EVALUATION OF SURVIVIN AND HYALURONIDASE AS URINE MARKERS IN PATIENTS WITH BLADDER CANCER

Thesis

Submitted for Fulfillment of Master degree in Biochemistry

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Acknowledgement

My gratitude is for Allah, His merciful, for what He has given me and for all the blessings, He supplied me with, thanks **God**.

It is a great pleasure to acknowledge the help of many indviduals without their help this work could not have been done.

First, I really find myself lucky to be supervised by **Prof. Dr.Maged Barakat**, professor of Biochemistry, Faculty of Pharmacy, Cairo University, for his continous encouragement and sincere advice which has been the main factors to complete this work.

No words could express my sincere appreciation and deepest gratitude to **Prof. Dr. Sanaa Eissa**, professor of Biochemistry, Faculty of Medicine, Ain Shams University. This work was not merely a project for her, but it was a special aim to which she gave her ultimate effort and attention. Words can not find their way to express my thanks to her expertise supervision.

Special thanks are owed to **Prof. Dr.Ashraf Zagloul** Prof of Surgical Oncology, National cancer institute, Cairo University.

Special thanks to **Dr.Soheir Badr**, Ass.Prof.of Biochemistry, Faculty of Medicine, Ain Shams University, for her help in revising this work, May God put this work in her good deeds balance.

I would remiss if I failed to acknowledge my parents, my sister and my friends, for their tolerance and their emotionally support that pushed me to finish this work, many thanks.

Marwa Mohanad El-Marzoki

List of contents

| INTROE | DUCTION | 1 | |
|--------------------------|--|-----|--|
| AIM OF | THE WORK | 5 | |
| REVIEW OF LITERATURE6 | | | |
| _ | Bladder Cancer | 6 | |
| _ | Epidemiology | 6 | |
| _ | Etiology and risk factors | 8 | |
| _ | Bladder Cancer Staging and Grading | 14 | |
| _ | Types of bladder cancer | 25 | |
| _ | Diagnosis of bladder cancer | 29 | |
| _ | Apoptosis | 51 | |
| ſ | Mechanism of action of IAPs: | 74 | |
| _ | Survivin | 77 | |
| _ | Hyaluronic acid and Hyaluronidase | 106 | |
| MATERIALS AND METHODS124 | | | |
| _ | Qualitative detection of bilharzial antibodies | | |
| | in sera | 130 | |
| _ | Estimation of protein concentration | 133 | |
| _ | Quantitative measurement of survivin in urine | 135 | |

| - | Detection of Hyaluronidase RNA in Urine pellets by RT-PCR | | |
|---------------------------|---|------|--|
| | A-Total RNA extraction from urine pellet sample | s141 | |
| | B- Reverse transcriptase polymerase chain (RT-PCR) | | |
| | C- Detection of PCR product | 153 | |
| _ | Statistical analysis | 157 | |
| Results | | 160 | |
| Discussion180 | | | |
| Summary and conclusion197 | | | |
| References203 | | | |

List of Figures

| Figure (1): Tumor staging in Bladder Cancer according to TNM system18 |
|---|
| Figure (2): Human Carcinogenesis is a multilayer process |
| Figure (3) : Hallmarks of the apoptotic and necrotic cell death process |
| Figure (4): Schematic representation of two major apoptotic pathways in humans the "extrinsic" and "intrinsic" |
| Figure (5): Schematic representation of some major apoptotic signaling pathways60 |
| Figure (6): Regulation of apoptosis by BcL2 Family64 |
| Figure (7): The IAPs family of proteins69 |
| Figure (8): An induced proximity model for XIAP's ubiquitin ligase activity |
| Figure (9): Mechanism of caspase Inhibition by IAPs and its prevention by the IAP antagonist DIABLO/Smac74 |
| Figure (10): Human Survivin Isoforms81 |
| Figure (11): Regulation of survivin86 |
| Figure (12): Potential mechanisms by which survivin inhibits apoptosis |
| Figure (13): A schematic representation of the different strategies to target survivin in cancer therapy105 |

| Figure (14): Scheme for Catabolic pathway of HA114 |
|--|
| Figure (15): The mean rank of survivin in the different investigated groups |
| Figure (16): The ROC curve analysis of survivin169 |
| Figure (17): The positivity rate of survivin in the different investigated groups according to the best chosen cutoff value 2537.25 pg/mg protein |
| Figure (18): RT-PCR product analysis of HAase and β -actin by agarose gel electrophoresis and ethidium bromide staining from urine samples of bladder cancer patients. 173 |
| Figure (19): RT-PCRproduct analysis of HAase and β -actin by agarose gel electrophoresis and ethidium bromide staining from urine samples of different investigated groups |

List of Tables

| Table (1): Grading of Bladder Cancer 24 |
|---|
| Table (2): Clinicopathological factors in malignant group 161 |
| Table (3): Positivity rate of urine cytology in relation to clinicopathological factors in bladder cancer groups . 163 |
| Table (4) : The concentration of survivin (pg/mg protein) in the different investigated groups |
| Table (5): Relation of survivin level (pg/mg protein) with different clinicopathological factors of bladder cancer |
| Table (6): The positivity of survivin in the different investigated groups according to the best chosen cutoff value 2537.25 pg/mg protein |
| Table (7): The positivity rate of survivin in relation to different clinicopathological factors in th bladder cancer according to cutoff value ≥ 2537.25 pg/mg protein |
| Table (8): Positivity rate of urine HAase RNA in different groups study 175 |
| Table (9): Positivity rate of urine HAase in relation to clinicopathological factors in bladder cancer group 177 |
| Table (10): Combined sensitivity and specificity with accuracy for the investigated parameters in bladder cancer |

List of Abbreviations

AC Adenocarcinoma

AJCC American Joint Committee of Cancer

Ala Alanine

ALA Amino levulinic acid

Apaf- 1 Apoptotic Protease Activating Factor-1

b-FGF Basic fibroblast growth factor

BIR Baculovirus IAP domain

BLCA-4 Bladder cancer-4

BSA Bovine serum albumin
BTA Bard tumor antigen

CAMs Cell adhesion molecules

CARD Caspase activation recuritment domain

CD34+ Bone marrow cells
CD44 Adhesion molecules

CDK Cyclin dependant kinase

cDNA Complementry deoxy ribonucleic acid

CIN Cervical intraepithelial neoplasia

CIS Carcinoma in situ

CK Cytokeratin

CT Computed tomography

CYFRA 21-1 Cytokeratin-19

Cys Cysteine

CYP1A2 Cytochrome P 1A2 dehydrate

DNA FCMDNA flow cytometry
DNA Deoxy ribonucleic acid

EC Endothelial cell

ECM Extracellular matrix

ELISA Enzyme linked immunosorbent assay **FDP** Fibronogen degradation producr

FGF Fibroblast growth factor

FISH Fluorescence in-situ hybridization

GPI Glycosyl phosphatidyl inositol

HAHyaluronic acidHAaseHyaluronidase

hALA Ester derivative of ALA

HBXIP Hepatitis B interacting protein

HSP90 Heat shock protein-90

hTERT Human telomerase reverse transcriptase

HyalHyaluronidase enzymeHYALHyaluronidase geneIAPInhibitor of apoptosis

ICE Interleukin -1ß-Converting Enzyme
IHA Indirect haemagglutination test

ISUP International Society of Urological Pathology

IVP Intravenous pyelogramMMP Matrix metalloproteinase

MRI Magnetic resonanace imaging

mt-DNA
 NAT1
 N-acetyl transferase 1
 NAT2
 NCI
 National Cancer Institute

NF Nuclear factor

NMP22 Nuclear matrix protein

NNA N-nitrosamines

NPV Negative predictive value

p53 Protein 53 (oncogene product)

PA Plasminogen activator

PAH Polycylic aromatic hydrocarbons
PARs Plasminogen activator receptors

PCR Polymerase chain reaction

Pg Picogram

PI3K Phosphoinositol-3 kinase
PPV Positive predictive value

PIN Prostatic intraepithelial neoplasia

pro-IL-18 Proinflammatory cytokinespro-IL-1ß Proinflammatory cytokines

PT Permeability transition

RASSFIA Tumor suppressor gene

Rb Retinoblastoma protein

RBCs Red blood cells
RNA Ribonucleic acid

ROS Reactive oxygen species

RT-PCR Reverse transcriptase polymerase chain reaction

RZ Ribozyme

siRNA Small interfering RNA

SCC Squamous Cell Carcinoma

SD Standard of deviation

STAT-3 Signal transducer and activator of transcription3

TBS Tris buffer saline

TCC Transitional Cell Carcinoma

Thr Threonine

TNF Tumor necrotic factor

TRAP Telomeric repeat action protocol

UBC Urinary bladder cancer test

VDAC Voltage dependent anion channel VEGF Vascular endothelial growth factor

WHO World health organization

Abstract

Purpose: To evaluate a convenient and non invasive procedure to diagnose bladder cancer by examining the usefulness of urinary survivin and HAase RNA in diagnosis of bladder cancer and to evaluate their sensitivity and specificity in comparison to urine cytology. Also, the study correlates these factors with different clinicopathological factors.

Materials and Methods: The study was done on 100 urine cases; 60 cases were collected from malignant bladder cancer patients; 20 cases were collected from benign bladder patients and 20 cases were collected from apparently normal individuals. Two factors survivin "antiapoptotic protein" and hyaluronidase were detected in urine specimens of the different studied groups. Survivin was quantitatively measured by using enzyme linked immunosorbent assay (ELISA). HAase was detected in urine pellets by RT-PCR.

Results: Survivin and HAase were significantly increased in malignant group than benign or normal control group. There was no significant relationship between survivin and HAase with different clinicopathological factors. The sensitivity of HAase alone was 86.67% while that survivin of and urine cytology was 78.33% and 38.33%, respectively. The sensitivity of urine cytology was increased on combination with either survivin or HAase. Also, combination of both markers increased overall sensitivity.

Conclusion: Inspite of slightly lower sensitivity of survivin 78.33% than HAase 86.67%, survivin detection has the advantage of being a quantitative test measured by ELISA which is of lower cost and easily performed than RT-PCR. Many precautions should also be considered on collection, transport and storage of samples when detecting HAase RNA on these samples. Combined use of cytology with survivin and HAase was the best recommended combination for bladder cancer detection.

KEY WORDS: Bladder cancer; apoptosis; survivin; hyaluronidase; cytology

INTRODUCTION

Bladder cancer represents a significant public health problem leading to more than 130,000 worldwide deaths annually. Disease prevalence is remarkable where more than 500,000 patients carrying the disease in United States alone (**Borden** *et al.*, 2003).

Carcinoma of the bladder is the most prevalent cancer in Egypt and in most African countries. At National Cancer Institute (NCI), Cairo, it constitutes 30.3% of all cancers (El-Mawla *et al.*, 2001).

Early detection of high grade carcinoma may help to improve the prognosis of these cases. Cystoscopy along with cytology is the main tool for diagnosis of bladder cancer. Cytology is specific for diagnosis of bladder carcinoma but less sensitive particularly in low-grade disease. Cystoscopy on the other hand is invasive but relatively costly technique and may be inconclusive at particularly in times case of cystitis. These characteristics have prompted the search for more reliable noninvasive markers of bladder cancers (Ianari et al., 1997).

A noninvasive method for the detection of urothelial carcinomas of urinary bladder would help improve assessment and follow up of patients with bladder carcinoma, as well as improve screening of high-risk groups, such as patients with schistomiasis and smokers, for the development of these malignancies (**Eissa** *et al.*, **2004**).

Regulation of cell proliferation by programmed cell death (apoptosis) contributes to tissue and organ homeostasis during development and differentiation. This process involves an evolutionary conserved multistep cascade and is controlled by proteins that promote or counteract apoptotic cell death (**Adida** *et al.*, **1998b**).

Increasing resistance to programmed cell death by an imbalance between proapoptotic protein and antiapoptotic protein plays a critical role during tumorigenesis and tumor progression facilitating the accumulation of transforming mutation and promoting evasion of tumor cells from immunosurveillence. A number of gene products with anti-apoptotic potential are known to modulate tumor cell viability and