

**Evaluation of the effect of omega-3 fatty acids on the induced hepatotoxicity
of methotrexate in acute lymphoblastic leukemia children**

A Thesis submitted for the fulfillment of master degree in pharmaceutical science
(clinical pharmacy)

By

Reham Kamel Farahat

Bachelor degree in pharmaceutical science ,
Faculty of Pharmacy , Ain Shams University

Under the supervision of

Prof. Dr . Manal Hamed El Hamamsy

Professor of Clinical Pharmacy

Vice Dean for community service and environmental development affairs

Faculty of Pharmacy, Ain Shams University

Prof . Dr . Nancy Samir El Barbary

Assistance professor of pediatrics

Faculty of medicine, Ain shams university

Faculty of Pharmacy

Ain Shams University

2015



سورة المؤمنون

آية : ١٠٥

Acknowledgments

First and fore most thanks to "Allah" the most beneficent and merciful

I wish to express my great appreciation to Prof. Dr. Manal Hamed El Hamamsy Professor of Clinical Pharmacy and Vice Dean for community service and environmental development affairs Faculty of Pharmacy, Ain Shams University for her great support , guidance and encourgment.

Words are not enough to thank Dr. Galila Mohamed Mokhtar professor of pediatrics , Ain Shams University for her great support in this study.

I Would like to express my deepest thanks and gratitude to Prof. Dr. Nancy Samir El Barbary assistant professor of pediatrics Faculty of medicine, Ain shams university , for her generous help and supervision throughout the work.

I Would like to acknowledge with appreciation Dr . Eman Ismail Consultant of clinical pathology Faculty of medicine, Ain shams university.

I wish to express great appreciation and gratitude to patients and their families , nurses and staff of Hematology and Oncology Clinic whose cooperation was indispensable for completion of this work.

Many special thanks for my mum for her kind and sincere help , love and support during the progress of the work.

Finally, I wish to express my great appreciation and gratitude to my family and best friends for their support.

Aim of the work

With the enlarged spectrum of clinical use of methotrexate , its toxicity on the liver has gained much more importance **(Suleyman and Veysef ., 2008)**.

Methotrexate associated hepatotoxicity is a significant clinical problem that affect the compliance with methotrexate containing treatment regimens. Hydrogen peroxide molecules were reported to act as mediators both in the therapeutic and toxic effects of some antineoplastic agents including MTX **(Zhang et al., 1992)**.

A growing body of evidence is emerging which suggest that oxidative stress and reactive oxygen derived free radicals play a crucial role in pathogenesis of MTX induced liver damage **(Ohta et al ., 2007)**.

Omega -3fatty acids are a type of unsaturated fat that are classified as an essential fatty acid .A large body of evidence exists to suggest that omega -3 fatty acids have considerable health benefits including anti-inflammatory effects , antioxidant effects and beneficial effects on cholesterol levels and improving non-alcoholic fatty liver disease **(Begin et al., 1986)**.

The aim of the study :

- 1) To study the role of oxidative stress in MTX induced hepatotoxicity.
- 2) To evaluate the possible hepatoprotective effect of omega-3 fatty acids on methotrexate associated liver injury as evidence by both clinical and biochemical parameters.
- 3) Patient education about disease and drug.

LIST OF CONTENTS

List of Contents

Contents	Page
List of Contents	I
List of Tables	III
List of Figures	IV
List of Abbreviations	V
Abstract	VIII
Introduction	1
Review of literature	
1-Acute lymphoblastic leukemia	4
1.1 Introduction	4
1.2 Risk factors	5
1.3 Classification of ALL	7
1.4 Signs and symptoms	9
1.5 Diagnosis	10
1.6 Prognostic factors	12
1.7 Treatment	13
1.8 Overall outcome for ALL	15
2 - Methotrexate	
2.1 History	17
2.2 Methotrexate mechanism of action	19
2.3 Methotrexate pharmacokinetics	21
2.4 Methotrexate dosage and administration	25
2.5 Clinical resistance mechanism	25
2.6 Methotrexate toxicity	26
2.7 Antidotal strategies for methotrexate toxicity	28
2.8 Methotrexate hepatotoxic effect	33
3- Malondialdehyde	36
4- Antioxidants	37
5- Poly unsaturated fatty acids	
5.1 Chemistry	40
5.2 Dietary sources for different lipids	45

LIST OF CONTENTS

5.3 Pathway for the synthesis of poly unsaturated fatty acids	46
5.4 Mechanism of action	49
5.5 Omega -3 daily requirment	50
5.6 Omega - 3 pharmacokinetics	50
5.7 Omega - 3 adverse effects	52
6-Role of clinical pharmacist	53
Aim of the work	56
Patients and methods	57
Results	64
Discussion	75
Summary	83
References	87
Appendix	105
Arabic summary	1

List of Tables

Tables	Page
Table (1): Risk factors for childhood leukemia	6
Table (2): FAB classification of ALL	7
Table (3): Clinical findings for childhood ALL	9
Table (4): Laboratory features at diagnosis in childhood ALL	11
Table (5): Important prognostic factors and their approximate incidence in childhood ALL	12
Table (6): Chemotherapeutic treatment of ALL	14
Table (7): MTX pharmacokinetics in plasma	24
Table (8): Fatty acids categories	42
Table (9): Measuring procedure for TAC	61
Table (10): Measuring procedure for MDA	62
Table (11): Demographic and clinical data of the enrolled patients	64
Table (12): Weight and height percentile of the enrolled patients	65
Table (13): Baseline laboratory data of the enrolled patients	66
Table (14): Post treatment laboratory data of the enrolled patients	67
Table (15): Comparison between pre- and post-treatment laboratory variables of MTX group	68
Table (16): Comparison between pre- and post-treatment laboratory variables of patients receiving MTX+Omega-3	69
Table (17): Correlation between the studied parameters at 0 month in MTX group	70
Table (18): Correlation between the studied parameters at 6 months in MTX group	70
Table (19): Correlation between the studied parameters at 0 month in MTX+Omega-3 group	72
Table (20): Correlation between the studied parameters at 6 months in MTX+Omega-3 group	72
Table (21): Clinical manifestations of hepatotoxicity in MTX group after six months of treatment	74

List of Figure

Figure	Page
Figure (1) : Survival rates for ALL	15
Figure (2) : 3-D picture of methotrexate molecule	16
Figure (3) : The structural formula of methotrexate	16
Figure (4) : Sites of action of major chemotherapeutic agents	19
Figure (5) : MTX mechanism of action	21
Figure (6) : Antidotal strategies for methotrexate toxicity	30
Figure (7): CPDG2 Mechanism of action	32
Figure (8): Nomenclature for fatty acids	40
Figure (9): Chemical structure of some fatty acids	43
Figure (10): Chemical structure of some fatty acids	44
Figure (11) : Dietary sources for different lipids	46
Figure (12): Synthesis of omega -3 and omega-6 polyunsaturated fatty acids	47
Figure (13): Pathway for synthesis of Very long chain polyunsaturated fatty acids	48
Figure (14) : Route of long- chain omega-3 fatty acids	51
Figure (15): Chemotherapy pharmaceutical care flow sheet	55
Figure (16): Study design	57
Figure(17): Baseline oxidative stress markers among enrolled patients	66
Figure (18): Post-treatment oxidative stress markers among enrolled patients	67
Figure (19): Pre- and post- treatment ALT levels among enrolled patients	69
Figure (20): Correlation between TAC and ALT among patients receiving MTX at 6 months of treatment	71
Figure (21): Correlation between TAC and MDA among patients receiving MTX at 6 months of treatment	71
Figure (22): Correlation between baseline TAC and MDA levels among patients receiving MTX+Omega-3	73
Figure(23): Percentage of the patients in MTX group shown signs and symptoms of hepatotoxicity	74

LIST OF ABBREVIATIONS

List of Abbreviations

AA	Arachidonic acid .
ACCP	American college of clinical pharmacy.
ADE	Adverse drug effect .
ADR	Adverse drug reaction .
AGE	Advanced glycation end product.
AICAR	5-aminoimidazole-4-carboxamide ribonucleotide.
AICART	5-aminoimidazole-4-carboxamide ribonucleotide transformylase enzyme.
ALA	Alpha linolenic acid .
ALE	Advanced lipoxidation end product.
ALL	Acute lymphoblastic leukemia.
ALT	Alanine amino transferase enzyme.
AML	Acute myleoid leukemia.
BM	Bone marrow.
CAT	Catalase
CBC	Complete blood count.
CCG protocol	Children Cancer Group
CMF	Cyclophosphamide , Methotrexate , 5-flurorouracil.
CNS	Central nervous system.
CPDG	Glutamate carboxypeptidase
CSF	Cerebro spinal fluid .
DAMPA	4-amino-4-deoxy-N-methylpteroic acid.
DHA	Docosahexaenoic acid .
DHFR	Di-hydrofolate reductase enzyme.
DHLA	Dihydro lipoic acid.
DMARDs	Disease modifying anti-rheumatic drugs.
DNA	Deoxy nucleic acid.
DPA	Docosapentaenoic acid .
DRP	Drug related problems .
dTMP	Deoxythymidine monophosphate.
dUMP	Deoxyuridine monophosphate .
EBV	Epstein Barr virus.
EPA	Eicosapentaenoic acid.
FAB	French - American - British system .
FAICAR	5-formyl aminoimidazole-4-carboxamide ribonucleotide.

LIST OF ABBREVIATIONS

FBP	Folate binding protein .
FDA	Food and drug administration .
FGAR	Formylglycinamide ribonucleotide.
FH2	Dihydrofolate.
FH4	Tetrahydrofolate.
FPGs	Foly polyglutamate synthase enzyme.
GAR	Glycinamide ribonucleotide.
GGH	Gamma glutamyl hydrosylase.
GHMT	Glycine hydroxymethyl transferase enzyme.
GI tract	Gastrointestinal tract.
GPX	Glutathione peroxidase .
GR	Glutathione reductase .
HDL	High density lipoprotein .
HDL-C	High density lipoprotein cholesterol.
H2O2	Hydrogen peroxide .
HIV	Human immune deficiency virus.
IT	Intra thecal.
IUPAC	International union of pure and applied chemistry.
IV	Intravenous.
JRA	Juvenile rheumatoid arthritis.
LA	Linoleic acid , Lipoic acid.
LNA	Linolenic acid.
LCFA	Long chain fatty acid .
LDL	Low density lipoprotein .
LDL-C	Low density lipoprotein cholesterol.
MAG	Monoacylglyceride.
MCFA	Medium chain fatty acids.
MDA	Malondialdehyde.
MRD	Minimal residual disease .
MTHFR	Methylene tetrahydrofolate reductase enzyme.
MTX	Methotrexate.
MUFAs	Monounsaturated fatty acids.
NADP	Cytosolic nicotinamide adenosine diphosphate dependant dehydrogenase enzyme.
NCI	National cancer institute.
O₂	Oxygen.
PCR	polymeraze chain reaction.
PL-OH	Phospholipid hydroxide.

LIST OF ABBREVIATIONS

PLOOH	Phospholipid hydroperoxide.
PMNs	Polymorphonuclear neutrophils.
P.O.	Per oral.
PPAT	Amido phosphoribosyl transferase enzyme.
PUFA	Poly unsaturated fatty acid.
Rb	Retinoblastoma protein .
RFC	Reduced folate carrier.
RNA	Ribonucleic acid.
ROH	Hydroxyl fatty acid.
ROOH	Lipid peroxides.
ROS	Reactive oxygen species.
SAM	S- adenosylmethionine.
SCFA	Short chain fatty acid.
SOD	Superoxidedismutase.
T_{1/2}	Half life.
TAC	Total antioxidant capacity .
TCR	T-cell receptor.
THA	Tetracosahexanoic acid .
TYMS	Thymidylate synthase enzyme.
UKCS	United Kingdom childhood cancer study.
U.S	United states .
Vd	Volume of distribution.
 VLCFA	Very long chain fatty acids .
WBC	White blood cell.

Abstract

Background: Previous studies have demonstrated that patients receiving oral methotrexate in maintenance phase of acute lymphoblastic leukemia suffer from many side effects, the most is hepatotoxicity . In the other hand there was different studies demonstrated the protective effect of Omega-3 fatty acids on liver cells .

Objective: To evaluate the protective effect of Omega-3 fatty acids on the induced hepatotoxic effect of oral methotrexate in childhood acute lymphoblastic leukemia .

Methods: A prospective, randomized, open labeled, controlled clinical trial, was conducted on 60 Egyptian children with acute lymphoblastic leukemia in maintenance phase. From June 2012 to March 2014. The patients were included into two groups, **group I:** 30 patients received oral methotrexate only (20mg/m²), **group II:** 26 patients received methotrexate (20mg/m²) in combination with omega-3(1000mg/day). Both groups were followed up for six months; data were collected at the beginning of the study and at the end of the study.

Results: At the beginning of the study there was no significant difference between the two groups neither in demographic nor baseline laboratory data, while after six months there was a significant difference ($p<0.001$) between the two groups regarding signs and symptoms of hepatic toxicity, better compliance and no one of the patients stopped the chemotherapy cycle in second group due to liver toxicity all over the study period.

Conclusion: The use of omega-3 fatty acids provides a significant protective effect on liver cells against methotrexate toxic effect upon liver cells. Improvement in quality of life due to absence of liver toxicity which causes fatigue and pain to the patient. The chemotherapy cycle was completed without any interruptions and no signs of hepatic damage was observed.

Key words: Acute lymphoblastic leukemia, Methotrexate, Omega-3, Oxidative stress, Malondialdehyde, Clinical pharmacy.

Introduction

Acute lymphoblastic leukemia:

Acute leukemia represents 97% of all childhood leukemia's .Acute lymphoblastic leukemia (ALL) accounts for about 75% of all childhood leukemia's, and nearly one third of all cancers of pediatric age (*Lanzkowsky and philip.,2005*).

ALL is the most common cancer diagnosed in children and represents 23% of cancer diagnosed among children younger than 15 years .There has been a gradual increase in the incidence of ALL (*Xie et al ., 2003; Ries et al., 2007 and Smith et al., 2007*).

In Egypt, the incidence of childhood cancers overall is approximately 78,010 new patients (as documented in the years 2002- 2005),seen at the NCI ,among which leukemia represents the most common childhood cancer, representing almost 35% of all cases (34.6% boys and 33.3% girls) (*Elattar et al ., 2006*) .

Bone marrow invasion results in anemia ,causing pallor , fatigability, tachycardia, dyspnea , and sometimes congestive heart failure (*Lankzkowsky and Philip., 2005*).

There may be neutropenia, causing fever, ulceration of the buccal mucosa, and infection (*Lankzkowsky and Philip., 2005*).

Among children with ALL , more than 95% attain remission and 75% to 85% survive free of leukemia recurrence at least 5 years from diagnosis with current treatment that in corporatre systemic therapy e.g combination chemotherapy and specific central nervous system preventive therapy (intrathecal chemotherapy) (*Moricke et al ., 2010; Pui et al., 2004*).

The improved survival for children with ALL reflects the impact of well-designed clinical trials conducted during this period, with treatment based on risk assessment (including response to initial treatment) (*Schrappe et al ., 2000*).

Methotrexate:

Methotrexate is a weak dicarboxylic acid with pKa 4.8 and 5.5, and thus it is mostly ionized at physiologic pH (*Widermann et al ., 2004*).

In maintenance therapy of ALL methotrexate is taken orally. Oral administration in tablet form is often preferred when low dose are being administered since absorption is rapid and effective.

ALL in pediatric patients and young adolescents is the most responsive to present day chemotherapy (*Seibel et al., 2008*). The length of maintenance therapy is 3 years for boys and 2 years for girls and adults (*Pui et al., 2004*).

Oral methotrexate is taken once weekly single dose according to the CCG protocol which is used in the Ain shams university blood and oncology clinic .The dose is 20mg / m² (*Hilden et al., 2006*).

Methotrexate (MTX), a folic acid antagonist, is widely used as a cytotoxic chemotherapeutic agent in the treatment of various malignancies such as acute lymphoblastic leukaemia as well as in the treatment of various inflammatory diseases (*Uzar et al., 2006; Cetinkaya et al., 2006*).

The efficacy of this agent is often limited by its toxicity which causes severe side-effects on liver histology related to MTX use and may lead to conditions such as fatty infiltration , liver cirrhosis, fibrosis of the liver, hypertrophy of the hepatocytes, hepatitis, hepato-cellular necrosis and death (*Uzar et al., 2006*).

MTX is known to be an oxidizing agent that suppresses antioxidant system and increases oxygen species in many organs (*Cetiner et al., 2005*).

Prolonged use of MTX leads to accumulation of polyglutamate forms of the drug in hepatocytes, which decreases hepatocellular folic acid levels and eventually leads to hepatocyte necrosis (*Kaplowitz., 2000*).

It is thought that the detrimental effects of MTX is partly due to its direct toxic action by increasing ROS production. It has further been reported that MTX administration induces oxidative stress and significantly reduces antioxidant enzymes such as superoxide dismutase, catalase and glutathione peroxidase in liver, intestinal mucosa and spinal cord tissues of rats (*Uzar et al., 2006; Uz et al., 2005*).

Few studies have demonstrated that methotrexate administration causes cell injury and reduction in the activities of antioxidant enzymes (*Cetinkaya et al., 2006*).

Polyunsaturated fatty acids are much less resistant to peroxidation than monounsaturated or saturated fatty acids . Furthermore , oxidative damage to lipids has widespread effect , as lipid peroxidation triggers a complex chain reaction involving a range of reactive intermediates that can also cause protein and DNA damage (*Hulbert et al., 2007*).

Omega-3 fatty acids:

Essential fatty acids constitute an important component of all cell membranes and influence membrane fluidity and the behavior of membrane-bound enzymes and receptors (*Das ., 2006*).

These fatty acids can modulate physiological and pathological conditions through multiple mechanisms, such as the inflammatory response (*Kang and Weylandt., 2008*), and have received great attention in recent years for their critical role in disease prevention and management (*Gebauer et al., 2006*).

The liver predisposes to oxidative stress presumably by amplifying the capacity of free radical chain reaction. An obvious sign of hepatic injury is the leakage of cellular enzymes into the plasma due to the disturbance caused in the transport functions of hepatocytes. When liver cell membrane is damaged, a variety of enzymes located normally in cytosol is released into the blood (*Sgroc Clinard and Ouazir ., 2002*).

Possible mechanisms that may be responsible for the protection against liver damage by omega-3-Fatty acids includes membrane stabilizing action on the hepatocytes. Although the exact mechanism of cytoprotection by omega-3-fattyacids remains unresolved, yet data suggests that this substance could serve as an antioxidant/cofactor that makes the hepatocytes less susceptible to the damaging action of noxious agents (*Sgroc Clinard and Ouazir ., 2002*).

α -Lipoic acid (LA), a naturally occurring sulphydryl compound found in virtually all plants and animal species, is a potent antioxidant with high efficacy of chemoprotection. It is also involved in the chelation of metal ions, regeneration of exogenous and endogenous antioxidants and repair of oxidized proteins (*Kolgazi et al ., 2007*).

It appears that LA exerts its beneficial effects in conditions where oxidative stress plays a critical role in the induction of cellular damage/target organ toxicity and hence it was considered for the present study and an attempt has been made to evaluate the possible protective effects of LA against MTX-induced toxicity (*Bajin et al ., 2006*).

It may cause gas, bloating, belching, nausea and diarrhea, very high doses may cause some but not so undesirable side effects as fishy body odor and/ or fishy breath (*Buckely et al 2004*), Care should be taken with patients taking blood thinning medications as aspirin and warfarin because these patient when using omega-3 fatty acids may bruise easily or suffer from bleeding disorders (*Buckely et al., 2004*).