## SOME STUDIES ON THE FACTORS OF PATHOGENIC DETERMINANTS OF STAPHYLOCOCCUS AUREUS ISOLATED FROM BOVINE MASTITIS

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By

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### بعض الدراسات عن عوامل الضراوة لميكروب المكور العنقودى الذهبى المعزول من إلتهاب الضرع في الماشية

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### بشمالله التخمر التحيم

### و ما أوتيته من العلم إلا قليسلاً

صدق الله العظيم سورة الإسراء الآية رقم ٨٥

# Dedicated To My Parents My Sisters & My Brother

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#### LIST OF ABBREVIATIONS

Y-DE Y-D electrophoresis

APHA American Public Health Association

bp Base pair

CLFa Clumping factor

CMT California mastitis test

CNS Coagulase negative staphylococci

Coa Coagulase gene

CPS Coagulase positive staphylococci

CRF Coagulase reacting factor DDW Double-distilled water

dNTPS Deoxynucleotide triphosphate solution EDTA Ethylene diamine tetra-acetic acid

ET Exfoliative toxin
Fnb Fibronectin binding A
FSF Fibrin stabilizing factor
hla Alpha-haemolysin
hlb Beta-haemolysin

IMI Intramammary infection

Luk Leukocidin gene

MLST Multilocus sequence typing

mPCR Multiplex polymerase chain reaction

nuc Thermonuclease gene O-F Oxidation-Fermentation

PAGE polyacrylamide gel electrophoresis.

PCR Polymerase chain reaction
PFGE Pulse field gel electrophoresis

RFLP Restriction fragment length polymorphism RPLA Reverse passive latex agglutination assay

rRNA Ribosomal ribonucleic acid

S. Staphylococcus SCC Somatic cell count

SDS Sodium dodecyle sulphate
SE Staphylococcal enterotoxin
SEA Staphylococcal enterotoxin A

Sp. Species

Spa Staphylococcal protein A

Str. Streptococcus

TCT Tube coagulase test

TE Tris-EDTA

TM Melting temperature
TPE Tris-phosphate EDTA

TSST-I Toxic shock syndrome toxin-

Tth Thermus thermophilus

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### **INTRODUCTION**

Worldwide bovine mastitis is the most common infectious disease affecting milk producing cows causing economic losses higher than any other disease of dairy cattle (Gillespie and Oliver,  $^{r} \cdot \cdot ^{o}$ ). Also, it possesses food safety and anti-microbial resistance threats (Kim et al.,  $^{r} \cdot \cdot ^{o}$ ), as it is the primary contamination source of milk and milk products especially in case of defective pasteurization (Joffe and Baranovics,  $^{r} \cdot \cdot ^{o}$ ).

The major causes of bovine mastitis are Staphylococcus (S.) aureus, Str. agalactiae, Str. dysgalactiae, Str. ubris (Phuektes et al.,  $\uparrow \cdot \cdot \cdot 1$ ). S. aureus is recognized worldwide as a major pathogen causing subclinical intramammary infections in dairy cows (Salasia et al.,  $\uparrow \cdot \cdot \cdot \cdot \cdot 1$ ).

Identification of bacterial pathogens in milk from cows with mastitis is the definitive diagnosis of mastitis infections. It also provides information important for prevention and control of the disease. In most clinical laboratories identification methods are based on microbiological culture of milk and biochemical identification of bacterial isolates recovered. However, these microbiological cultures are limited by the dynamic nature of infections. Subclinically infected cows are intermittent shedders of organisms through low and high shedding patterns during lactation leading to negative cultures (*Phuektes et al.*, \*\*\*\*\*).

In bovine mastitis, coagulase positive staphylococci (CPS) (S. aureus and some strains of S. hyicus) are common pathogenic than coagulase negative staphylococci (CNS) isolates (Ali-Vehmas and Sandholm, 1992). The TCT has the potential to detect not only S. aureus but also other CPS in milk.

The ability of *S. aureus* to form clumps in the presence of plasma has been known since the turn of the century (*Much*, 19.4). Staphylococcal components which interact with fibrinogen and can be purified from *S. aureus* culture supernatant fluids have been described. These include a staphylocoagulase and a cell wall protein (clumping factor, CLFa) which binds with fibrinogen (*Boden and Flock*, 1997). Staphylocoagulase, produced by *S. aureus*, form an active molecular complex with prothrombin which converts fibrinogen to a fibrin clot (*Hendrix et al.*, 1947). Studies of gene sequences have demonstrated that coagulase and CLFa factor are genetically distinct (*McDevitt et al.*, 1997). Detection of staphylocoagulase and CLFa are used for identification of *S. aureus* in diagnostic laboratories (*Kloos and Schleifer*, 1947). Molecular epidemiological analysis of bovine *S. aureus* population suggested that a small numbers of clonal types were responsible for most infections, and that strains had a broad geographic distribution (*Kalorey et al.*, 1998). A better knowledge on the distribution of *S. aureus* in dairy animals might help formulate strategies to reduce the spread of infection (*Salasia et al.*, 1998).

The analysis of relationship between presence/absence of the different genes and the udder inflammatory response measured by milk somatic cell count (SCC) showed that at least one cluster, which made by classification of isolates in cluster by virulence genes, induced a higher inflammatory response. Moreover, the analysis of the association between virulence genes and the presence of subclinical mastitis, support that it could be related to strain characteristics, and the expression of specific combination of virulence factor specially the spa (staphylococcal protein A), COA (coagulase gene) and SEJ (one of the SEs) (**Zecconi et al.**, \*\*.\*\*\*7). No remarkable difference was recognized in the identification ratio of the isolates which belonged to the two major lineages between mastitis of subclinical origin and clinical origin (**Hata et al.**, \*\*.\*\*7).

This work aimed to study some factors of the pathogenic determinants of *S. aureus* isolated from clinical bovine mastitis. This will be accomplished through:

- 1. Isolation and classification of staphylococci recovered from samples collected from clinically mastitis cows examined.
- 7. Determining the incidence of *S. aureus* in milk samples collected from cases of bovine clinical mastitis.
- γ. Studying certain pathogenic determinants of *S. aureus* recovered from clinical mastitis milk. Moreover, identification of *S. aureus* enterotoxin (SEA) by using PCR.