Detection Of *Blastocystis hominis* In Stools Of Patients Attending Beni Suief University Hospital

Thesis

Submitted In Partial Fulfillment Of M.Sc.Degree In Parasitology

By

Doaa Ahmed Hamdy Mohamed M.B.,B.Ch.

Supervisors

Prof. Dr. Hoda Abd El Megid El Bolaky

Professor Of Parasitology Faculty Of Medicine Cairo University

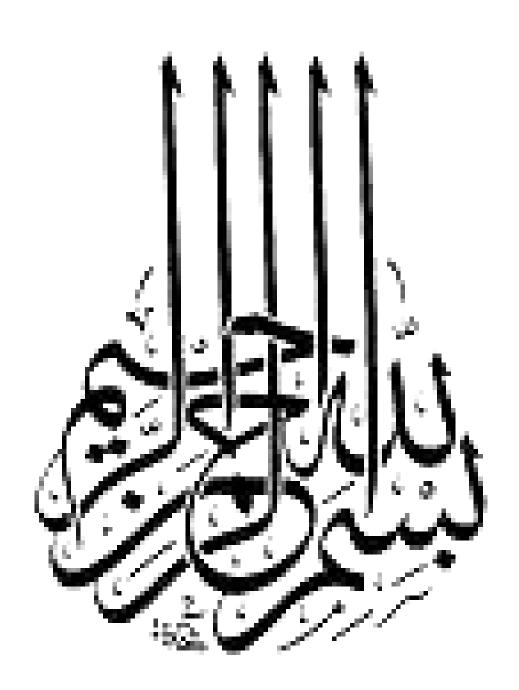
Prof. Dr. Olfat Mohamed El Matrawy

Professor Of Parasitology Faculty Of Medicine Cairo University

Dr. Hanaa Mohamed Ezzat Moussa

Lecturer Of Parasitology Faculty Of Medicine Cairo University

Faculty Of Medicine Cairo University 2009



Dedication

To the soul of my parents, to my husband, my daughter, my sisters and my brother who gave me all love and support.....

<u>Acknwledgement</u>

First and foremost thanks are due to ALLAH the most beneficent and merciful.

I would like to express my deepest thanks to **Prof. Dr. Hoda Helmy El Rahimy** Head of the Department Of Parasitology, Faculty

Of Medicine, Cairo University, for her advice and help.

My deepest appreciation goes to **Prof. Dr. Hoda Abd El Megid El Bolaky** Professor Of Parasitology, Faculty Of Medicine, Cairo
University, for her encouragement, interest, valuable advice and
constructive criticism throughout the present work.

I am also grateful to **Prof. Dr. Olfat Mohamed El Matrawy**Professor Of Parasitology, Faculty Of Medicine, Cairo University, for her kind supervision, guidance and help.

My thanks also to **Dr. Hanaa Mohamed Ezzat Moussa**Lecturer Of Parasitology, Faculty Of Medicine, Cairo University,
for her cooperation, advice and help in the laboratory work.

Finally, I would like to express my warm feelings to the staff members of Parasitology Department, Faculty Of Medicine, Cairo University.

List of Abbreviations

AIDS Acquired immunodeficiency syndrome

B.hominis Blastocystis hominis

HPF High power field

P Probability value

ROC Receiver operating characteristic

List of Figures

Figure	Page No.
Figure (1): Proposed life cycle of <i>B.hominis</i> by Singh et	20
al., (1995).	
Figure (2): Vacuolar form of <i>B.hominis</i> stained with	58
iodine by X100 objective.	
Figure (3): Amoeboid form of <i>B.hominis</i> stained with	58
iodine by X100 objective.	
Figure (4): B.hominis three days after in-vitro cultivation	59
using Jones' medium by X100 objective.	
Figure (5): Vacuolar form of <i>B.hominis</i> stained with	59
Giemsa by X100 objective.	
Figure (6): Amoeboid form of <i>B.hominis</i> stained with	60
Giemsa by X 100 objective.	
Figure (7): Vacuolar form of <i>B.hominis</i> stained with	60
modified trichrome by X100 objective.	
Figure (8): Cyst form of <i>B.hominis</i> stained with modified	61
trichrome by X 100 objective.	
Figure (9): Granular form of <i>B.hominis</i> stained with	61
modified trichrome by X 100 objective.	
Figure (10): Vacuolar form of <i>B.hominis</i> stained with	62
modified safranin by X 100 objective.	
Figure (11): Granular form of <i>B.hominis</i> stained with	62
modified safranin by X 100 objective.	
Figure (12): B.hominis stained with modified safranin	63
showing binary fission by X 100 objective.	
Figure (13): Vacuolar form of <i>B.hominis</i> stained with	63
modified Ziehl Neelsen by X 100 objective.	
Figure (14): Granular form of <i>B.hominis</i> stained with	64
modified Ziehl Neelsen by X 100 objective	

List of Tables

Title	Page No.
Table (1): Prevalence of <i>B.hominis</i> infection in the 90 patients	44
attending the pediatric and tropical medicine outpatient clinics.	
Table (2): Presence of gastrointestinal symptoms in the 39	45
B.hominis infected cases.	
Table (3): Type of residential areas of <i>B.hominis</i> infected cases.	46
Table (4): The relation between <i>B.hominis</i> infected cases and	47
the close contact to animals.	
Table (5): B.hominis infected cases among different age groups.	48
Table (6): Relation between infected immunosuppressed cases	49
with <i>B.hominis</i> and those infected only by the parasite among	
different age groups.	
Table (7): The relation between mixed infected cases with	50
B.hominis and those infected only by the parasite.	
Table (8): Detection of positive <i>B.hominis</i> cases among 90	52
patients using different diagnostic techniques. Sensitivity and	
specificity and p values as referred to in-vitro culture positive	
cases.	

List of Graphs

Title	Page No.
Graph (1): Comparison of ROC curves for diagnostic	53
performance of direct smear, concentration and staining	
techniques as referred to in-vitro culture.	
Graph (2): Comparison of ROC curves for diagnostic	55
performance of different staining techniques as referred to in-	
vitro culture.	

Abstract

Blastocystis hominis is a unicellular organism found commonly in the intestinal tract of humans and many other animals. Very little is known about the basic biology of this organism. The common morphological forms are vacuolar, granular, and amoeboid. Other additional forms were recently detected. Infections with the organism are worldwide and appear in both immunocompetent and immunodeficient individuals. Gastrointestinal symptoms generally attributed to blastocystosis infection. In the present study detection of this organism was done in Beni Suief University Hospital by using different diagnostic techniques which gave different variations in the obtained results.

(**Key words:** *Blastocystis hominis*, Intestinal parasites, Prevalence, Diagnosis).

Introduction

Introduction

Blastocystis hominis (**B.hominis**) is a single celled parasitic protozoan with a worldwide distribution. Its geographic distribution appears to be global, with infections common in tropical, subtropical and developing countries (**Minvielle et al., 2004**).

This protozoan can be transmitted as a cyst by the faecal-oral route, especially in areas with poor hygiene and sanitation (Logar et al., 1994 and Stenzel and Boreham, 1996).

Despite its prevalence throughout the world, its taxonomy remains unresolved till confirmed by analysis of some genes (Arisue et al., 2002).

The signs and symptoms commonly associated with *B.hominis* infection appear with other organisms. Experts were not sure whether it causes disease or merely serves as an indicator of other agents that might cause diarrhoeal symptoms (**Barret et al., 1999**).

B.hominis had been previously considered as a normal inhabitant of the intestinal tract, but recently it has been accepted as a controversial pathogenic parasite causing gastrointestinal symptoms (**Tan et al., 2006**).

Morphological heterogeneity is observed in *B.hominis* parasite. There are the amoeboid form which is commonly detected in stools of diarrhoeic patients and the vacuolated form which has been considered to be the typical cell form and is generally used for its diagnosis. There is also the granular form which has an ultrastructure similar to that of the vacuolar form (**Tan and Suresh**, **2006**).

Different diagnostic methods are used for detection of *B.hominis* infections, such as serologic testing which has been used with very limited success (Chen et al., 1987). However Garavelli et al., (1992) reported the success of the immunofluorescence assay and enzyme-linked immunosorbent assay in detecting serum antibodies to *B.hominis* infection.

In addition, staining and culturing of the parasite from faecal samples have been used in an attempt to identify it (Nascimento and Moitinho, 2005).

Recently various genetic analyses showed that there is no single species of *Blastocystis* that infects humans. **Stensvold et al.**, (2007b) proposed discontinuing the use of the term *B.hominis* and referred to *Blastocystis* from humans and animals as *Blastocystis* species.

Aim of The Work

Aim of The Work

The aim of the present work is to compare between different diagnostic techniques in detecting *Blastocystis hominis* in stools of gastrointestinal symptomatic patients attending Beni Suief University Hospital.

Review of Literature