Effect of Genetic Polymorphism on the Clinical Response to Valproate Therapy in Epileptic Children

Thesis

Submitted for the Partial Fulfilment of the Philosophy Doctor (PhD) Degree in Pharmaceutical Sciences (Clinical Pharmacy)

By

Pharmacist / Engi Abd-el Hady Abd-el Hady Ibrahim

Master Degree in Clinical Pharmacy Ain Shams University

Under the Supervision of

Prof. Dr. Osama Ahmed Badary

Professor of Clinical Pharmacy
Faculty of Pharmacy
Ain Shams University
Chairman of National
Organization for Drug Control &
Research

Prof. Dr. Mohamad Abd-el Adl Elsawy

Professor of Pediatrics and Genetics Faculty of Medicine

Prof. Dr. Manal Hamed El Hamamsy

Professor of Clinical Pharmacy Vice Dean for Community Services and Environmental Development Affairs Faculty of Pharmacy Ain Shams University

Prof. Dr. Sahar Mohamed Ahmed Hassanein

Professor of Pediatrics Head of Neuropediatric Unit Faculty of Medicine Ain Shams University

Prof. Dr. Manal Zaghlol Mahran

Professor of Clinical Pathology Faculty of Medicine Ain Shams University

2014

Acknowledgements

First and foremost, I am deeply thankful to Allah by the grace of whom, this work was achievable.

I thank my supervisors whose assistance and dedicated involvement in every step throughout this work made it accomplished. I would like to thank them very much for their support and understanding over these past five years.

I am deeply indebted to **Prof. Dr. Osama Ahmed Badary**, Professor of Clinical Pharmacy, Faculty of pharmacy, Ain Shams University, for his earnest support, valuable foresight, and careful guidance throughout the preparation of this study.

I am very grateful to **Prof. Dr. Mohamad Abd-el Adl El-Sawy**, Professor of Pediatrics, Faculty of Medicine, Ain Shams University and former Head of Genetics center, for helping me in selecting the topic, his encouragement, and continuous support.

I would like also to express my sincerest gratitude and appreciation to **Prof. Dr.**Manal Hamed El Hamamsy, professor of Clinical Pharmacy, whose contribution in stimulating suggestions and encouragement helped me to coordinate my work especially in writing this dissertation.

I am very thankful to **Prof. Dr. Sahar Mohamed Ahmed Hassanein**, Professor of Pediatrics and Head of Neuropediatric Unit, Faculty of Medicine, Ain Shams University, for her generous assistance, beneficial discussions and valuable guidance with the patients' facility aspects.

A special gratitude I give to **Prof. Dr. Manal Zaghlol Mahran**, Professor of Clinical Pathology, Faculty of Medicine, Ain Shams University, for her keen supervision and unfailing advice.

Furthermore I would also like to acknowledge with much appreciation the crucial role of the staff of Clinical pathology and immunology laboratory, for their technical and instrumental support to perform the genetic analysis. My deep gratitude goes especially to **Dr. Nesrine Ali Mohamed**, lecturer of Clinical Pathology & Immunology, Faculty of Medicine, Ain Shams University.

I also express my gratefulness to **Prof. Dr. Inas Elattar**, professor of Biostatistics, Department of Biostatistics & Cancer Epidemiology, National Cancer Institute, Cairo University, who kindly gave me the assistance in conducting the statistical analysis and interpretation of data.

Acknowledgements

The patients in whom I performed the study and their parents deserve my sincere gratitude.

Getting through my dissertation required more than academic support, and I would like to thank my colleagues at the department of Clinical pharmacy for their constant personal as well as professional support. I cannot begin to express my gratitude and appreciation for their friendship.

Most importantly, none of this could have happened without my husband, my children, my mother and the rest of my family members. This dissertation stands as a testament to your unconditional love and encouragement.

Table of Contents

Acknowledgements	i
Table of Contents	iii
List of Tables	vi
List of Figures	ix
List of Abbreviations	X
Abstract	xii
Introduction	1
Review of literature	4
Definitions	4
Incidence	4
Pathophysiology	5
Etiology	6
Genetic influences on the epilepsies	8
Classification of epileptic seizures	10
Generalized versus localization-related epileptic seizures	10
1-Generalized seizures	10
2- Focal Seizures	10
Epilepsy syndromes	12
Status Epilepticus	12
Seizure precipitants	13
Risk factors	14
Clinical Presentation and Diagnosis	14
Aura	16
Diagnosis	16
1-Description of events and history taking	16
2-Physical Examination	16
3-Laboratory studies	17
4-Electroencephalograph (EEG)	17
5-Electrocardiogram (ECG)	17
6-Neuroimaging	18
Differential diagnosis of seizures	18
Therapy	18

Table of Contents

Antiepileptic Drug Therapy	19
Mechanism of AEDs	19
Selection of AEDs	20
Basis for selection of AED	20
Dosing consideration	20
Complications of pharmacotherapy	24
1-Adverse Dug effects	24
2-Drug Interactions	27
The Potential of Pharmacogenetics in the Treatment of Epilepsy	30
The contribution of clinical pharmacy to the implementation and integration of	
pharmacogenetics in health care system	30
Genetic factors contributing to AED response	32
(1) Genes encoding drug transporters	32
(2) Genes encoding drug metabolizing enzymes	32
(3) Genes encoding AED targets	33
Valproic Acid	34
Mechanism of action	34
Clinical efficacy	35
Adverse Effects	35
Pharmacokinetics	37
Absorption	37
Distribution and Protein Binding	37
Metabolism	37
Excretion	38
Drug Interactions	39
Clinical Use and Dosing	40
Monitoring of VPA	43
Glucuronidation	44
The UDP Glucuronosyltransferase Gene Family	44
Genetic polymorphisms of UGTs	45
Aim of the work	47
Patients and Methods	48
[1] Patients	49
[2] Methods	50

Table of Contents

Results	66
Discussion	107
Summary and Conclusions	
References	124
Appendix	147
الملخص العربي	

List of Tables

Table 1. Causes of seizure disorders according to age groups	7
Table 2. Genes and loci for pediatric genetic epilepsies	9
Table 3. Types of epileptic seizures	. 1
Table 4. Status epilepticus precipitating events	.3
Table 5. Seizure types and characteristics	.5
Table 6. Differential diagnosis of seizures	.8
Table 7. Evidence-based guidelines for initial monotherapy treatment of epilepsy 2	23
Table 8. Effect of Antiepileptic drugs on hepatic drug metabolizing enzymes2	28
Table 9. Drugs that affect the pharmacokinetics of VPA	1
Table 10. Interactions of VPA with an effect on the pharmacokinetics of other drugs 4	12
Table 11. Checklist for the most common treatment-related adverse drug reactions of	of
valproic acid	52
Table 12. Demographic characteristics of epileptic children	57
Table 13. Seizures types and clinical characteristics of epileptic children	59
Table 14. Treatment characteristics of epileptic children	12
Table 15. Genotype frequencies of UGT1A6 541A>G and 552A>C in epileptic patients of	эf
the study as compared with different populations	15
Table 16. Genotype frequencies of both UGT1A6 541A>G and 552A>C polymorphisms is	in
epileptic patients	¹ 6
Table 17. Allele frequency calculated from observed genotype frequency in epilept	ic
patients7	16
Table 18. Comparison of Age, weight and gender distribution between wild and varian	nt
genotypic groups of UGT1A6 541A>G and 552A>C polymorphisms in epileptic patients 7	18
Table 19. Association of UGT1A6 541A>G and 552A>C genotypes with maximum VP.	A
dosage, C_{ss} and CDR, in epileptic children	19
Table 20. Association of UGT1A6 541A>G and 552A>C genotypic groups with maximum	m
VPA dosage, C _{ss} and CDR, in epileptic children	31
Table 21. Association of allelic groups of UGT1A6 541A>G and 552A>C polymorphism	1S
with maximum VPA dosage, C _{ss} and CDR, in epileptic children	31
Table 22. Seizure characteristics at follow up for epileptic children	3
Table 23. Association of UGT1A6 541A>G and 552A>C genotypic groups with seizur	re
frequency in epileptic children	33

Table 24. Association of UGT1A6 541A>G and 552A>C genotypes with seizure severity
score on Chalfont scale, in epileptic patients
Table 25. Association of UGT1A6 541A>G and 552A>C genotypic groups with seizure
severity score on Chalfont scale, in epileptic patients
Table 26. Frequency of adverse drug reactions of valproic acid experienced by epileptic
children
Table 27. Association between genotypic groups of UGT1A6 541A>G and fatigue, difficulty
in concentration and/ poor scholastic achievement and hyperactivity as ADR to VPA 90
Table 28. Association between genotypic groups of UGT1A6 552A>C and fatigue, difficulty
in concentration and/ poor scholastic achievement and hyperactivity as ADR to VPA 91
Table 29. Association between genotypic groups of UGT1A6 541A>G and decreased
appetite +/- poor weight gain, alopecia +/- curling and insomnia, as ADR to VPA
Table 30. Association between genotypic groups of UGT1A6 552A>C and decreased
appetite +/- poor weight gain, alopecia +/- curling and insomnia, as ADRs to VPA94
Table 31. Association between genotypic groups of UGT1A6 541A>G and drowsiness,
obesity with/without increased appetite, irritability and headache, as ADRs to VPA 96
Table 32. Association between genotypic groups of UGT1A6 552A>C and drowsiness,
obesity with/without increased appetite, irritability and headache, as ADRs to VPA 97
Table 33. Association between wild type and variant genotypic groups of UGT1A6 541A>G
and UGT1A6 552A>C polymorphisms, and the number of ADRs per patient
Table 34. Laboratory data at baseline and follow up, for epileptic children
Table 35. Effect of co-medication with 1-carnitine on seizure frequency in epileptic children
Table 36. Effect of co-medication with 1-carnitine on the frequency of headache, irritability
and hyperactivity as ADRs to treatment with VPA in epileptic children 101
Table 37. Effect of co-medication with 1-carnitine on the frequency of drowsiness, fatigue
and insomnia as ADRs to treatment with VPA in epileptic children
Table 38. Effect of co-medication with 1-carnitine on the frequency of decreased appetite +/-
poor weight gain, alopecia+/- curling and obesity+/- increased appetite as ADRs to treatment
with VPA in epileptic children
Table 39. Effect of co-medication with 1-carnitine on the frequency of difficulty in
concentration and/ poor scholastic achievement as an ADR to treatment with VPA in
epileptic children 104

List of Tables

Table 40. Correlation between serum VPA concentration and daily VPA dose,	number of
seizures per month (during 6 months follow up), number of ADRs per patient ar	d Chalfon
seizure severity score	106

List of Figures

Figure 1. A timeline of antiepileptic drugs
Figure 2. Overview of antiepileptic drug treatment response
Figure 3. Pathways of valproic acid elimination with candidate genes involved in metabolism
39
Figure 4. Glucuronidation reaction of valproic acid
Figure 5. UGT1 gene locus consisting of four common exons, nine first exons and four
pseudogenes
Figure 6. Schematic representation of the study design
Figure 7. Chalfont Seizure Severity Scale
Figure 8. Distribution of different seizure types among epileptic children
Figure 9. Valproic acid brands used in the treatment of epileptic children
Figure 10. Electrophoresis pattern of PCR fragments after digesting with NsiI for the
UGT1A6 541A>G polymorphism
Figure 11. Electrophoresis pattern of PCR fragments after digesting with Fnu4HI for the
UGT1A6 552A>C polymorphism
Figure 12. Adverse drug reactions of valproic acid experienced by epileptic children 88
Figure 13. The response and prognosis of epileptic children to valproic acid therapy 105

List of Abbreviations

AAN American Academy of Neurology.

ABCB1 ATP-Binding Cassette, Sub-Family B, Member 1

ADR Adverse Drug Reaction

AEDs Antiepileptic Drugs

AUC Area Under the Plasma Concentration-Time Curve

BID bis in die, twice a day

CDR Concentration to Dose RatioCT Computerized Tomography

CYP P450 Cytochrome P450

DNTP Deoxynucleotide Triphosphate

EDTA Ethylenediaminetetraacetic Acid

EEG Electroencephalography

EPC Epilepsia Partialis Continua

FLAIR Fluid-Attenuated Inversion Recovery

G6PDH Glucose-6-Phosphate Dehydrogenase

GABA-A γ-Aminobutyric Acid, Subunit A

GCSE Generalized Convulsive Status Epilepticus

ILAE International League Against Epilepsy

LD Linkage Disequilibrium

MDR Multi-Drug Resistance Gene

MRI Magnetic Resonance Imaging

NAD Nicotinamide Adenine Dinucleotide

NICE The National Institute For Clinical Excellence In The United Kingdom.

PCR Polymerase Chain Reaction

PCR-RFLP Polymerase Chain Reaction-Restriction Fragment Length Polymorphism

PT Prothrombin Time

PTT Partial Thromboplastin Time

SCN1A Sodium Channel Gene

SD Standard Deviation

SIGN Scottish Intercollegiate Guidelines Network.

SMA Supplementary Motor Area

SNP Single Nucleotide Polymorphism

List of Abbreviations

TAE Tris-Acetate-EDTA

TIA Transient Ischemic Attack

TID ter in die, three times a day

TNF Tumor Necrosis Factor

UDGPA Uridine 5´-Diphosphate-Glucuronic Acid

UGT Uridine Diphosphate Glucuronosyltransferase

VPA Valproic Acid

Abstract

Background/objective: Valproic acid (VPA) is widely used in pediatric epilepsy. It is mainly eliminated through conjugation by UDP-glucuronosyltransferases which are known to be polymorphic. The aim of the study was to investigate the effect of UGT1A6 polymorphism at 541A>G and 552A>C loci on the serum level of VPA and overall clinical response. Seizure control and incidence of adverse drug reactions (ADR) were investigated in a cohort of Egyptian children with idiopathic epilepsy.

Methods: Genetic polymorphisms were detected in 48 patients receiving VPA monotherapy by PCR-RFLP. Steady state concentrations at trough level were determined by homogenous enzyme immunoassay technique. All patients were monitored for seizure frequency and seizure severity (SS) using Chalfont SS scale as well as for ADRs.

Results: Patients of variant genotype group (AC & CC) had lower concentration dose ratios (CDRs) than those with (AA) genotype for UGT1A6 552A>C (p=0.029). For both UGT1A6 541A>G and 552A>C, the median CDR of variant allele carriers was significantly lower than wild-type allele carriers (p= 0.047 and p= 0.001, respectively). Higher SS scores on Chalfont scale were associated with (AA) genotype of UGT1A6 552A>C than variant genotypes group (AC & CC) (p=0.020). No significant effect was detected on seizure control, while fatigue and cognitive adverse effects were significantly higher in wild genotype group of UGT1A6 552A>C and variant genotype group of UGT1A6 541 A>G, respectively.

Conclusion: UGT1A6 polymorphisms at 541A>G and 552A>C loci may be associated with increased VPA metabolism in Egyptian epileptic children. The severity of seizures as well as susceptibility to certain ADRs may be linked to the presence of certain UGT1A6 genotypic variants.

Introduction

Epilepsy is a chronic neurological condition manifesting as recurrent, unprovoked epileptic seizures. It is one of the most common chronic neurological diseases, with an estimated 50 million people affected worldwide (*Duncan et al.*, 2006). Epilepsy is an extremely heterogeneous disorder, comprising a large spectrum of different seizure and syndrome types with multiple underlying etiologies (*Bhalla et al.*, 2011).

Antiepileptic drugs (AED) are the primary form of treatment for seizures and epilepsy, while brain surgery and vagal nerve stimulation are reserved for selected refractory cases (*Elger and Schmidt*, 2008). Pharmacotherapy is fraught with problems mainly related to the unpredictability of efficacy, adverse drug reactions (ADRs) and optimal dosing in individual patients (*Depondt*, 2008).

The choice of drug and initial dosing is mainly based on factors such as epilepsy type, patient's age, gender, co-medication, and concomitant diseases (*Elger and Schmidt*, 2008). Further dose adjustments are based on seizure frequency and occurrence of ADRs. This may consume a considerable time of trial and error before an acceptable balance is achieved between efficacy and toxicity (*Berg and Chadwick*, 2000; *Loscher et al.*, 2009).

AED efficacy, toxicity and dosing are influenced by multiple factors including environmental factors, patient-related factors, as well as genetic factors (*Evans and McLeod*, 2003). Identification of genetic factors influencing AED response could enable prediction of response in individual patients which is the essense of the pharmacogenetic studies. This could lead to more rapid seizure control with fewer ADRs and thus to an improved quality of life for patients with epilepsy (*Loscher et al.*, 2009).

Genetic association studies are currently the most widely used approach where correlations between genetic variants and phenotypical differences are assessed on a population scale, to identify genetic variations contributing to variable drug responses (*Goldstein et al.*, 2003). Increased knowledge of genetic associations with drug response is becoming more available making it possible that genetic information could be commonly used to guide drug therapy decisions in the near future (*Johnson*, 2013).

Clinical pharmacists have an established role in the health care setting, including education for both patients and providers, selecting and monitoring drug therapies for individual patients, ensuring safe and appropriate use of medications in populations, as well as conducting clinical research (*El-Ibiary et al.*, 2008). Pharmacogenetic information serves

as a potential tool providing unique opportunities for clinical pharmacists to expand these roles, thereby, optimizing drug treatment for individual patients (*Brock et al.*, 2003).

Valproic acid (VPA) is one of the major antiepileptic drugs with high efficacy against multiple seizure types, including both primarily generalized and partial seizure in adults and children (*Loscher*, 2002). Serious adverse effects, despite being rare, including fatal hepatotoxicity, acute pancreatitis, encephalopathy as well as bone-marrow suppression, have been associated with VPA treatment (*Gerstner et al.*, 2007).

Therapy with VPA is clinically complicated by high interindividual variability in both pharmacokinetics and pharmacodynamics giving rise to a wide dosing range and therapeutic plasma level of 50-100 mg/l which necessitates its blood level monitoring during therapy (*Jiang and Wang*, 2004; Ferraro and Buono, 2005).

Glucuronidation and β-oxidation are the principal pathways of VPA metabolism, with glucuronidation reaching up to 50% of the total metabolism of the initial dose (*Ito et al.*, 1990; Argikar and Remmel, 2009). Glucuronidation involves the conjugation of a glucuronic acid moiety to a range of functional groups of a specific substrate increasing their polarity which facilitates their excretion in bile or urine (*Guillemette*, 2003). It is carried out by Uridine diphosphate glucuronosyltransferase (UGTs), a superfamily of enzymes (>16) expressed on the inner membrane of the endoplasmic reticulum and are categorized into three subfamilies: UGT1A, UGT2A, and UGT2B (*Mackenzie et al.*, 1997). Glucuronidation of VPA has been reported to be carried out by UGT1A3, UGT1A6, UGT1A9, UGT2B7, and UGT2B15 (*Ethell et al.*, 2003; Argikar and Remmel, 2009).

UGT1A6 gene was demonstrated to be highly polymorphic with at least four alleles characterized by three single nucleotide polymorphisms (SNPs) including polymorphisms rs2070959 (541A>G) and rs1105879 (552A>C), rs6759892 (19T>G) in the coding sequence which can lead to both transcriptional and functional changes of its encoded enzymes (*Ciotti et al.*, 1997; Nagar et al., 2004). Genetic polymorphisms in UGT1A6 are highly prevalent in different racial populations and the frequency of some variant alleles has shown different distribution between Caucasians and Asians (*Lampe et al.*, 1999).

Studies investigating the clinical implications of these variant alleles are of great importance for clinical practice as they help assess the required dose of VPA in different populations.

Few studies are available on the effect of UGT1A6 gene polymorphisms on the levels of VPA in epileptic patients. All of them were carried out in Asian populations of different ethnic backgrounds; moreover, the results generated from these studies were not entirely