

Uses of amniotic membrane in ophthalmic practice

Eassy

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LIST OF ABBREVLATIONS

5-FU	5 –flurouracil
AGV	ahmad glaucoma valve
AM	amniotic membrane
AMG	amniotic membrane graft
AMP	amniotic membrane patch
AMT	amniotic membrane transplantation
AS	absorbable suture
B FGF	basic fibroblast growth factor
DMSO	dimethyle sulfoxide
EDTA	ethylene –diamine tetra-acetic acid
EGF	epidermal growth factor
ELISA	enzyme linked immunoabsorbed assay
FDA	food and drug administration
GDD	glaucoma drainage device
HAM	human amniotic membrane
HGF	hepatocyte growth factor
HIV	human immunodeficiency virus
HLA	human leucocytic antigen
IL	interleukin

KGF	keratinocyte growth factor
MMC	mitomycin C
m-RNA	massenger –ribonucleic acid
NAS	non absorbable sutures
NGF	nerve growth factor
ОСР	ocular cicatricial pemphigoid
PAA	para acetic acid
PCR	polymerase chain reaction
PTK	photo refractive keratectomy
RA	receptor antagonist
SJS	steven Johnson syndrome
TEN	toxic epidermal necrosis
TGF	transforming growth factor
TNF	tumour necrosis factor

Anatomy and Histology of the Amniotic Membrane

Amniotic membranes (AM) develop from extra-embryonic tissue and consist of a fetal component (the chorionic plate) and a maternal component (the deciduas). These two parts are held together by the chorionic villi and connect the cytotrophoblastic shell of the chorionic sac to the decidua basalis. (*Parry S and Strauss JF.1998*)

The fetal component, which includes the amniotic and chorionic fetal membranes, separates the fetus from the endometrium. The amniochorionic membrane forms the outer limits of the sac that encloses the fetus, while the innermost layer of the sac is the AM. The AM consists of an epithelial monolayer, a thick basement membrane, and an avascular stroma. (*Parry S and Strauss JF.1998*)

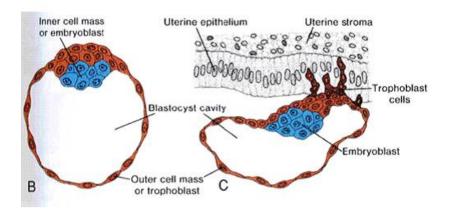


Fig. (1.1): Embryonic membranes and placenta are derived from the outer cell mass and the fetus is derived from the inner cell mass of the blastocyst. (Rafii et al.2007)

The AM, which is normally 0.02 mm to 0.05 mm in thickness, lines the amniotic cavity and has its inner apical surface bathed by the amniotic fluid, whereas the outer basal surface is in direct proximity to the chorion. Unlike the chorion, amnion is devoid of any vasculature. (Bourne GL. 1960)

The AM consists of five layers from within outward:

- **a.** A single layer of highly metabolically active, columnar to cuboidal epithelium.
- **b.** A thin basement membrane.
- **c.** A compact layer made of reticular fibers virtually devoid of cells.
- **d.** A loose network of reticulum containing fibroblasts, called the fibroblast layer.
- **e.** A spongy layer of wavy bundles of reticulum bathed in mucin, which forms the interface with the chorion. (*Bourne*, 1960)

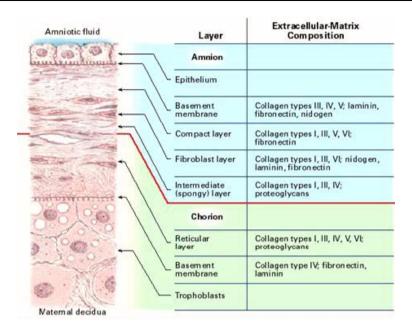


Fig. (1.2): Schematic presentation of the structure of the fetal membrane at term. The extracellular matrix components of each layer are shown. (Parry S and Strauss JF. 1998)

It was documented that matrix amniotic basal lamina contains large quantities of proteoglycans rich in heparin sulphate, where amnion contains a large amount of collagen, hyaluronan and predominantly smaller proteoglycans such as biglycan and decorin, with decorin being more prominent of the two, and is located in close connection with the collagen fibrils. (*Meinert M et al. 2001*)

Collagen types I, III, IV, V and VII, laminin and fibronectin, have been identified in amniotic basement membrane and stromal amnion. Similarities between the lamini-1, laminin-5, fibronectin and type VII collagen components of the basement

membranes of conjunctiva, cornea and AM have been demonstrated. (Fukuda K et al. 1999)

Furthermore, the α -subchain components of collagen IV have been shown to be similar between AM and conjunctiva but different between AM and cornea. (*Fukuda K et al.1999*)

Pathophysiology of AM

Some components of the membrane that are relevant in the context of its mechanism of action, and that help understand its limitations and indication.

Enzymes: Important amongst these are enzymes involved in prostaglandin synthesis such as phospholipases, prostaglandin synthase and cyclo-oxygenase as the presence of prostaglandins in the membrane would promote inflammation, whereas, the presence of prostaglandin inactivating enzyme would suppress inflammation (*Smieja et al.*, 1993)

Secretory leukocyte protease inhibitor: a potent inhibitor of human the AM. Its concentrations can be up regulated by exposing amniotic cells to interleukins(IL)-1a, IL-1b, and tumour necrosis factor (TNF α), whereas, the presence of secretory leukocyte protease inhibitor would suppress inflammation. (*Zhang et al., 2001*).

Cytokines: IL-6 and -8 are the predominant cytokines associated with amnion cells. Expression of these cytokines was increased in the presence of IL-1b, TNF α and bacterial lipopolysaccharide. IL-10 and IL-1RA (receptor antagonist), both anti-inflammatory cytokines, have been also shown in amnion epithelial and mesenchymal cells. (*Keelan et al.*, 1997)

The presence of anti-inflammatory cytokines such IL-1Ra and IL-10 would suppress inflammation but the presence of IL-6 and IL-8would promote inflammation. In eyes that are inflamed due to injury, other proinflammatory cytokines such as IL-1a, IL-1b and TNFa could also promote both pro- and anti inflammatory cytokines and enzymes. (*Hao et al.*, 2000)

Growth Factors: Studies on human amniotic membrane have revealed the presence of EGF, TGFα, KGF, HGF, bFGF, TGF-b1, and - b2 by RT-PCR for the mRNA and by ELISA for the protein products. (*Koizumi et al.*, 2000)

TGF-b3 and growth factor receptors KGFR and HGFR were also detected by RT-PCR, with a higher level of various growth factors was found in AM with epithelium than without epithelium, indicating an epithelial origin for these growth factors, the presence of various growth factors like EGF would support epithelial growth and TGF would support wound healing. (*Koizumi et al.*, 2000)

However, TGF would itself promote scar tissue formation and be contradictory to the 'anti-adhesive or scar suppressing' action proposed for the membrane in preventing corneal and conjunctival cicatrisation (*Koizumi et al.*, 2000)