

PRODUCTION OF DEVELOPED GRAPE ROOTSTOCKS USING *IN VITRO* MUTATIONS

By

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ABSTRACT

The present study was carried out during the period from ٢٠٠٨ to ٢٠١١. It aimed to induce and propagate an *in vitro* drought resistant mutagens of two Grape rootstocks *Vitis* spp., SO٤ and Freedom *via* the following steps:

First: Sterilizing shoot tips with Mercuric chloride (HgCl₂) and Sodium hypochlorite (NaOCl) at different concentrations and periods. This part results show that using ١٠ or ١٠% of Clorox for ١٠ minutes is beneficial in maintaining an aseptic culture since they record high survival and minor contamination percentages on the opposite of the higher sterilization concentrations and periods which are growth inhibitors. HgCl₂ treatments were lethal or inhibitors.

Second: a. Irradiating *in vitro* cultured plantlets of both rootstocks with Gamma rays at various doses to induce a mutation. Results indicate that all doses affected plantlets characters and ٤٠ Gy of Gamma rays is lethal for both rootstocks.

b. Testing irradiated plantlets susceptibility to drought through subculture on PEG ٦٠٠٠ containing medium at several concentrations. Five and ١٠ Gy irradiated plantlets had high survival and growth on high PEG concentrations except for the ١٠g/ L PEG which had dehydrated the plantlets within a week. Overall, the ١٠ Gy dose seem to be a promoter for survival %, leaf area, No. of leaves and roots/ plantlet, and root length. The ١٠ Gy shared or succeeded the ١٠ Gy dose in survival and most growth parameters, especially when combined with a low PEG concentration or under dehydration conditions. Twenty Gy of Gamma stimulated roots formation. However, the ٣٠ Gy dose inhibited survival and all growth parameters when cultured on different media. PEG strongly affected plantlets, since ١٠,٠ g/ L enhanced survival and growth, on the contrary of the concentration of ٨ g/ L.

Third: RAPD- PCR technique proved a genetic structure alteration in Gamma irradiated plantlets. Polymorphism in Freedom reached about ٥٧,١ % with the primer OP-C٠٩, while the Primers OP-C١٣ and OP-D١٣ showed ٣٧,٠% polymorphism in SO٤ rootstock.

Both rootstocks biologically had the same trend under all treatments even if SO٤ had superior root length, while Freedom scored a greater number of roots. Freedom was more polymorphic with Gamma irradiation than SO٤.

Key words: Gamma irradiation, Mutation, Drought, PEG, RAPD- PCR, Grape

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INTRODUCTION

Grape (*Vitis vinifera* L.) is one of the most favored fruits that have high nutritional benefits also it has a valuable commercial importance Worldwide; it is ranking the second fruit tree crop in Egyptian exportation after Citrus. This is why Grape takes a great interest from growers, which led to progressive stages in the World production and exportation. Grape production of Egypt reached about ١.٣٦٠.٢٥٠ tons cultivated in ١٥٣,٦٨٢ Feddan according to the statistics of FAO (٢٠١٠).

The Egyptian Agricultural strategy aims to continue expansion of fruit trees cultivation in the newly reclaimed areas and maximizing fruits exports.

Economic viability of a fruit tree production is linked directly to orchard productivity and management efficiency. Optimal levels of productivity require tree survival, managed vigor, and good marketable yields over the expected life span of the orchard.

The grower's choice of a rootstock is a quite important limiting factor when fruits are grown on dry, lime soils having pathogens, or other soil problems. In the Egyptian reclaimed regions, where one or more of these conditions is present, rootstocks that better address these site and vigor problems will become a crucial need. New rootstocks introduced through commercial nurseries need extensive testing to avoid commercial failures from rootstocks that prove poorly adaptation to local climatic and soil conditions. In addition, putative resistance of new rootstocks to soil problems has sometimes failed when planted in other production regions away from their origin.

Environmental stresses result in water deficiency for the plants, thus impairing its numerous biological roles. *In vitro* screening for stress tolerance will have its significance in identifying cultivars with optimal stress tolerance and productivity.

Most of rootstocks are hybrid Grape varieties that are the product of a crossing of two or more *Vitis* spp. or an inter-specific crossings. Due to their excellent tolerance to powdery mildew, other fungal diseases, nematodes and phylloxera. Hybrid varieties exhibit a mixture of traits from their European, Asiatic, and North American parentage.

Two of the best and common Grape rootstocks in Egyptian vineyards are SO⁴ or Teleki (*V. berlandieri* x *V. riparia* Michx.) and Freedom ((*V. champinii* Planch. x (*V. solonis* hort. Berol. ex Planch. x *V. othello*)). *Vitis berlandieri* is primarily known for good tolerance against soils with a high content of lime. It is poorly adapted to grafting. Various rootstocks resistant against both phylloxera and lime, and suitable for viticulture, were produced by crossing *Vitis berlandieri* and *Vitis riparia*, *Vitis rupestris* or *Vitis vinifera*.

SO⁴ and Freedom characteristics shown in Table (1), however, their behaviour against drought conditions is not satisfactory which is why this study attempted to improve drought tolerance of these two rootstocks through genetic improvement *via* tissue culture and gamma irradiation inducing mutations techniques.

The high percentage of Gamma-ray irradiated mutants indicates that mutation breeding *via* Gamma-ray irradiation is an effective and highly successful approach for the generation of

commercial cultivars and new types for growers (Nakagawa, ٢٠٠٩).

In a mutant micropropagation program, it is of paramount importance to preserve the new genetic forms, but perhaps even more important is their genetic stability. Since the traditional methods of identifying Grape cultivars have relied on morphological characters whose expression is affected by developmental and environmental factors, RAPD this PCR- based technique has been adopted as a convenient and powerful means of detecting genetic differences among closely related forms (Gupta and Varshney ١٩٩٩ and Kim *et al.* ٢٠٠٢).

Table ١. Characteristics of SO٤ and Freedom Grape rootstocks.

Type	Vigor	Root system	Nematode resistance	Drought resistance	Alkalinity tolerance
SO٤	Moderate	Shallow	Moderate-High	Low	Fair
Freedom	Moderate-High	Moderate	High	Moderate-High	?

Source: Creasy and Creasy (٢٠٠٩).

Therefore, the present work was designed to study the following:

١. Effect of surface sterilization methods on aseptic cultures establishment of SO٤ and Freedom Grape rootstocks.

٢. Gamma irradiation doses capability in inducing drought tolerance mutation in both rootstocks and testing their mutants on PEG containing media.

٣. Induced mutations detection *via* RAPD- PCR analysis.

REVIEW OF LITERATURE

The previous work dealing with the aseptic cultures establishment, effect of Gamma irradiation and PEG in nutrient medium and examining mutagenesis using RAPD on different plants will be discussed as follows:

1. Sterilization

Sterilization of different *in vitro* cultured explants is a primary and basic step to establish an aseptic culture; so that several sterilization protocols were set up by researchers and the following are some of them:

On Artichoke (*Cynara scolymus*), Alphonse *et al.* (۲۰۰۲) revealed that the most effective sterilizing and disinfectant treatment for survival and bacterial contamination prevention was dipping shoot tips in ethanol ۷۰٪ (۳۰ sec) + sodium hypochlorite (NaOCl) ۱,۵٪ (۲۰ min) + mixture of streptomycin and gentamycin each at ۵۰ mg/ L (۳۰ min).

While, Ebrahim *et al.* (۲۰۰۲) used sodium hypochlorite ۳٪ solution (contained ۰,۱٪ Tween ۲۰) alone for ۲۰ minutes for surface sterilization of four Banana cultivars shoot tips (*Musa* spp.).

Also, Lee *et al.* (۲۰۰۲) mentioned that (۱۰ mm) nodal stem cuttings of two Sweet Potato cultivars (*Ipomoea batatas* L.) were sterilized prior to culture with sodium hypochlorite (۵٪) for ۱۰ minutes then washed with sterile distilled water.

Moreover, Ashok *et al.* (۲۰۰۳) sterilized two *Cucumis sativus* L. Cvs. flower buds with ۵٪ sodium hypochlorite solution for ۲۰ min,

then in a horizontal laminar air flow using 0.1% mercuric chloride for 5 min. Flower buds were rinsed several times in sterile distilled water.

Working on Apple (*Malus domestica*), El-Sabrouh (1993) sterilized shoot tips (5 mm) by immersing in 70% ethanol for a minute then by dipping them in 0.1% solution of mercuric chloride (HgCl₂) for 3 minutes, followed by three rinses in sterile distilled water.

Stevia rebaudiana seeds were sterilized by washing with 0.1% detergent solution for 5 min, rinsed before surface sterilization in 70% ethanol for 30 sec followed by soaking in 1% commercial bleach (sodium hypochlorite) for a minute and rinsed thoroughly with sterile distilled water; Maharik and El- Gengaihi (1993).

Moreover, El-Hammady *et al.* (1995) indicated that dipping explants in 1% of sodium hypochlorite for 10 minutes was effective in sterilizing Almond shoots (*Prunus dulcis* Mill).

In another study made by Mkuya *et al.* (1995) they indicated that Rice anthers (*Oryza sativa*) were surface disinfected by immersing in 70% ethanol, then in 1.5% antiformine for 20 min and in 3% hydrogen peroxide (H₂O₂) for 10 min, and finally they were washed 3-4 times with sterile distilled water.

Whereas, Sim (1996) surface sterilized Grape tissue (*Vitis vinifera*) by submersion in sodium hypochlorite (1%) plus surfactant drops for 10 minutes. Tissue removed under aseptic conditions and serially transferred through three rinse containers containing sterile distilled water.

Also, Congfen *et al.* (1997) pointed out that the use of 0.1% sodium hypochlorite (30 min) for Aloe (*Aloe barbadensis* Miller)

sterilization was less detrimental to the explants than 0.1% mercuric chloride (10 min).

On the other hand, Yogeshwar *et al.* (1998) treated nodal segments explants (2.5 – 3.5 cm length) of *Bambusa tulda* with a quick dip for 30 sec in 70% ethanol before being administered three doses, viz., 0.05, 0.1 and 0.2% each of aqueous sodium hypochlorite or mercuric chloride solution separately for 10 min. They showed that administration for 10 min of 0.05 and 0.1% mercuric chloride to explants collected facilitated optimum culture establishment. 0.1 – 0.2% mercuric chloride enhanced aseptic culture establishment but inhibited bud break due to toxicity to explants.

Whereas, Sujatha and Kumari (1998) maintained aseptic culture of *Artemisia vulgaris* by surface sterilization of young nodal explants (1–1.5 cm) with 0.1% (w/v) aqueous mercuric chloride for 1–3 minutes.

El-Shamy *et al.* (1999) indicated that the highest percentage of survival and free contamination explants of *Pyracantha fortuneana* Roem were observed when shootlet explants soaked for 1 minute in 70% ethanol, then immersed for 30 minutes in 4% Clorox plus drops of Tween 20 and rinsed three times with sterile distilled water.

Hamouda *et al.* (1999) had surface sterilized *Pelargonium graveolens* L'Herit explants (1 cm length; internodes segments) with 0.8% sodium hypochlorite for 30 minutes.

On the other hand, Sakr *et al.* (2000) compared two sterilizer's effects; sodium hypochlorite and hydrogen peroxide on *Echinacea purpurea* seeds germination. They reported that the highest seed

germination percentage was obtained when coated seeds were sterilized with hydrogen peroxide.

Nodal segments (1, 1–1.5 cm) of *Solanum nigrum* L. plants obtained from the field were sterilized as the following method: washed in tap water for 2–3 minutes then surface sterilized in laminar air hood with 70% ethanol for 30 s followed by 0.05% mercuric chloride for 3–5 minutes, and rinsed 5–6 times with sterile double distilled water, Satish *et al.* (2010).

Wang *et al.* (2011) employed autoclaving and mercuric chloride sterilization for two-liquid-phase (TLP) soil slurry system and reported that no microorganisms were detected in the HgCl₂-sterilized soil slurries during the whole incubation period, indicating that the sterilization efficiency and effectiveness of HgCl₂ on soil slurry was much higher than those of autoclaving at 121 °C for 40 minutes.

2. Gamma irradiation and Polyethylene Glycol

a. Gamma irradiation

Gamma is an electromagnetic ionizing type of radiation affects the molecules and ions in the cells of the living creatures; this is why it is useful in inducing mutations as the following reviews:

Pandini *et al.* (1997) had induced mutations by Gamma radiation (0, 5, 10, 20 and 40 kR doses) for plant height in two Triticale cultivars (*X triticosecale* Wittmack). All radiation doses showed increase in genetic amplitude for this trait, being suitable for increasing variability in breeding programs.

Moreover, Yoshioka *et al.* (1999) had maintained nine resistant mutants of Japanese Pear (*Pyrus communis*) to Black Spot Disease

derived by chronically irradiation of susceptible cultivars with Gamma-rays induced from acutely irradiated dormant scions. Although, irradiation had displayed unfavorable characteristics, all of the mutants showed an intermediate resistance to Black Spot Disease and conferred various levels of resistance.

Another study conducted by Lamseejan *et al.* (१०००) on Gamma rays at ०, १०, ३०, ५०, ७०, ९० and ११० Gy effect on *in vitro* culture of Chrysanthemum (*Chrysanthemum morifolium*). They noticed that, M¹V⁴ shoots irradiated at ५० Gy and over died. Lethal Dose ५० (LD₅₀) was १४ Gy. Controls and treated plants at १० Gy were able to survive and gave rise to the full grown plants. After १० days, control and treated plants were found to be different in average height and number of leaves. The treated plants had much more variation than the controls.

In this respect, Morishita *et al.* (१००१) concluded that the rate of mutation in Buckwheat (*Triticum aestivum* L.) M² generation was increased with Gamma ray dose. Gamma ray LD₅₀ estimated about ३५० Gy and १०० Gy, respectively. However ३००-४०० Gy irradiation, lower than LD₅₀, was desirable to obtain higher mutation rates and fewer dead plants in this dose range.

Also, Lee *et al.* (१००२) induced somaclonal variations in Sweet Potato (*Ipomoea batatas* L.) regenerates from Gamma irradiated embryogenic callus at different levels (०, ३०, ५०, ७० and ९० Gy). The frequency of morphological variants derived from the irradiated callus ranged from ३- ४.८ % compared to ०.१- १.१% of that derived from the non- irradiated callus. Polymorphisms of the irradiated callus were १,१