolerance to self-antigens. After the presentation of an antigen in conjunction with human leucocyte antigen (HLA) molecules on the surface of an antigen presenting cell to a specific T-cell receptor (TCR), costimulatory signals are required for the T-cell responses. A number of co-stimulatory molecules are present on the T-cell membrane, which may have stimulatory or inhibitory effects on T lymphocytes. The cytotoxic T lymphocyte associated antigen- 4 (CTLA-4), also known as CD152, is expressed on T lymphocytes and inhibits the T-cell responses (**Teft et al, 2006**).

The CTLA-4 gene is located on chromosome 2 (2q33). Many single nucleotide polymorphisms (SNPs) have been identified in the CTLA-4 gene that contains four exons and three introns. Among them, the –318C > T SNP (rs5742909) located in the promoter region, the 49A > G SNP (rs231775) located in exon 1, and the 6230A > G SNP (CT60, rs3087243) located in 3'-

untranslated region (3'UTR) have attracted more attention (**Chistiakov et al, 2006**).

The A49G polymorphism in exon 1 of the *CTLA-4* gene is especially important because it alters the structure of the CTLA-4 protein by causing Thr17Ala amino acid substitution. It has been suggested that this polymorphism reduces the inhibitory function of CTLA-4. The *CTLA-4* A49G polymorphism was found to be associated with autoimmune diseases such as insulin-dependent diabetes, rheumatoid arthritis, systemic lupus erythematosus (SLE), multiple sclerosis, primary biliary cirrhosis, Hashimoto's thyroiditis, and Graves' disease (**Dallos & Kovacs , 2005**).

Aim of the work

The objective of this study is to examine the genetic association of A49G *CTLA 4* gene single nucleotide polymorphism in patients with idiopathic thrombocytopenic purpura.

Chapter 1

Immune Thrombocytopenic Purpura

Introduction

Immune thrombocytopenic purpura (ITP) is a clinical syndrome in which a decreased number of circulating platelets (thrombocytopenia) manifests as a bleeding tendency, easy bruising (purpura), or extravasation of blood from capillaries into skin and mucous membranes (petechiae) (* **Stasi et al,2008**).

Thrombocytopenia may be subdivided into four grades as follows: grade 1 with a platelet count of 75,000-150,000/ μ L; grade 2 with a platelet count of 50,000-<75,000/ μ L; grade 3 with a platelet count of 25,000-<50,000/ μ L; and grade 4 with a platelet count of <25,000/ μ L. The causative mechanisms of ITP are varied, making ITP a heterogeneous disorder. Thrombocytopenia may be caused by failure of or reduction in platelet production or by increased platelet destruction (**Sekhon & Roy, 2006**).

Classification of ITP

ITP is classified in to two types according to duration of disease :Acute (less than six months) and chronic (6 months or more) (**George et al, 1996**) & (**Cines & Blanchette, 2002**).

Acute ITP usually occurs within few weeks after a viral or bacterial infection in approximately two thirds of patients. It may also follow an immunization such as for measles, mumps and rubella (MMR) . The majority of children with acute ITP (70-90%) spontaneously attain complete remission within 12 months after the initial diagnosis (**Di Paola & Buchanan, 2002**).

The remaining 10-30% of children develop chronic ITP. Some children with chronic ITP improve with the passage of additional time past 6 months from diagnosis. Others, especially those who are more markedly thrombocytopenic, may continue to require repeated treatments, frequent

follow-up and modification of their lifestyle. Some children may continue to respond to, but require, ongoing treatment, while others may never have respond well to treatment(s) or have initially responded to a treatment but then subsequently become refractory to it (**Imbach et al, 1995**).

The term chronic refractory ITP refers to patients with persistent thrombocytopenia, (i.e. platelet counts of 20 000/ μ L or less) for more than 6-12 months and at least minor bleeding manifestation , it occurs in 2-10% children presenting with acute ITP . They may (or may not) have failed splenectomy, and generally require further treatment to increase and sustain their platelet count to avoid bleeding complications. Since management of these patients is complicated and challenging, it creates frustrations for the patient, family and physicians (**Impach et al, 1995**).

Thrombocytopenia is classified as primary, also referred to as idiopathic thrombocytopenic purpura, or as secondary to an underlying disorder . Both malignant and non malignant disorders can be associated with ITP. Within the non malignant disorders a great variety of conditions such as infections, immunodeficiency states, drug treatments (e.g. quinidine, heparin) and autoimmune diseases are included (**Cines & Blanchette, 2002**).

Incidence :

The annual incidence of ITP is estimated to be 5 cases per 100,000 children and 2 cases per 100,000 adults (**Fogarity & Segal, 2007**) , but these data are not from large population-based studies. Most cases of acute ITP, particularly in children, are mild and self-limited and may not received medical attention. Therefore, estimated incidences of acute ITP are difficult to determine and are likely to understate the full extent of the disease (**Segal & Powe, 2006**).