# BIOLOGICAL STUDY OF HIGH RISK HPV IN LARYNGEAL CARCINOMA

#### Thesis

Submitted for partial fulfillment of the doctoral degree in otorhinolaryngology

By Tamer M. EL-Sayed Seleem Azab (M.Sc)

#### Supervised by

Prof. Dr. Mohammed Nassar Abd EL-Raheem Nassar

Professor of Otorhinolaryngology

Faculty of Medicine — Ain Shams University

Prof. Dr. Mohammed Adel Ismael Lotfy

Professor of Otorhinolaryngology

Faculty of Medicine — Ain Shams University

Prof. Dr. Samar K. Abd EL-Hameed Kassem

Professor of Biochemistry and Molecular Biology

Faculty of Medicine — Ain Shams University

Dr. Aly M. Nagy Aly EL-Makhzangy

Lecturer of Otorhinolaryngology

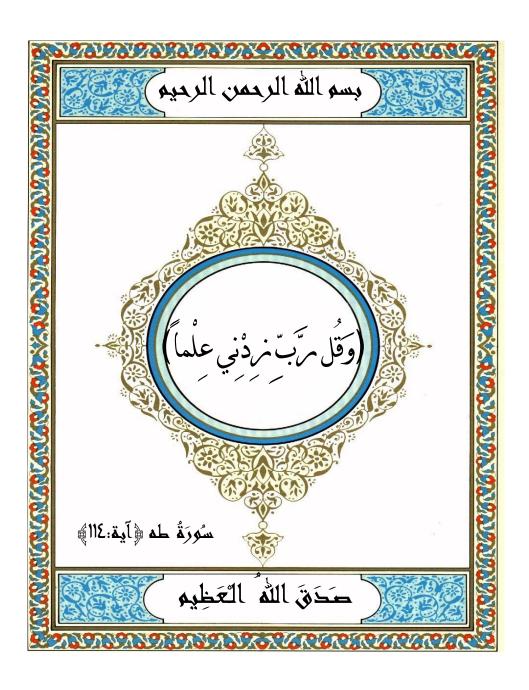
Faculty of Medicine — Ain Shams University

Dr. Hosam M. Kamal Rabea

Lecturer of Otorhinolaryngology

Faculty of Medicine — Ain Shams University

Faculty of Medicine Ain Shams University



### **Acknowledgement**

With this work ready for evaluation and criticism, I pray to God Almighty thankfully and hopefully for multiplying its value and use.

I wish to express my sincere thanks to **Professor Dr. M. Nassar A. Nassar**, Professor of Otorhinolaryngology, Faculty of Medicine, Ain Shams University. His guidance with the steps of this work from the very beginning of idea generation and all the way to the accomplishment of the aim, saving no effort or time for this end, has always been enlightening.

I also would like to re-acknowledge **Professor Dr. Professor Dr. M. Adel Lotfy** Professor of Otorhinolaryngology, Faculty of Medicine, Ain Shams University whose patience and perseverance were fruitful

I extend my deep thanks and gratitude to **Professor Dr. Samar Kamal**, Professor of Biochemistry, Faculty of Medicine, Ain Shams University, for her relentless care, and scientific advice on the various problems that I had to go through to attain the goal of this research.

**Dr. Aly EL-Makhzangy** lecturer of otorhinolaryngology generously provided much of his precious time and devoted effort to contribute to the process of production of the present work. My expressions of gratitude for his generosity shall be always lesser than what he deserves.

Many thanks go to **Dr. Hosam Rabea** who helped me a lot

I would like to thank also Professor **Dr. M. EL-Begermy** Professor of Otorhinolaryngology, Faculty of Medicine, Ain Shams University, who keeps teaching me and shall –god willing- keep as such

To the eminent Professors of the Otorhinolaryngology Department at the Faculty of Medicine, Ain Shams University, I wish to express my appreciation for permitting me to attend at the Operating Theater to learn from their elegant surgical skills, as well as permitting the extirpated specimens to be used for the fulfillment of the quest of this research.

Technical support considering RT-PCR analysis was continuously given by **Dr. Aisha M. Abd EL-Kareem** Ph.D. Biochemistry faculty of science -Ain Shams University, many thanks to her.

None of this work would have been possible without the co-operation of the patients, the *raison d'être* of all the Medical Profession. I thank them all.

## **CONTENTS**

Introduction.		١
Aim of the W	ork	٧
Review of the	e Literature	٨
0	Cancer as a genetic disease	٨
0	Molecular biology of head and neck cancer	۲٦
0	Human Papillomavirus	٣٦
Patients & Mo	ethods	٤٨
Results		0 8
Discussion		٧٨
Summary an	d Conclusion	٨٨
References		٩.
Appendix		
Arabic Sumn	1ary	

## **LIST OF TABLES**

<u>List of Tables</u> P.	
<b>Table (^):</b>	Mechanisms of TPor inactivation in human cancer
Table (7):	Classification of the patients٤٩
Table (*):	Sub-sites of the tumors in the study group (G1) $\xi 9$
Table (٤):	Types of lesions in the control group $(G^{\gamma})$
Table (°)	HPV 17 DNA and RNA primers
Table (٦):	T-staging for patients in study group
Table (Y):	N-staging for patients in study group
Table (^):	TNM-staging of patients in study group
Table (4):	Histopathlogical grading distribution in patients with laryngeal carcinoma
Table (' '):	Modality of treatment of patients with laryngeal carcinoma
<b>Table</b> ( ' '):	Vital status of patients in study $group(G)$ $^{\circ A}$
Table ( \ \ \ \ ):	Comparison of age distribution between study and control group
Table ( ۱۳):	Comparison of HPV T DNA and HPV T ET RNA detection in tumor core

Table (\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	Sensitivity, specificity, PPV & NPV of HPV DNA assay to HPV RNA assay	٦٣.
Table (۱°):	Comparison of HPV 17 E7 RNA detection in tumor core and benign lesions	.7٤
Table ( ۱٦):	Comparison of HPV RNA in tumor core and resection margins	٥٦.
Table ( \ \ \ ):	Comparison of HPV RNA detection in tumors of the different sub-sites	.77
Table (۱۸):	Comparison of HPV RNA detection in tumors of the different TNM stages	۲۲.
Table (۱۹):	Comparison of HPV RNA detection in safety margins of the different TNM stages	۸۲.
Table (۲۰):	Comparison of HPV RNA detection in tumors in different histopathological grading	.٦٩
Table (۲۱):	Comparison of HPV RNA detection in safety margins in different histopathological grading	.٦٩
Table (۲۲):	Comparison of HPV RNA in tumor core regarding to recurrence	٠٧.
` ,	Comparison of HPV RNA in resection margins regarding to recurrence	. ۷۲
Table (۲٤):	Comparison of HPV RNA in tumor core regarding to mortality	.٧٤
Table (۲°):	Comparison of HPV RNA in safety margins regarding to mortality	۲۷.

## **LIST OF FIGURES**

	Title	Page
١	Cell cycle	9
۲	Cellular localization of oncogene proteins	10
٣	A sketch of the TP° network	77
٤	Genome of HPV E'-E' early genes, L'-L' late genes	٣٨
0	Analytical gel electrophoresis shows the 'AV' bp product of the PCR amplification reaction of the HPV'7 DNA in study group	09
٦	Analytical gel electrophoresis showing the ' ' bp product of the RT-PCR amplification reaction of the HPV ' E' mRNA in study group	·
٧	Analytical gel electrophoresis showing the ' ' bp product of the RT-PCR amplification reaction of the HPV ' E mRNA in control group	٦١
٨	Kaplan-Meier log-log survival curve by weeks shows no significant statistically difference between HPV '7 E7 Protein positive and negative groups in tumor core regarding the recurrence	٧١
4	Kaplan-Meier log-log survival curve by weeks shows no significant statistically difference between HPV '7 E7 Protein positive and negative groups in safety margins regarding the recurrence	٧٣
١.	Kaplan-Meier log-log survival curve by weeks shows no significant statistically difference between HPV 17 E7 Protein positive and negative groups in	٧٥

	tumor core regarding the mortality	
11	Kaplan-Meier log-log survival curve by weeks shows no significant statistically difference between HPV '7 E7 Protein positive and negative groups in safety margins regarding the mortality	<b>YY</b>

## **LIST OF ABBREVIATIONS**

AJCC	American Joint Committee on cancer
CAK	Cyclin activated kinase
CDK	Cyclin dependent Kinase
CIN	cervical intra-epithelial neoplasia
CKI	Cyclin dependent kinases associated inhibitors
E', E' E°, E', E'	Early proteins
EGF	Epithelial growth factor
FGF <sup>1</sup>	fibroblast growth factor
FISH	fluorescence in situ hybridization
G٠	Non-dividing cells stage
G١	stage stands for "GAP \"
G٢	stage stands for "GAP 7"
Hdm-Y	Human double minute
HNSC	Head and Neck Squamous cell carcinoma
HPV	Human papillomavirus
IARC	International Agency for Research on Cancer
L1,L7	late" proteins
LOH	loss of heterozygosity
M	stage stands for "mitosis"
Mdm-۲	Murine double minute
ORF	Open reading frames

ТРот	Protein product of tumor suppressor gene with molecular weight or Kd
PCR	Polymerase chain reaction
R point	Restriction point
RB	Retinoblastoma
RT-PCR	reverse transcription-PCR
S	stage stands for "Synthesis"
SEER	Surveillance, Epidemiology & End Results
SV:	Simian virus ٤٠
TGF alpha	transforming growth factor alpha
TSGs	tumor suppressor genes
WNT \	wingless/int-\

#### INTRODUCTION

Tobacco smoking and extensive alcohol drinking are known risk factors in the aetiology of head and neck squamous cell carcinoma (HNSC). Other known risk factors include environmental exposure to wool dust, wood dust and mineral fibers. However increasing number of (HNSC) in the absence of exposure to above risk factors suggests the presence of additional risk factors (*Scully*, \*\(\mathcal{F}\)\cdot\(\mathcal{F}\)\).

In 1977, Shope and Hurst observed that infection with the cottontail rabbit Papillomavirus led to subsequent development of keratinous lesions, some of which epithelial progressed to invasive neoplasms. This observation led to the discovery of the first DNA virus that caused tumours in mammals. Substantial evidence now supports the role of HPV in the development of (premalignant) lesions of the vulva, penis, anus and uterine cervix. In addition, epidemiological and molecular data suggest that HPV may also promote head and neck carcinogenesis (Zur Hausen, Y·· Y).

HPVs are small epitheliotrophic DNA viruses  $\approx \xi \circ$  nm in diameter with circular double strand DNA genome of  $\approx \Lambda$  Kb. Based on their capsid structure they belong to the family of papovaviridae. Nearly  $\Upsilon$  HPV

genotypes have been identified, subdivided into benign and malignant or oncogenic subtypes, the benign subtypes, including HPV-7 and -17, are associated with mucosal warts and papillomatosis, such as occurs in the larynx, while others particularly HPV-17 and -14 are strongly associated with malignancy (*Harriet et al.*, \*\*••\*\*\*).

Human papilloma virus share a common life cycle that can be divided into several steps. In the first step virus particles infect squamous epithelial cells. Initially, a latent infection is established, in which the tissue is clinically and histologically normal but the viral DNA is present in low copy numbers. The latent infection can persist in the absence of disease for the life of the host. The mechanism of activation of latent HPV infection is not known but two possibilities have been suggested; first minor irritation, wounding or exposure to ultraviolet (UV) light. Second, transient or local immunosuppression could permit viral activation (*Benton et al.*, 1997).

Activation of latency is accompanied by a marked increase in expression of HPV RNA in suprabasal cells, amplified synthesis of viral DNA in upper layers, and production of new virus particles in a subset of cells in the uppermost layers which is accompanied by increase level of viral proteins. Human papillomavirus DNA is organized into three regions. The early region contains the open

reading frames (ORFs) for all viral particles. These regions are denoted by an E preceding the number defining the individual ORF. Et and Et of the high risk HPV (HPV-1) and -1A) can both immortalize and transform cell. The "late" regions L<sub>1</sub> and L<sub>2</sub>, code for the viral capsid proteins. These genes are expressed only late in viral life cycle after the viral DNA has been amplified to high levels. The third region of the genome, the upstream regulatory region (URR), does not code for viral proteins. It regulates viral gene expression and regulation (*McMurray et al.*, \*\*••\*\*).

The possible mechanisms associated with HPV-mediated carcinogenesis include both TP° mutation dependent and mutation independent pathway. The former mainly acts in upper aerodigestive tract tumors and the latter acts in cervical tumors. Mutation of the TP° tumor suppressor gene in upper aerodigestive tract is associated with alternations in the apoptotic regulatory bcl- and bak genes, leading to down-regulation of programmed cell death and increased cell proliferation. HPV infection is also associated with increased tissue angiogenesis and activation of telomerase (*Pillai and Nair*, \*···).

Human papillomavirus can be detected by various methods. Southern blot is very specific and informative. However it has several limitations it is labor intensive, it can't be carried out on formalin-fixed and paraffin

However its very sensitivity is the major limitation of PCR. Great care must be taken in handling specimens and preparing the reactions, to eliminate contamination that gives false-positive results. Although PCR can detect a small number of infected cells, it is not clear that this is biologically meaningful. Thus, some level of positive signal must be established to suggest a relationship between HPV positivity by PCR and disease. It is now recommended that semiquantitative RT-PCR be used to detect E<sup>¬</sup> or E<sup>¬</sup> transcripts, to assure that the virus is actively expressed in tumors (*Matzow et al.*, 199A).