Cairo University

Faculty of Medicine





Diagnostic Significance of Quantiferon TB Gold in Tuberculous Pleural Effusion

Thesis Submitted in Partial Fulfillment of the Requirements of the Doctorate Degree in Chest Diseases

By Walaa Zein El Abedin Awd

Master Degree, Cairo University, 2008

Supervisors
Prof. Dr. Khaled Eid Sobhy

Professor of Chest Diseases

Faculty of Medicine

Cairo University

Prof. Dr. Somaya Abd El-Latif Eissa

Professor of Microbiology & Immunology

Faculty of Medicine

Cairo University

Prof. Dr. Mohamed Galal Morsy

Professor of Chest Diseases

Faculty of Medicine

Cairo University

Dr. Hamed Abd El-Hafez Abd Allah

Lecturer of Chest Diseases

Faculty of Medicine

Cairo University

Faculty of Medicine Cairo University

2014



جامعة القاهرة كلية الطب قسم الأمراض الصدرية

دلالات الكوانتيفيرون في الأنسكاب البللوري الدرني

رسالة مقدمة تمهيداً للحصول على درجة الدكتوراه فى الأمراض الصدرية

مقدمة من الطبيب

و لاء زين العابدين عوض ماجستير الأمراض الصدرية - طب القاهرة - 2008

تحت إشراف

أد/ خالد عيد صبحي

أستاذ الأمراض الصدرية

كلية الطب - جامعة القاهرة

أد/ سمية عبد اللطيف عيسي

أستاذ الميكروبيولجي

كلية الطب - جامعة القاهرة

أد/ محمد جلال مرسى

أستاذ الأمراض الصدرية

كلية الطب - جامعة القاهرة

د/ حامد عبد الحفيظ عبد الله

مدرس الأمراض الصدرية

(ويشارك في الأشراف)

كلية الطب - جامعة القاهرة

كلية الطب

جامعة القاهرة

2014م

Acknowledgement

To **ALLAH** every thing in life is resumed, in this work, he has helped me a lot, if only one to be thanked, Allah is the first and the last, also those offered by Allah to advice and guide have to be thanked.

I wish to express my deepest gratitude to **Prof. Dr. Khaled Eid Sobhy**, Professor of Chest Diseases, Cario University, for his sincere guidance, kind supervision, continuous encouragement through the whole work.

I am very grateful to **Prof. Dr. Somaya Abd El-Latif Eissa**, Professor of Microbiology and Immunology, Cairo University, for her precious supervision.

I am very grateful to **Prof. Dr. Mohamed Galal Morsy**, Professor of Chest Diseases, Cairo University, for his constant assistance and valuable support.

I sincerely appreciate the aid of **Dr. Hamed Abd El-Hafez Abd Allah**, Lecturer of Chest Diseases, Cairo University, Without him, this work would not have been completed.

I am very grateful to **Dr. Nadiah Mohammed Madny**, Lecturer of Microbiology and Immunology, Cairo University, for her precious supervision.

Lastly I would like to thank my **Father** for his guidance and always being my backbone.

Aim of the Work

The aim of this study is to evaluate the efficacy of Quantiferon TB Gold in diagnosing tuberculous pleural effusion.

Contents

	Title	Page
Introduction		1

Title	Page
Aim of the Work	3
Review of Literature	4
- Chapter (I): Pleural Effusion	4
- Chapter (II): Tuberculous Pleural Effusion	11
- Chapter (III): Quantiferon	52
- Chapter (IV): Quantiferon TB Gold and Tuberculous Pleural Effusion	70
Patient and Methods	75
Results	94
Discussion	115
Summary and Conclusion	130
Recommendations	133
References	134
Appendix	_
Arabic Summary	_

List of Tables

Title	Page
Review of Literatures:	
Table (A): General Causes of Pleural Effusions	5
Table (B): Etiology of Transudative Effusions	8
Table (C): Etiology of Exudative Effusions	9
Table (D): Recommended Dosage of First-Line Anti-TB Drugs	
Patients and Methods:	
Table (E): A2 x 2 table for evaluating a screening test disease status	89

Title	Page
Results:	
Table (1): Age and Gender distribution between Tuberculous	
group and Non Tuberculous group	94
Table (2): Comparison of positive Quantiferon in blood test	0.0
results in TB group and Non TB group	96
Table (3): Comparison of positive Quantiferon test results in	
pleural fluid in TB group and Non TB group	97
Table (4): Comparison of Quantiferon in blood and pleural	
fluid in Tuberculous group	98
Table (5): Comparison of Quantiferon in blood and pleural	
fluid in control group (Non Tuberculous)	99
Table (6): Comparison between diagnostic tests in the	
Tuberculous group and the Control subgroups	100
Table (7): Results of ROC curve analysis of Quantiferon in	
blood for diagnosis of TB	104
Table (8): Results of ROC curve analysis of Quantiferon in	
pleural fluid for diagnosis of TB	106
Table (9): Results of ROC curve analysis of Tuberculin test for	
diagnosis of TB	108
Table (10): Results of ROC curve analysis of ADA for	
diagnosis of TB	110
Table (11): Pair-wise comparisons between the diagnostic tests	
for TB	112
Table (12): Shows area under the curve (AUC), standard error	
(SE), 95% confidence interval (95% CI) and results of	
z-test for pair-wise comparisons	112

List of Figures

Title	Page
Patients and Methods:	
Figure (I): Blood incubation steps	82
Figure (II): Fresh dilutions prepared of the kit standard for each ELISA session	84
Figure (III): ELISA steps Cellestis Limited (Australia) and Cellestis Inc	85
Figure (IV): The Roc curve.	92
Results:	
Figure (1): Bar chart representing the mean age in Tuberculous and Non Tuberculous patients	95
Figure (2): Bar chart representing the Gender distributions in Tuberculous and Non Tuberculous patients	95
Figure (3): Bar chart representing the positive results of Quantiferon in blood in Tuberculous and Non Tuberculous group	96
Quantiferon in pleural fluid in Tuberculous and Non Tuberculous group	97
Figure (5): Bar chart representing mean ADA in Tuberculous and Non Tuberculous subgroups	102
Figure (6): Bar chart representing Tuberculin test results in Tuberculous and Non Tuberculous subgroups	102
Figure (7): Bar chart representing Quantiferon in fluid results in Tuberculous and Non Tuberculous subgroupsFigure (8): Bar chart representing Quantiferon in blood test	103
results in Tuberculous and Non Tuberculous subgroups.	103
Figure (9): ROC curve for Quantiferon in blood	105
Figure (10): ROC curve for Quantiferon in fluid	107
Figure (11): ROC curve for Tuberculin test	109
Figure (12): ROC curve for ADA	111

Title Page

Figure (13): Diagrammatic chart representing comparison

between ROC curves of different diagnostic tests 113

List of Abbreviations

ADA : Adenosine deaminase.

ARDS : Acute respiratory distress syndrome.

CD : Cluster of differentiation.

CFP-10 : Culture filtrate protein 10.

CHF : Congestive heart failure.

CR3 : Complement receptor 3.

CT : Computed tomography.

ELISA : Enzyme linked immunosorbent assay.

EPTB: Extra pulmonary tuberculosis.

ESAT-6 : Early secretory antigenic target-6.

HCW: Health care workers.

HIV : Human immunodeficiency virus.

IGRAS : Interferon gamma release assays.

IL: Interleukin.

INFδ : Interfeone gamma.

LDH : Lactate de hydrogenase enzyme.

NK: Natural killer.

PBMCs: Peripheral blood mononuclear cells.

PPD: Purified protein derivative.

QFN-TB: Quantiferon-TB Gold-test.

QFT-G-IT: Quantiferon-TB Gold in tube method.

TACO: Tryptophan aspirate coat protein.

TB: Tuberculosis.

TLR : Toll like receptor.

TST: Tuberculin skin test.

Chapter (I) Pleural Effusion

Anatomy of the Pleura:

The pleura is the serous membrane that covers the lung parenchyma, the mediastinum, the diaphragm, and the rib cage. This structure is divided into the visceral pleura and the parietal pleura. The visceral pleura covers the lung parenchyma, not only at its points of contact with the chest wall, diaphragm, and mediastinum but also in the inter-lobar fissures. The parietal pleura lines the inside of the thoracic cavities. In accordance with the intrathoracic surfaces that it lines, it is subdivided into the costal, mediastinal, and diaphragmatic parietal pleura (*Light* 2007).

A film of fluid (pleural fluid) is normally present between the parietal and the visceral pleura. This thin layer of fluid acts as a lubricant and allows the visceral pleura covering the lung to slide along the parietal pleura lining the thoracic cavity during respiratory movements. The space, or potential space, between the two layers of pleura is designated as the pleural space (*Light*, 2007).

The normal pleural space is approximately 18 to 20 mm in width, although it widens at its most dependent areas (Albertine et al., 1991).

The pleural membranes do not touch each other and that the pleural space

is a real, not a potential, space, it is likely that the primary function of the pleural membranes is to allow extensive movement of the lung relative to the chest wall. If the lung adhered directly to the chest wall, its expansion and deflation would be more limited (*Deschamps and Rodarte*, 1988).

Pathogenesis of Pleural Effusions:

Pleural fluid accumulates when the rate of pleural fluid formation exceeds the rate of pleural fluid absorption. The main factors that lead to increased pleural fluid formation or decreased pleural fluid absorption are tabulated in Table (A). Normally, a small amount (0.01 mL/kg/hour) of fluid constantly enters the pleural space from the capillaries in the parietal pleura. Almost all of this fluid is removed by the lymphatics in the parietal pleura, which have a capacity to remove at least 0.20 mL/kg/hour (Light, 2007).

Table (A): General Causes of Pleural Effusions (Light, 2007).

• Increased pleural fluid formation:	
- Increased interstitial fluid in the lung	
■ Left ventricular failure, pneumonia, and pulmonary embolus	
- Increased intravascular pressure in pleura	
■ Right or left ventricular failure, superior vena caval syndrome	
- Increased permeability of the capillaries in the pleura	
■ Pleural inflammation	
■ Increased levels of vascular endothelial growth factor	
- Increased pleural fluid protein level	
- Decreased pleural pressure	
 Lung atelectasis or increased elastic recoil of the lung 	
- Increased fluid in peritoneal cavity	
■ Ascites or peritoneal dialysis	
- Disruption of the thoracic duct	
- Disruption of blood vessels in the thorax	
• Decreased pleural fluid absorption:	
- Obstruction of the lymphatics draining the parietal pleura	
- Elevation of systemic vascular pressures	
Superior vena caval syndrome or right ventricular failure	
- Disruption of the aquaporin system in the pleura	

For pleural fluid to accumulate to form an effusion, it is likely that both the entry rate of liquid must increase and the exit rate must decrease. If only the entry rate increased, it would require a sustained rate more than 30 times normal to exceed the reserve lymphatic removal capacity; if the exit rate decreased, it would take more than a month at the normal entry rate of 12 mL/day to produce an

effusion detectable by chest radiograph thus, in the clinical setting, it is most likely that excess pleural fluid accumulates due to changes in both entry and exit rates (*Broaddus*, 2008).

Increased entry of fluid may result from increased filtration across systemic or pulmonary capillaries or entry of another fluid (e.g., chyle, cerebrospinal fluid [CSF], urine, intravenous fluids). Decreased exit of fluid may result from interference with lymphatic function (e.g., obstruction of the parietal pleural stomata, inhibition of lymphatic contractility, infiltration of draining parasternal lymph nodes, or elevation of the systemic venous pressure into which the lymph drains) (Broaddus, 2008).

There are few studies on the rate of removal of liquid in humans; however, decreases in lymphatic clearance have been confirmed in patients with tuberculous and malignant effusions and in those with the yellow nail syndrome, a disease of lymphatic function *(Leckie and Tothill, 1965)*.

Pathogenesis of Transudative and Exudative Pleural Effusion:

In some cases, it is likely that the same disease process acts both to increase the entry and to decrease the exit of fluid, whereas, in other cases, different disease processes may act cooperatively to produce an effusion to determine the origin of effusions, a classic and useful distinction is between transudates and exudates (Runyon et al., 1979).

Transudates form by leakage of fluid across an intact capillary barrier owing to increases in hydrostatic pressures or decreases in osmotic pressures across that barrier. Transudates generally indicate that the pleural membranes are not themselves diseased. Exudates results from leakage of fluid and protein across an altered capillary barrier with increased permeability. The protein ratio, the lactate dehydrogenase (LDH) ratio, and the absolute pleural LDH concentration constitute Light's criteria (Light et al., 1972).

Transudates include various low-protein fluids that arise from non injured capillary beds. The majority of transudates are caused by CHF. These transudates have been shown to result from leakage of edema across normal pulmonary

capillaries into the pulmonary interstitium and then across the leaky visceral pleura into the pleural space *(Wiener and Broaddus, 1993)*.

Other transudates, those associated with the nephrotic syndrome or atelectasis, may form because of altered pressures (osmotic or hydrostatic) across the pleural capillaries. Some transudates, usually small, may develop primarily because of an isolated decrease in exit rate (*Broaddus*, 2002).

Hepatic hydrothorax and effusions from peritoneal dialysis develop when fluid flows from the peritoneal space into the lower pressure pleural space across macroscopic holes in the diaphragm (Wong et al., 2009).

Finally, other very low protein fluids such as urine or CSF or intravenous liquids may find their way to the pleural space if their normal course is disrupted (*Broaddus*, 2008).

Exudates arise from injured capillary beds, in the lung, the pleura, or adjacent tissues. Most exudates, such as those associated with pneumonia or pulmonary embolism, probably form following lung inflammation and injury when a high-protein lung edema leaks into the pleural space. Another large category of exudates arises from pleural injury due to inflammation, infection, or malignancy. Exudates can also form when exudative liquid in the mediastinum (esophageal rupture or chylothorax), retroperitoneum (pancreatic pseudocyst), or peritoneum (ascites with spontaneous bacterial peritonitis or Meigs' syndrome) finds its way into the lower-pressure pleural space (*Broaddus*, 2008).

As stated, for either transudates or exudates, lymphatic obstruction may contribute to the accumulation of the effusion. Nonetheless, because lymphatic clearance does not alter the pleural fluid protein concentration, the protein concentration gives information about the formation of the fluid, not its removal (*Broaddus*, 2008).

Table (B): Etiology of Transudative Effusions (Sahnl, 2008).

- Congestive heart failure	Urinothorax
- Cirrhosis	Meig's syndrome
- Nephrotic syndrome	Pulmonary embolism
- Peritoneal dialysis	Superior vena cava obstruction

- Myxedema	Hypoalbuminaemia
- Atelectasis(early)	

Table (C): Etiology of Exudative Effusions (Light, 2007).

Do non non one o	Simple
Parapneumonic	Complicated
Tuberculosis	Hypersensitivity reaction in hematogenous phase
	Empyema as complication of post primary pulmonary T.B
Other infections	Fungal
	Parasitic
Wallamant	Metastatic disease
Malignant	Mesothelioma
Pulmonary Embolism	
	Rheumatoid arthritis
	Systemic lupus erythematosus
Collagen Vascular disease	Wegener's granulomatosis
	Churg-Strauss syndrome
	Familial Mediterranean fever
	Pancreatitis
Abdominal diseases	Subphrenic abscess
	Postoperative
	Atelectasis
	Acute respiratory distress syndrome, (ARDS)
Miscellaneous	Asbestos exposure
	Hemothorax
	Chylothorax
	Cholesterol effusions
	Esophageal rupture
	Radiation therapy
	Ovarian hyperstimulation syndrome

Light Criteria for Exudative Pleural Effusion:

Transudative and exudative pleural effusions are distinguished by measuring the lactate dehydrogenase (LDH) and protein levels in the pleural fluid. Exudative pleural effusions meet at least one of the following criteria, whereas transudative pleural effusions meet none:

- 1. Pleural fluid protein/serum protein >0.5.
- 2. Pleural fluid LDH/serum LDH > 0.6.

3. Pleural fluid LDH more than two-thirds normal upper limit for serum.

The above criteria misidentify approximately 25% of transudates as exudates. If one or more of the exudative criteria are met and the patient is clinically thought to have a condition producing a transudative effusion, the difference between the albumin levels in the serum and the pleural fluid should be measured. If this gradient is greater than 12 g/L (1.2 g/dL), the exudative categorization by the above criteria can be ignored because almost all such patients have a transudative pleural effusion. If a patient has an exudative pleural effusion, the following tests on the pleural fluid should be obtained: description of the fluid, glucose level, differential cell count, microbiologic studies, and cytology (*Light*, *2002*).

Chapter (II) **Tuberculous Pleural Effusion**

Tuberculosis (TB) is a leading cause worldwide of preventable morbidity and mortality from an infectious agent. The interaction of HIV with Mycobacterium Tuberculosis has lead to its resurgence in developed nations and has increased the burden of TB cases in developing countries (Who, 2006).

TB pleural effusion, considered as a form of extrapulmonary TB (EPTB), constitutes a frequent clinical problem *(Sharma, 2004)*.

It's particularly important in the present era of HIV infection, when EPTB is more commonly encountered in clinical practice (*Harries, 1990*).

A pleural effusion occurs in approximately 5% of patients with TB *(Siebert et al., 1991)*.

Epidemiology:

A total of nine million new cases and approximately two million deaths from TB were reported in 2004 *(Who, 2006)*.

Although the African region has the highest estimated incidence (356 per 100,000 population per year), the majority of patients with TB live in the most populous countries of the Asian subcontinent, which accounts for nearly half of the new cases that arise yearly *(Dye, 2006)*.