SERUM SELENIUM LEVEL, LIPID PEROXIDATION AND GLUTATHIONE PEROXIDASE ACTIVITIES IN PATIENTS WITH ACUTE LYMPHOBLASTIC LEUKEMIAS PRE-AND POST-INDUCTION CHEMOTHERAPY

A Thesis

Presented to The Medical Research Institute Alexandria University In Partial Fulfillment of the Requirements for the Degree

of

Master

in

Applied Medical Chemistry

By

Nahla Abdel Hakam Ahmed

B.Sc, (Special Chemistry), Faculty of Science, El Mansoura University, 1998

Advisor's committee

Prof. Dr. Safinaz Mahmoud El-Zoghbi Professor of Applied Medical Chemistry Medical Research Institute University of Alexandria	•••••••••••••••••••••••••••••••••••••••
Prof. Dr. Afaf Aly Hassan El-Faras Professor of Physiology Medical Research Institute University of Alexandria	
Dr. Yosria El-Sayed Ali Assistant Consultant of Physiology Medical Research Institute University of Alexandria	
Dr. Samia Abd El-Wahab Mohammed Colleague of Haemtology Medical Research Institute University of Alexandria	

ACKNOWLEDGEMENT

Thanks to **God** all mighty for the help and endurance offered to me to accomplish this work

It is with great pleasure that I record my special appreciation to **Prof. Dr. Safinaz El-Zoghby** Professor of applied medical chemistry. Medical Research Institute. University of Alexandria for her close supervision encouragement and support.

I am also a greatly indebted to **Prof. Dr. Afaf El-Fares** Professor of physiology. Medical Research Institute. University of Alexandria for her sincere criticism, guidance, and precious advice.

My appreciation also extends to **Dr. Yousria El-Sayed Ali** Assistant Consultant of Physiology. Medical Research Institute. University of Alexandria for her help, encouragement, kindness and continuous support during the study.

I am sincerely thankful to **Dr. Samia Abdel Wahab** Colleague of haematology. Medical Research Institute. University of Alexandria for her great help and instructions throughout this work.

I am of great thankful for **Dr**. **Nesreen Essam El-Din Shaheen** Colleague of applied medical chemistry. Medical Research Institute. University of Alexandria for her helping in the practical work of this study.

I am greatly thankful to all personnel in applied medical department for providing facility the work of this study and co-operation and support.

At last but not least, my faithful gratitude to my family and friends for their continuous helps and support in all my life.

LIST OF ABBREVAIATION

ADP : Adenosine Di Phosphate

AIDS : Acquired Immunodeficiency syndrome

ALL : Acute Lymphoblastic Leukemia

AML : Acute Myeloid Leukemia

ATP : Adenosine Tri Phosphate

BM : Bone Marrow

CAT : Catalase

CD : Cluster of differentiation

COX : Cycloxygenase

CSF : Cerebrospinal Fluid

Cyt Ig : Cytoplasmic immunoglobulin

DTNB : 5,5'-dithiobis-(2-nitrobenzoic acid)

DV : Daily Value

EDTA : Ethylene diamine tetra acetic acid

eNO : Endothelial nitric oxide

FAB : French - American - British

FAD : Flavin Adenine Dinucleotide

FDA : Food and Drug Administration

FR : Free Radical

GPX : Glutathione Peroxidase

GSH : Glutathione

GSHr : Glutathione reductase
GSSG : Glutathione disulfide

GST : Glutathione-S-transferase

H₂O₂ : Hydrogen peroxide

H₂Se : Hydrogen selenide

HIV : Human Immunodeficiency Virus

HLA-DR : Histocompatibility antigens

HOCI : Hypochlorous acid

iNOS : Inducible nitric oxide synthase

kDa : Kilo Dalton

L1-L3 : Categories of acute Lymphoblastic leukemia

LOO : Lipid alkoxyl radical : Lipid peroxyl radical

LOX : Lipoxygenase

M0-M7 : Categories of acute myeloid leukemia

MAbs : Mono Clonal Antibodies

MDA : Malondialdehyde

MIC classification : Morphology, Immunophenotyping and Cytogenetic classification

MPO : MyeloperoxidaseNADPH : Reduced NADP

NFKB : Nuclear Factor-kappa B

NK : Natural Killer cells

nNO : Neural tissue nitrogen oxide

Non Se-GPX : Se-independent Glutathione peroxidase

NOS : Nitrogen oxide synthase

NO_x : Oxides of nitrogen

NSE : Non-specific esterases

ONOO : Peroxynitrite

OS : Oxidative Stress

8-OHdG : 8-Hydroxy deoxyguanosine

PAS : Periodic Acid Schiff

PB : Peripheral Blood

PBS : Phosphate Buffer Saline

PUFA : Poly Unsaturated Fatty Acids

RDA : Recommended Dietary Allowance

RNS : Reactive Nitrogen Species

ROOH : Organic hydroperoxide

ROS : Reactive Oxygen Species

SBB : Sudan black B

SDS : Sodium Dodecyl Sulphate

Se : Selenium

Se IW : Selenoprotein W

Sec : Selenocysteine

Se-GPX : Se-dependent Glutathione peroxidase

Non Se-GPX : Se-independent Glutathione peroxidase

SeP : Selenoprotein P

SmIg : Surface membrane immunoglobulin

SOD : Super Oxide Dismutase

SPPS : Statistical Package for Social Sciences

TBA : Thiobarbituric Acid

TCA : Trichloroacetic acid

TdT : Terminal Deoxynucleotidly transferase

tGPX : Total Glutathione Peroxidase Activity

TMP : Tetramethoxypropane

TNF-α : Tumor Necrosis Factor- α

tRNA : Transfer RNA

LIST OF CONTENTS

Chapter
ACKNOWLEDGEMENT
INTRODUCTION1
I. Leukemia1
II. Free radicals 6
1. Production of free radicals
2. Oxidative stress
3. Lipid peroxidation
4. DNA
5. Protein
6. Scavenging of free radicals
III. Antoxidant
1. Antioxidant and cancer
2. Glutathione Peroxidase
3. Selenium
AIM OF THE WORK26
SUBJECTS AND METHODS
RESULTS35
DISCUSSION63
SUMMARY
REFERENCES70
ARABIC SUMMARY
PROTOCOL

LIST OF FIGURES

Figure		Page
(1)	Production of oxygen and nitrogen free radicals and other reactive species in mammalian cells.	9
(2)	Lipid peroxidation chain reaction.	12
(3)	Removal of oxygen and nitrogen free radicals and other reactive species in mammalian cells.	16
(4)	Antioxidant groups and actions.	17
(5)	Exogenous antioxidants.	19
(6)	A metabolic scheme showing Se metabolism.	24
(7)	Standard curve of malondialdehyde (MDA).	29
(8)	Se standard addition calibration curve.	32
(9)	Hg level in children with ALL (at diagnosis and after remission), and healthy control groups.	44
(10)	RBCs count in children with ALL (at diagnosis and after remission), and healthy control groups.	44
(11)	WBCs count in children with ALL (at diagnosis and after remission), and healthy control groups.	45
(12)	Platelets count in children with ALL (at diagnosis and after remission), and healthy control groups.	45
(13)	Peripheral lymphocyte cells in ALL (at diagnosis and after remission), and healthy control groups.	46
(14)	Bone marrow lymphocyte cells in children with ALL (at diagnosis and after remission).	48
(15)	Bone marrow blast cells in children with ALL (at diagnosis and after remission).	48
(16)	Serum level of MDA (nmol/ml) in children with ALL (at diagnosis and after remission) and healthy control group.	51
(17)	Serum total glutathione peroxidase activity (tGPX) IU/ml in children with ALL (at diagnosis and after remission) and healthy control group.	54
(18)	Serum selenium dependent glutathione peroxidase activity IU/ml in children with ALL (at diagnosis and after remission) and healthy control group.	56
(19)	Serum non- selenium dependent glutathione peroxidase activity IU/ml in children with ALL (at diagnosis and after remission) and healthy control group.	58
(20)	Serum level of selenium ($\mu g/L$) in children with ALL (at diagnosis and after remission) and healthy control group.	61
(21)	Relation between Se ($\mu g/L$) and Hb in children with ALL after remission	62
(22)	Relation between MDA (mmol/ml) and blast cells count in bone marrow in children with ALL at diagnosis.	62



LIST OF TABLES

Table		Page
(1)	A summary of FAB classification of acute leukemia.	2
(2)	A summary of FAB classification of acute lymphoblastic leukemia.	4
(3)	Immunophenotyping of B-ALL subtypes.	5
(4)	Immunophenotyping of T-ALL subtypes.	5
(5)	Summary of antioxidants defenses present in biological system.	18
(6)	Selected food sources of selenium.	22
(7)	Different concentrations of malondialdehyde which was used in standard curve.	29
(8)	Different added concentrations of Se which used in addition standard curve.	32
(9)	Peripheral Hematological Profile in Control Healthy Subjects.	36
(10)	Peripheral Hematological Profile in Acute Lymphoblastic Leukemia (ALL) Patients at Diagnosis.	37
(11)	Bone Marrow Profile in Acute Lymphoblastic Leukemia (ALL) Patients at Diagnosis.	38
(12)	Peripheral Hematological Profile in Acute Lymphoblastic Leukemia (ALL) Patients after Remission.	39
(13)	Bone Marrow Profile in Acute Lymphoblastic Leukemia (ALL) Patients after remission.	40
(14)	Descriptive data of Peripheral blood findings.	43
(15)	Descriptive data of Bone Marrow findings.	47
(16)	Serum level of Lipid Peroxidation (MDA) nmol/ml.	50
(17)	Serum level of Total Glutathione Peroxidase Activity (tGPX) IU/ml.	53
(18)	Serum level of Se-dependent Glutathione peroxidase (Se-GPX) IU/ml.	55
(19)	Serum level of Se-independent Glutathione peroxidase (Non Se-GPX) IU/ml.	57
(20)	Serum Selenium Levels (µg/L).	60

INTRODUCTION

I-LEUKEMIA

Acute leukemias are a heterogeneous group of neoplasms affecting uncommitted or partially committed hematopoietic stem cell ⁽¹⁾. They could be defined as malignant proliferation of hematopoietic progenitor cells involving primarily the peripheral blood (PB) and the bone marrow (BM) leading to increased numbers of blast cells at the expense of normal elements⁽²⁾. They have been recognized as a genetic disease, resulting from a series of acquired or inherited mutations in the structure of certain genes⁽³⁾. These mutations are passed from the original transformed progenitor cells to its clonal descendants. Most of the genetic aberrations fall into genetic classes of functional dysregulation that subvert normal hematopoietic developmental programs by circumvention of cell cycle controls, inhibition of differentiation and resistance to therapeutic apoptosis in leukemic blasts ⁽⁴⁾.

The disease can be originated from lymphoid cells of different lineage, giving rise to B- or T- cell leukemias or sometimes mixed-lineage leukemia. It is a curable disease with an expected long term survival rate of at least 70%, when treated with modern therapeutic regimen. In general, ALL standard treatment protocols consist of induction and maintenance remission with chemotherapeutic drugs ⁽⁵⁾.

The diagnosis of acute leukemia usually follows the presentation of a patient with clinical features suggestive of this disease, with relevant laboratory evaluation being performed. In some patients, diagnosis of acute leukemia can be made from a blood count and blood film but in others bone marrow aspiration is essential for diagnosis, for further classification and to provide optimal material for cytogenetic analysis. Cases with hypocellualr or fibrotic BM, only trephine biopsy permits a definitive diagnosis ⁽⁶⁾.

Classification

A uniform classification system for acute leukemias was developed by an international group of investigators in 1976 known as French - American -British (FAB) classification, this system is based on the morphologic appearance of bone marrow and blood leukemic blasts in Romanovsky -stained smears, supplemented when necessary with cytochemical stains ⁽⁷⁾. Selective cytochemical stains improve the accuracy and reproducibility of lineage assessment and acute myeloid leukemia (AML) subclassification. Myeloperoxidase (MPO), Sudan black B (SBB) and Non-specific esterases (NSE) are useful in this regard ^(8,9).

Table (1): A summary of FAB classification of acute leukemia (4)

Categories of AML	Categories of ALL
M0, AML with minimal evidence of myeloid differentiation	L1, ALL with fairly small uniform lymphoblasts
M1, AML without differentiation	L2, ALL with more pleomorphic lymphoblasts
M2, AML with differentiation	L3, ALL with basophilic vacuolated lymphoblasts
M3, AML and M3 variant, acute promyelocytic leukemia and its hypogranular/microgranular variant	
M4, AML acute myelomonocytic leukemia	
M5, acute monoblastic (M5a) or monocytic (M5b) leukemia	
M6, acute leukemia with at least 50% erythroblasts in the bone marrow	
M7, acute megakaryoblastic leukemia	

Cytochemistry:

- The MPO reaction: is positive in cells of granulocytic series and weakly positive in monocytes, while lymphocytes and erythriod precursors are negative.
- SBB stains: the pattern of staining closely parallels MPO, but SBB positivity is usually stronger than MPO.
- NSE activity: is found in monocytes.
- A periodic acid schiff (PAS): ALL lymphoblasts often have prominent PAS staining in the form of coarse granules or blocks. Myeloblasts may be positive or negative. The pattern of staining is faint and diffuse.
- Acid phosphatase: is not useful in diagnosis of acute leukemia (10).

The definition of acute leukemia and the distinction between AML and ALL according to the FAB system are based on 2 criteria:

- 1- Percentage of blasts in the bone marrow is > 30% of all nucleated cells.
- 2- Three percentage or more (\geq 3%) of blasts show a positive reaction for MPO or SBB in the bone marrow (11).

When facilities are limited, acute leukemia can be classified on the basis of cytology and cytochemistry, as initially proposed by the FAB group. This means that cases of AML with minimal evidence of myeloid differentiation could be misclassified as ALL. If cytology and cytochemistry are supplemented by immunophenotyping in all cases that are not obviously myeloid, the diagnosis will be further refined. Also cases of acute biphenotypic leukemia will remain unrecognized without extensive use of immunophenotyping (12).

Introduction

The optimal approach to the classification of acute leukemia is based on a combination of morphology, immunophenotyping and cytogenetic analysis (MIC classification), or with the addition of molecular genetic analysis (MIC M classification). This approach has been adopted by the World Health Organization (WHO classification) (13). Thus WHO classification incorporates and interrelates morphology, cytogenetic, molecular genetics and immunologic markers in an attempt to construct a classification that is universally applicable and prognostically valid (14).

The definition of acute leukemia according to WHO classification include lowering of the threshold for the percentage of blast cells to 20% in the blood or bone marrow (12).

Immunophenotyping

The impact of immunophenotyping by flow cytometry in the diagnosis and management of acute leukemia has expanded rapidly. This advance can be attributed mainly to the production of several hundred monoclonal antibodies (MAbs) labeled with flurochromes to a variety of antigens expressed by hematopoietic cells ⁽¹⁵⁾.

The panel used includes at least one highly sensitive marker (CD19 for B-lineage cells, CD7 for T- lineage cells, and CD 13 or CD33 for myeloid cells) and one marker that are highly specific cytoplasmic CD79a for B-lineage cells, cytoplasmic CD3 for T-lineage cells, and cytoplasmic myeloperoxidase for myeloid cells ⁽¹⁶⁾.

Cytogenetic study:

Genetic analysis is a diagnostic tool in acute leukemia. The results contribute to the confirmation of the diagnosis, but, more important, the karyotype of the leukemic blasts gives important information concerning the prognosis of the disease ⁽¹⁷⁾.

Molecular genetic analysis has an advantage over conventional cytogenetic analysis that it can yield results when cytogenetic analysis has failed or when only normal metaphases are seen (18).

Acute Lymphoblastic Leukemia (ALL)

ALL is a malignant disorder that originates in a single B- or T-lymphocyte progenitor. The proliferation and accumulation of blast cells in the marrow results in suppression of hematopoiesis and, therefore, anemia, thrombocytopenia and neutropenia. Extramedullary accumulation of lymphoblasts may occur in various sites especially the meninges, gonads, liver, spleen or lymph nodes ⁽¹⁹⁾.

ALL is the most common malignancy diagnosed in patients under the age of 15 years accounting one quarter of all cases, 76% of all leukemias are in this age group (20).

Clinical Features:

Clinical presentation is variable reflecting marrow failure and extramedullary spread.

About one half of patients present with fever which occurs either from cytokines released from leukemic cells or from infection regardless of its origin.

Introduction

Fatigue and lethargy are frequent manifestations of anemia. More than one quarter of patients may present with limp, bone pain, arthralgia or refusal to walk owing to leukemic infiltration of the periosteum, bone or joint or expansion of marrow cavity by leukemic cells (21-23).

Laboratory Features:

Anemia, neutropenia and thrombocytopenia are common findings in patients with newly diagnosed ALL, and their severity reflects the degree of marrow replacement by leukemic lymphoblasts.

Hyperleucocytosis (> $100 \times 10^9 / L$) occur in 10-16 % of the patients ⁽²¹⁾.

Serum lactate dehydrogenase and serum uric acid are usually elevated, serum immunoglobulin levels are markedly decreased reflecting decreased number and impaired function of normal lymphocytes ⁽²⁴⁾.

CSF (Cerebrospinal Fluid) examination is an essential diagnostic procedure since leukemia blast cells may be identified in CSF at diagnosis⁽²⁵⁾.

FAB classification of ALL:

Three subtypes of ALL are distinguished on the basis of cell size, nuclear shape, prominence of nucleoli and relative amount of cytoplasm.

- L1 lymphoblasts \rightarrow are small with high nucleocytoplasmic ratio.
- L2 lymphoblasts → larger blasts with moderatory abundant cytoplasm.
- L3 lymphoblasts → the largest with abundant intensely basophilic cytoplasm (25)

Table (2): A summary of FAB classification of acute lymphoblastic leukemia (10)

	L1	L2	L3
Cell Size	Small	Large	Large
Nuclear chromatin	Fine or clumped	Fine	Fine
Nuclear shape	Regular, may have a cleft or indentation	Irregular, may have cleft or indentation	Regular, oval to round
Nucleoli	Indistinct or not visible	One or more per cell large prominent	Once or more cell large prominent
Amount of cytoplasm	Scanty	Moderately abundant	Moderately abundant
Cytoplasmic basophilia	Slight	Slight	Prominent
Cytoplasmic vacuoles	Variable	Variable	Prominent

Introduction

Immunologic classification of ALL:

Monoclonal antibodies allowed the subclassification of ALL into T and B lineage and each of these can be further subdivided according to the expansion of certain maturation antigen molecules $^{(26)}$.

Table (3): Immunophenotyping of B-ALL subtypes (27).

ALL-subtype	Tdt	HLA-DR	CD19	CD10	Cyt Ig	SmIg
Pro B or early B	+	+	+	ı	_	_
Common ALL	+	+	+	+	_	_
Pre-B ALL	+	+	+	+	+	_
B-ALL	-	+	+	-/+	_	+

Table (4): Immunophenotyping of T-ALL subtypes (27).

ALL_ subtypes	Tdt	HLA_DR	CD2	CD5	CD7	CD4	CD8	CD3
Pre-T ALL	+	+	I	I	+	I	_	ı
Early T- ALL	+	-	+	+	+	_	_	-
Cortical T-ALL	+	_	+	+	+	+	+	-
Late T-ALL	+	_	+	+	+	+	+	+