

***THE RELATION BETWEEN SKIN
DISORDERS AND VITAMIN D***

Essay

**Submitted for Partial Fulfillment of Master Degree in
Dermatology, Venereology and Andrology**

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2015

*Introduction and
Aim of the Work*

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Vitamin D is a prohormone produced in the skin through ultraviolet irradiation of 7-dehydrocholesterol. It is biologically inert and must be metabolized to 25-hydroxyvitamin D in the liver and then to 1 α ,25-dihydroxyvitamin D in the kidney before function. The hormonal form of vitamin D, ie; 1 α ,25-dihydroxyvitamin D, acts through a nuclear receptor to carry out its many functions, including calcium and phosphate absorption in the intestine, calcium mobilization in the bone, and calcium reabsorption in the kidney. In addition, vitamin D has several non-calcemic functions in the body (*Deluca, 2004*).

This overview provides a brief description of the physiologic, endocrinologic, and molecular biologic characteristics of vitamin D. It also provides information about new selective therapeutic analogues of vitamin D and highlights the role of vitamin D in pathogenesis and treatment of some inflammatory, infectious, autoimmune, neoplastic and other skin disorders.

Review of Literature

Chapter (1) Vitamin D

Chapter (1)

1. Vitamin D

1.1. Introduction

Vitamin D is a secosteroid (i.e., A steroid in which one of the bonds in the steroid rings is broken), fat-soluble prohormone of critical importance for a broad variety of independent physiological functions (*DeLuca, 2004; Wolpowitz and Gilchrest, 2006; Holick, 2007*). It was identified after the discovery of the anti-rachitic effect of cod liver oil (*Wolpowitz and Gilchrest, 2006*). It is considered as a key regulator of bone metabolism and calcium and phosphorous homeostasis through a negative feedback with the parathyroid hormone (*PTH*) (*DeLuca, 2004; Wolpowitz and Gilchrest, 2006; Holick, 2007*). It also regulates the growth and differentiation of multiple cell types, and displays a number of immunoregulatory and anti-inflammatory properties (*Adorini and Penna, 2008*).

It acts via binding to a corresponding intranuclear receptor [vitamin D receptor (VDR)], in target tissues (*Stumpf et al., 1979; Gniadecki, 1996*). Nearly, every tissue in the body has receptors for the active form of vitamin D (*Di Rosa et al., 2011*), including cells involved in innate and adaptive immune responses which can both produce and respond to the active form of vitamin D (*Adorini and Penna, 2008*).

1.2.Nomenclature of vitamin D precursors and metabolites

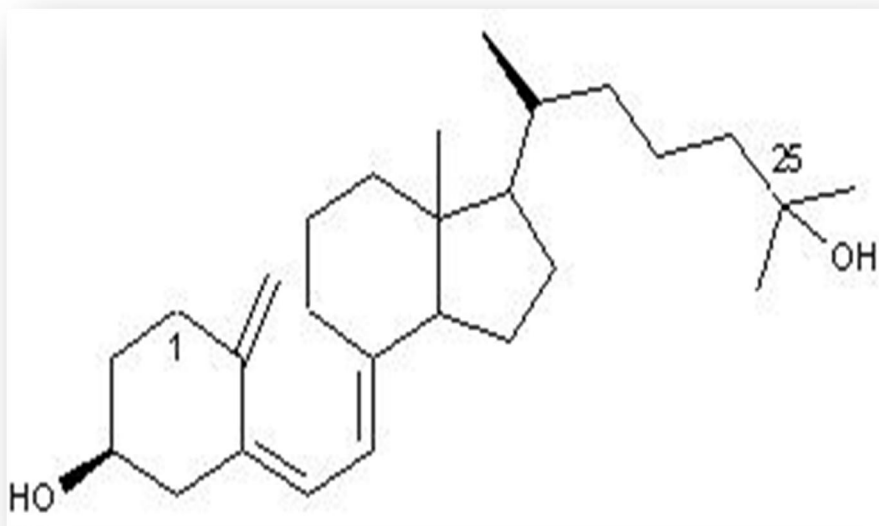
Vitamin D exists in nature in different forms as shown in table (1).

Table (1): Different forms, nomenclature and sources of vitamin D (Wolpowitz, and Gilchrest, 2006).

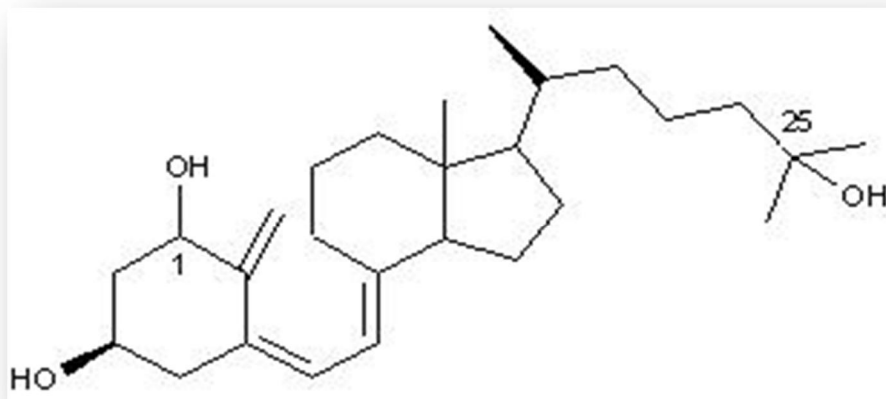
Common name	Clinical name	Abbreviation	Comments
7-Dehydrocholesterol	Provitamin D3	7-DHC	Lipid in cell membranes
Cholecalciferol	Previtamin D3	Previt D3	Photosynthesized in skin or obtained from diet
Ergocalciferol	Previtamin D2	Previt D2	Obtained from diet; equivalent to vit D3 as precursor for active vit D
Calcidiol	25-Hydroxyvitamin D	25-(OH) vit D	Circulating “storage” form of vit D, biologically inactive
Calcitriol	1,25-Dihydroxyvitamin D	1,25-(OH) ₂ vit D	Active form of vit D, tightly regulated

The two main forms of vitamin D in blood are **25(OH)D (Calcidiol) (fig.1)**, the principal circulating form of vitamin D and **1,25(OH)₂D(calcitriol) (fig. 2)**, the principal active hormonal form of vitamin D and which is responsible for most of its biologic actions (**Bikle,2009**).

The difference in the chemical structure of the active metabolites is situated in the side chain (**fig.1,2**) (**Feldman et al., 2005; Hollis and Wagner, 2004**). 1,25(OH)₂D, is different from 25(OH) D in that it possesses an additional 1-alpha hydroxylation. This structural difference alters binding to the carrier protein [vitamin D binding protein (DBP)] as well as metabolism (**Bikle, 2009**).



(Fig.1): The Chemical Structure of Calcidiol (**Deluca, 2004**).

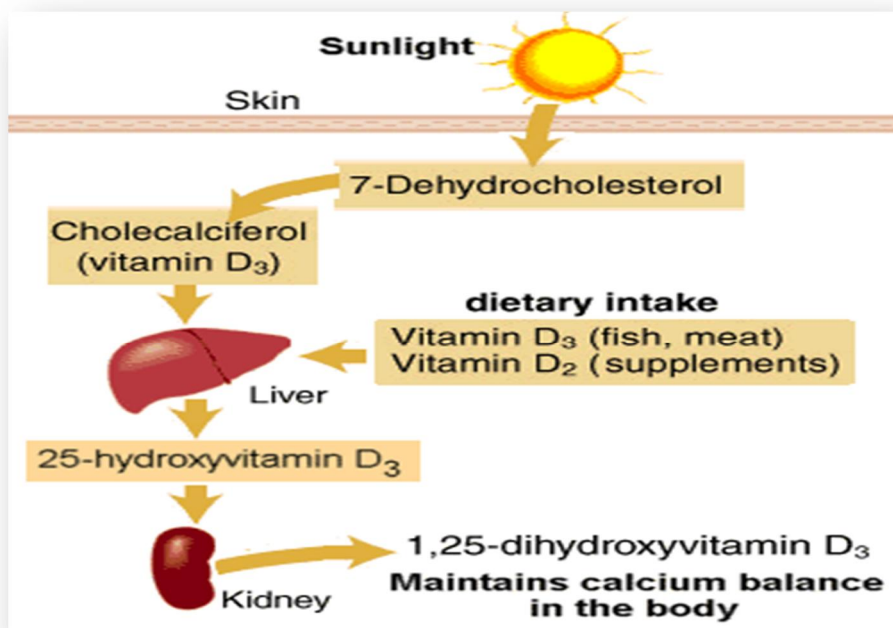


(Fig.2): The Chemical Structure of Calcitriol (*Deluca, 2004*).

1.3. Sources of Vitamin D

Vitamin D ($1, 25(\text{OH})_2\text{D}$) is produced in relatively large quantities in humans and in the majority of vertebrate animals. The main production of $1, 25(\text{OH})_2\text{D}$ occurs in the course of photosynthesis (Photochemical conversion of 7-dehydrocholesterol (7-DHC) to previtamin D in the skin), in the presence of ultraviolet (UV) light (*Di Roso et al., 2011*) during summer months (*Feldman et al., 2005; Hollis and Wagner, 2004*). Only a few foods naturally contain appreciable amounts of vitamin D: fish liver, fish liver oils, fatty fish, and egg yolks. Oily fish such as salmon, mackerel, and bluefish are excellent sources of vitamin D (*Lu et al., 2007; Hollis and Wagner, 2004*) (fig.3). Some countries practice fortification of certain foods with vitamin D, most often milk, margarine, and/or butter. The mean intake of

vitamin D, in different studies varies with age group, food, supplementation habits and gender (Lu *et al.*, 2007; Hollis and Wagner, 2004).



(Fig.3): Vitamin D sources (Hollis and Wagner, 2004).

1.4. Level of Vitamin D in adult serum

Although 1, 25(OH)₂D is the biologically active form of vitamin D, its half-life is less than 4 hours. In fact, 1, 25(OH)₂D may remain normal or even increase in vitamin D-deficient states (Dlugos *et al.*, 1995 ; Holick, 2004), so, It is universally accepted that the circulating level of 25-hydroxyvitamin D should be used as an indicator of vitamin D because it is easy to measure, has long half-life in circulation (approximately 2 or 3 weeks), and there is

correlation between its level and clinical disease states (*Adams et al., 1982; Reichel et al., 1989; Wolpowitz and Gilcrest, 2006*).

Vitamin D deficiency is defined as a level of 25-hydroxyvitamin D of less than 20 ng per milliliter (50 nmol per litre) (*Thomas et al., 1998; Holik et al., 2005; Lips et al., 2006; Michael, 2007*). A level of 25-hydroxyvitamin D of 21 to 29 ng per millilitre (52 to 72 nmol per litre) is considered as an insufficiency of vitamin D, and sufficient vitamin D should reach a level of 30 ng per millilitre or greater (*Dawson-hughes et al., 2005*).

On the other hand, vitamin D intoxication is extremely rare. Studies showed that doses of more than 50,000 IU per day, which raises 25-hydroxyvitamin D to more than 150 ng per milliliter, is associated with hypercalcemia and hyperphosphatemia(*Bouillon, 2001; Holick and Garabedian., 2006; Holik, 2006*).

So, the serum values of 25-hydroxyvitamin D are interpreted as follows:

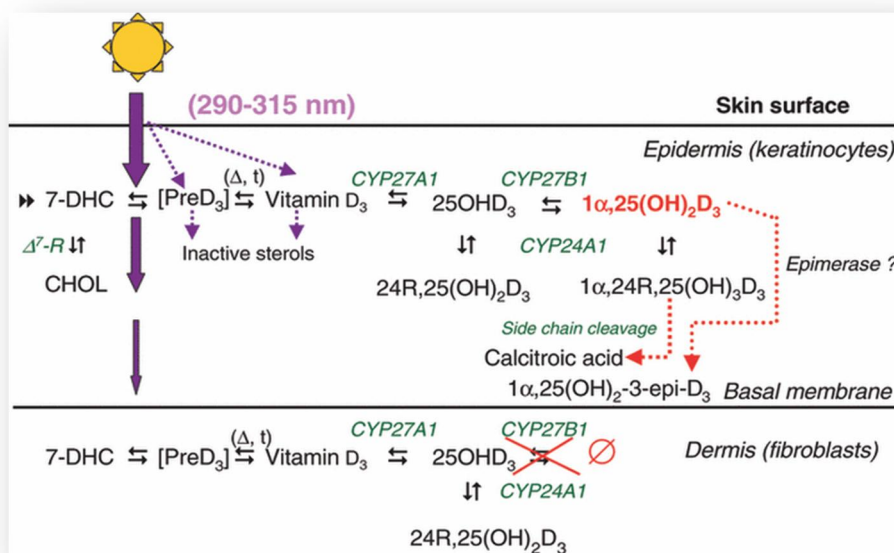
- <20 ng/mL (<50 nmol/L): deficient.
- 21–29 ng/mL (51–74 nmol/L): insufficient.

- >30 ng/mL (>75 nmol/L): sufficient.
- >150 ng/mL: intoxication (*Holick and Chen, 2008*).

1.5. Metabolism of Vitamin D

1.5.1. Photochemical reactions in the skin

Human epidermal keratinocytes contain the complete machinery needed to produce the hormone $1\alpha,25(\text{OH})_2\text{D}$ (calcitriol) from its initial precursor 7-dehydrocholesterol (7-DHC) (*fig.4*). These cells also express the nuclear vitamin D receptor (VDR) that mediates the effects of this hormone on keratinocytes. Thus, it is reasonable to assume that calcitriol acts in an autocrine and paracrine manner in the epidermis (*Lehmann, 2009*).



(Fig.4): Vitamin D pathway in epidermal keratinocytes (*Lehmann, 2009*).

7-dehydrocholesterol (7-DHC) is already stored in the basal and suprabasal layers of skin (*Holick et al., 1980; Mac Laughlin et al., 1982*), and is photolyzed to previtamin D (calciferol) during exposure to ultraviolet (UV) light (*Prosser and Jones, 2004; Birlea et al., 2008*). The photochemical reaction is maximum at wave length spectrum from 297 to 302 nm (*Holick et al., 1980*). So, vitamin D synthesis is almost confined to the UVB region (290 - 320 nm) and occurs at minimal rates in the UVA region (320 - 400 nm) (*Wolpowitz and Gilchrest, 2006*).

If newly formed previtamin D continues to be irradiated, it is converted into additional products such as lumisterol 3, and tachysterol 3 that do not possess vitamin D activity any more, i.e.:. Excessive sunlight exposure cannot cause vitamin D toxicity because UVB converts excess vitamin D to biologically inert isomers (*Prosser and Jones, 2004; Birlea et al., 2008*).

1.5.2. Limiting factors of the cutaneous vitamin D synthesis

There is a biochemical equilibrium between 7-DHC and cholesterol adjusted by the activity of 7-DHC-D7-reductase in epidermal keratinocytes. Thereafter, the conversion of 7-DHC into previtamin D in the skin depends on several individual and environmental factors (*Chen et al., 2007; Lehmann, 2009*):

1.5.2.1. The concentration of 7-dehydrocholesterol (7-DHC) in the skin. The 7-DHC level in normal human adult skin ranges between 1.9 and 75 µg/cm² (*Lehmann, 2009*). Under normal physiological circumstances, human skin has ample quantities of 7-DHC available in the stratum spinosum and stratum basale and which are mainly regulated by the activity of the 7-DHC-D7-reductase that catalyzes the conversion of 7-DHC to cholesterol and vice versa (*Bonjour et al., 1987*),

1.5.2.2. *The energy of photons* that – in turn - depends upon the wavelength of the ultraviolet rays,

1.5.2.3. *Both the solar zenith angle (which is a function of latitude and season) and time of the day.* Solar zenith means the distance between the sun and the earth,

1.5.2.4. *Skin pigmentation* which is determined by the concentration of melanin in the skin. Melanin, which absorbs UVB in the 290–320 nm range, functions as a light filter and, therefore, determines the proportion of the incident UVB that is actually able to penetrate the outer epidermal layers and arrive at the stratum basale and stratum spinosum (*Chen et al., 2007*).

1.5.2.5. *Use of sunscreens*, which considerably suppresses photolysis of 7-DHC.

1.5.2.6. *Temperature*, which regulates the enzymatic conversion of previtamin D to vitamin D, and

1.5.2.7. *Age*, as there is an inverse relation between the epidermal concentrations of 7-DHC and the age (*Feldman et al., 2005*).

1.5.3. Vitamin D activation and degradation pathways

After being synthesized, vitamin D is translocated to the circulation where it binds to vitamin D binding protein (DBP) to reach the peripheral tissues (*Prosser and Jones, 2004; Birlea et al., 2008*).

•*Transport in Blood*

Vitamin D metabolites are transported in blood bound primarily to vitamin D binding protein (DBP) (85-88%) and albumin (*Bikle et al., 1984; Cooke and Haddad, 1989*). Vitamin D binding protein (DBP) concentrations are normally 4-8 times the concentration of the vitamin D metabolites, so that DBP is only about 2% saturated. DBP has high affinity for the vitamin D metabolites, such that under normal circumstances only approximately 0.03% of

25(OH)D and 24, 25(OH)₂D and 0.4% of 1, 25(OH)₂D are free (*Bikle et al., 1984; Bikle et al., 1986*). Conditions such as liver disease and nephrotic syndrome, usually associated with reduced DBP and albumin levels will lead to a reduction in total 25(OH)D and 1, 25(OH)₂D levels, without necessarily affecting the free concentrations (*Bikle et al., 1986*).

However, vitamin D intoxication can increase the degree of saturation sufficiently to increase the free concentrations of 1, 25(OH)₂D and so cause hypercalcemia without necessarily raising the total concentrations (*Pettifor et al., 1995*).

Vitamin D binding protein (DBP) was originally known as group specific component (Gc-globulin) before its properties as a vitamin D transport protein became known. It has three common polymorphisms which are useful in population genetics but which do not appear to alter its function. DBP is a 58kDa protein with 458 amino acids that is homologous to albumin and α -fetoprotein (α FP). It is made primarily-but not exclusively-in the liver. Other sites include the kidney, testes, and fat (*Horiuchi et al., 1977*). DBP like other steroid hormone binding proteins is increased by oral (not transdermal) estrogens and