

Introduction

Diabetes has been considered an independent serious risk factor for development of myocardial dysfunction (*Fein et al., 1990 and Shiomi et al., 2003*). Diabetic cardiovascular problems represent the leading cause of death in diabetic patients (*Aronson et al., 1999 and Borges et al., 2006*).

Diabetic metabolic dysregulation is characterized by reduction of glucose utilization and reliance most exclusively on fatty acids to generate energy, disturbed oxygen metabolism with increased production of oxygen radicals such as reactive oxygen species (ROS) (*Halliwell, 1991 and Bocci, 2005*), lipid oxidation products (LOP) (*Leinonen et al., 1999 and Schleicher et al., 1999*) and insufficient antioxidants (*Szaleczky et al., 1999 and Bocci, 2005*) altering oxidant/antioxidant balance and producing oxidative stress disorder (*Ye et al., 2003*).

Diabetic chronic oxidative stress could possibly be among other factors the linking element between many diabetic complications (*Du et al., 2000 and Nishikawa et al., 2000*) mediates structural and functional cardiovascular derangement, impairment of

various intracellular signaling pathways (*Feuvray, 2004*) and dysfunction in cardiac myocytes (*Suematsu et al., 2002*) as well as neutrophils adhesion to endothelial cells (*Kokura et al., 1999*).

Antioxidants might represent a new addition to the measures used in diabetic patients for preventing oxidative stress complications (*Haidara et al., 2006*). However, use of conventional exogenous antioxidants cannot rehabilitate cells to actively increase intracellular antioxidant capacity to achieve a therapeutic result in contrast to medical ozone therapy (*Salvemini et al., 1999 and Bocci, 2005*).

In the light of rising of numbers of cardiac interventions invoking transient ischemia then reperfusion, the incidence of reperfusion injury (*Zahler et al., 2000*) described at least in part as a misguided inflammatory response resulting from transient oxidative stress production of excess oxygen by-products or ROS (*Belke et al., 2000*) is gaining ever-increasing relevance for development of protective strategies.

However, beneficial effects of reperfusion therapies have been limited by ischemic damage that

occurs before reperfusion and during reperfusion injury (*Bucciarelli et al.*, २००१).

A powerful tool to render an organ resistant to subsequent severe ischemia and reperfusion is confronting it to brief episodes of ischemia or oxidative stress ascribed as “ischemic preconditioning” (*Ma et al.*, १९९१; *Schreck et al.*, १९९१ and *Zahler et al.*, २०००). Brief levels of oxidative stress produces transient levels of ROS which can reduce subsequent injury to reperfused vessels (*Akimitsu et al.*, १९९१) and cardiomyocytes (*Zhou et al.*, १९९१) after submitting them to ischemia and can reduce leukocytic adhesion to vascular wall (*Kubes et al.*, १९९१).

Also, transient levels of ROS that can confer protection or adaptation to oxidative stress and control diabetic complications (*Al Dalain et al.*, २००१) can be adopted by intermittent controlled ozone therapy ascribed as “oxidative preconditioning” (*Leon et al.*, १९९१ and *Candelario et al.*, २००१) analogous to ischemic preconditioning (*Murry et al.*, १९९०) which act by inducing a transient calculated oxidative stress by ROS and LOP that activate many cellular responses (*Viebahn*, १९९९ and *Bocci*, २०००).

Aim of the work

The present study is designed to assess the possible cardiac protective effect of oxidative conditioning by medical ozone and ischemic preconditioning by femoral artery ligation, in streptozotocin-diabetes. Also, the consequence of ischemic insult is evaluated after short- and long-term preconditioning mechanisms.

Cardiovascular Disease In Diabetes

Diabetes is considered as a risk factor for cardiovascular disease (*Laakso, 1999*) as well as for myocardial dysfunction (*Fein et al., 1990*). Diabetic cardiovascular problems represent the leading cause of death in diabetic patients (*Aronson et al., 1997 and Borges et al., 2006*).

Diabetes is characterized by increased production of oxygen radicals such as reactive oxygen species (ROS) (*Halliwell, 1991 and Bocci, 2000*) and lipid oxidation products (LOP) (*Leinonen et al., 1997 and Schleicher et al., 1997*).

Diabetes is characterized by insufficient antioxidants (*Szaleczky et al., 1999 and Bocci, 2000*) that alters oxidant/antioxidant balance (*Ye et al., 2003*) which could link between many diabetic complications (*Du et al., 2000 and Nishikawa et al., 2000*).

Vascular tissue damage in diabetes is due to glucose-mediated four mechanisms: increased advanced glycated end products (AGE) formation, polyol

pathway, activation of protein kinase and hexosamine pathway flux (*Brownlee, २००१*) and due to increase free fatty acids-mediated mechanisms (*Brownlee, २००६*).

Cardiac endothelium:

Endocardial endothelium and pulmonary endothelium constitute central vascular endothelium (*Rosenberg and Aird, १९९९*) which contains the largest source of endothelial enzymes and endothelial mediators acting through specific receptor signaling pathways (*Brutsaert, २००२*).

Vascular endothelial cells produce nitric oxide (NO) which regulates vasodilatation, anticoagulation, leukocyte adhesion, smooth muscle proliferation and antioxidative capacity of endothelial cells (*Rakhit and Marber, २००१*).

Moreover, vascular endothelium influences greatly synthetic and motor functions of the underlying smooth muscle and similar regulatory mechanisms and vessel wall cross-talk exist between pericytes and microvascular endothelium (*Chakravarthy and Gardiner, १९९९*).

Endothelial activation and dysfunction:

Cardiac endocardial endothelium (EE) and myocardial capillary endothelium (MyoCapE) are involved in modulating myocardial growth, contractile performance and rhythmicity (*Brutsaert, 1990; Brutsaert and Andries, 1992 and Andries et al., 1996*).

Evidences suggest that EE syncytium exhibits strong electrochemical coupling between EE cells (*Beny, 1999*) that function as a sensor that amplify release of endothelium-derived paracrine substances which likely can cause modulation of interstitial ionic homeostasis and disturbances in excitability and conduction of subendocardial myocardial layers including terminal purkinje fibers and dense subendocardial plexus exhibiting a positive chronotropic effect and or arrhythmia (*Hassanabad et al., 1998*).

Moreover, EE functions as an autocrine or paracrine organ-oriented modulator of cardiac contractile performance, rhythmicity and growth, mostly right ventricular EE (*Arai et al., 1999*).

Right ventricular EE cell to cardiomyocyte number ratio is high, so right ventricular pump performance will be adjusted to incoming blood-born

signals of mixed venous blood (*Brutsaert, 2003*) unlike left ventricular EE which optimizes cardiac pumping performance through coronary circulation and MyoCapE (*Brutsaert, 2003*).

Vascular endothelial cells contain nitric oxide synthase (NOS) which synthesizes NO (*Moncada et al., 1999*).

Three NOS isoforms are characterized: type I neural constitutive (ncNOS), type II inducible (iNOS) and type III endothelial constitutive (ecNOS). All three NOS isoforms are present in human myocardium (*Kelly et al., 1997*).

Low level of endogenous basal non-stressed physiological conditions produces NO almost exclusively by MyoCapE, EE and coronary vascular endothelium (*Schulz et al., 1991 and Andries et al., 1998*) which causes +ve inotropic action (*Mohan et al., 1997*).

Nitric oxide is known to be a modulator of biological phenomena from cell signal to effector and from physiology to pathology. NO is an important physiological regulator in maintenance of cell physiology from immunomodulation to calcium signaling. Low NO concentrations are associated with

cytostasis and high concentrations are associated with cytotoxicity. Balance between cytostatic and cytotoxic effects of NO may lie in tissue concentration in which NO produced, particular NO synthase isoform is activated and complex interaction with other ROS, is particularly pertinent in context of ischemia-reperfusion (I/R) (*Rakhit and Marber, 2001*). A cardioprotective role was addressed to NO mainly by myocardial preconditioning and modulating mitochondrial function (*Cassina and Radi, 1997*) and antioxidant effects (*Siow et al., 1999*).

Endogenous nitric oxide (NO) synthesis during ischemia reperfusion was found to protect isolated guinea pig hearts versus apoptotic cell death (*Czarnowaska et al., 2001*).

Diabetic endothelial dysfunction:

Endothelial damage is a major cause of associated vascular complications (*Inguchi et al., 2000*) in diabetes and ischemia reperfusion (*Okada et al., 1998 and Qi et al., 1998*). NO reacts with superoxide in vascular endothelial cells to form peroxynitrite (ONOO⁻) which causes tissue injury (*Beckman et al., 1990; Ishida et al., 1997 and Wang and Zweier, 1997*), NO can inhibit oxygen radical

pathways e.g., xanthine oxidase and leukocyte NADPH oxidase (*Lee et al.*, 2000).

Tumor necrosis factor- α (TNF- α) levels can induce iNOS (*Habib et al.*, 1996; *Torre et al.*, 1996; *Kasai et al.*, 1997; *Sato et al.*, 1997 and *Sasayama et al.*, 1999) which provokes further TNF- α expression (*Kalra et al.*, 2000) in cardiomyocytes and cardiac endothelium (*Sigusch et al.*, 2000) causing cardiac depression (*Oral et al.*, 1997; *Kubota et al.*, 1997 and *Bozkurt et al.*, 1998).

TNF- α is a key mediator of reperfusion injury released early after I/R (*Squadrito et al.*, 1993 and *Kupatt et al.*, 1999) which increases IL β and IL α secretion that chemotactically activate leukocytes (*Tritto et al.*, 1997).

Hemodynamic outcome in streptozotocin (STZ)-induced diabetic rats is implicated to be resulted from reduced subendocardial coronary perfusion which impaired subendocardial coronary reserve (*Borges et al.*, 2006) and sympathetic innervation of heart and vessels (*Fazan et al.*, 1997; *Fazan et al.*, 1999; *Schann et al.*, 1997 and *Akiyama et al.*, 1989), as well as decreased cardiac output (*Ren and Bode*, 2000).

Postperfusion increased expression of angiotensin II receptor type I (ATI-R). Angiotensin II and catecholamines (*Imai et al., 1992 and Ito et al., 1993*) could have enhanced ET-1 release which induce superoxide radicals mediating post ischemic endothelial dysfunction, P-selectin expression and neutrophil adhesion (*Ergul, 2002 and Duda et al., 2006*).

Cardiac endothelial cells ATI-R is activated in diabetic rats by oxidative stress which could contribute to superoxide-mediated NO activation (*Mehra et al., 2005*) resulting in peroxynitrite formation which is toxic to cardiac myocytes (*Ishida et al., 1996*). This represents a preferential site for neutrophil activation, increased adhesion molecules expression (*Sohn et al., 2003*) and inflammatory cytokines as TNF- α and IL-1 which themselves potentiate neutrophil activation (*Takahashi et al., 2001*).

Diabetic cardiomyopathy:

Trost et al., (2002) proved that STZ model is a valuable tool to study diabetic cardiomyopathy using mice as an animal model in which extensive focal endomyocardial necrosis is observed in 12 weeks

diabetic rat (*Akula et al.*, २००२) while diabetic hearts in acute phase are more resistant to irreversible cell damage (*Ravingerova et al.*, २००२).

There is a body of evidence showing a predominant decrease in number of β_1 -adrenergic receptor in chronic diabetes (*Maeda et al.*, १९९०; *Savarese and Berkowitz*, १९९१ and *Dincer et al.*, २००१) while during development of diabetes (*Maeda et al.*, १९९० and *Fazan et al.*, १९९१) an increase in number of β_1 -adrenergic receptors of short duration was demonstrated (*Austin and Chess-Williams*, १९९२ and *Uekita et al.*, १९९१).

Extracellular signal regulated kinase V_γ (ERKV $_\gamma$) which protects rat myocardium against heart injury was reduced by approximately १/३ in long-term diabetes unlike short-term diabetes (*Xu et al.*, २००६).

Moreover, increased superoxide radicals in diabetic chronic oxidative stress could have induced focal mitochondrial DNA oxidative damage (*Rolo and Palmeira*, २०००) characterized by dissipation of membrane potential and calcium overload (*Rakhit and Marber*, २००१) leading to calcium handling defect in cardiomyocytes (*Muscari et al.*, २००६) together with cardiac sympathetic innervation impairment

frequently observed in diabetes (*Maeda et al.*, 1990 and *Fazan et al.*, 1999) and altered beta-adrenoceptor sensitivity (*Kaneko et al.*, 1991) could have decreased cardiac contractility (*Saini et al.*, 2000) and ejection fraction (*Joffe et al.*, 1999).

Diabetic oxidative stress could have reduced activity of Ca/calmodulin-dependent sarcoplasmic reticulum Ca^{++} uptake, sarcoplasmic reticulum Ca^{++} /calmodulin-dependent protein kinase II (Ca MkII) (*Osada et al.*, 2000), the expression of sarcoplasmic reticulum calcium-ATPase, $\text{Na}^{+}/\text{Ca}^{++}$ exchanger (*Choi et al.*, 2002 and *Brevig et al.*, 2006) and responsiveness of contractile proteins to intracellular Ca^{++} concentration (*Wang and Morgan*, 1992 and *De Keulenaer and Sys*, 1990).

Furthermore, low levels of cytosolic Ca^{++} can be attributed to reduce ryanodine sensitive Ca^{++} channel expression of cardiac myocytes in rats (*Choi et al.*, 2002 and *Brevig et al.*, 2006) and decreases inward Ca^{++} current which triggers ryanodine Ca^{++} channels to release Ca^{++} (*Choi et al.*, 2002 and *Bers*, 2002) from sarcoplasmic reticulum.

ROS-induced decline in mitochondrial respiration could have increased mitochondria capacity to

produce ROS (*Petrosillo et al.*, 2003 and *Saini et al.*, 2005), thus mitochondria can be both the first target and initiator of oxidative stress (*Christophe and Nicolas*, 2006).

In addition, focal mitochondrial DNA oxidative damage-induced calcium defect in diabetes could have resulted from inhibition of eNOS which together with increased ET-1 (*Brunner et al.*, 1999 and *Hoshino et al.*, 2005) could have interacted with NO and decreased NO availability.

Mitochondrial dysfunction also could result in persistent depression of energy production (*Du et al.*, 2000 and *Nishikawa et al.*, 2000) and cardiolipin damage mainly in mitochondrial complex I and III (*Paradies et al.*, 2004).

Oxidative stress:

Oxidative stress refers to circumstances in which ROS are excessively formed or when there are inadequate endogenous defences (*Hoeldtke et al.*, 2003). ROS include free radicals as superoxide ($\text{O}_2^{\cdot -}$), hydroxyl (OH^{\cdot}), peroxy (RO_2^{\cdot}), hydroxyperoxy (HRO_2^{\cdot}), hydrogen peroxide (H_2O_2) and hydrochlorous acid (HOCL) (*Turko et al.*, 2001 and *Evans et al.*, 2002).

West, (२०००) proposed interaction between hyperglycemia and enhanced production of ROS. Enzymatic sources of diabetic oxidative stress include NOS, NADPH oxidase and xanthine oxidase (*Guzik et al.*, २०००; *Guzik et al.*, २००२ and *Aliciguzel et al.*, २००२) while mitochondrial respiratory chain is non enzymatic source (*Green et al.*, २००६). ROS may injure cells by causing lipid peroxidation which triggers cell death (*Park and Lucchesi*, १९९९) that can be prevented when oxygen radicals are inactivated by specific scavengers (*Zweier and Villamena*, २००२).

Antioxidants:

Antioxidant is any substance that when present at low concentrations prevents oxidation of oxidizable substrate (*Halliwell*, १९९१).

Examples of enzymatic antioxidants are: superoxide dismutase (SOD), catalase in lysosomes, glutathione peroxidase (GP) in mitochondria and xanthine oxidase enzyme. While non enzymatic antioxidants are vitamins A, C and E, glutathione, alpha lipoic acid, carotenoids, trace elements like copper, zinc, selenium, coenzyme Q₁₀ (CoQ₁₀) and cofactor like folic acid, albumin and vitamins B₁, B₆, B₁₂ and B₁₂ (*Evans et al.*, २००२).