IMPLEMENTATION OF BIOTECHNOLOGY FOR IMPROVING SALT STRESS TOLERANCE IN PLANT USING PRROTEOMIC STUDY

Submitted By

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A thesis submitted in Partial Fulfillment
Of
The Requirement for the Doctor of Philosophy Degree
In
Environmental Sciences

Department of Environmental Agricultural Sciences Institute of Environmental Studies and Research Ain Shams University

APPROVAL SHEET

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ABSTRACT

A proteomic approach was used to identify proteins affected by salt in the halophyte plant Atriplex nummularia L. A total of twenty seven protein spots were reproducibly detected and analyzed on 2 DE gel. These proteins showed changes under salinity condition in Atriplex leaves and were up-regulated. The identified protein spots were involved in biosynthesis, ATP generation, metabolism and photosynthesis. In an attempt to explore the data of the proteomic experiment, we have studied the effect of some compounds on wheat plant under salinity condition. The relevant of such compounds have been used as exogenous application under salinity. The selected compounds were proline, ascorbic acid, arginine, glutathione and glutamine. The results obtained from this experiment revealed that growth parameters (shoot length, root weight and grain weight), biochemical compounds (chlorophyll, carbohydrate, starch, protein, fiber, ash and fat content) and some antioxidant enzymes such as ascorbate peroxidase, peroxidase and catalase activity were affected by both salt and treatments. The selected compounds mitigate the negative effects of salt stress and improved growth parameters and biochemical compounds compared with control plants under different salinity level.

Key words: Salt stress, salt tolerance, proteomic approach, exogenous application, *Atriplex* plant and wheat plant.

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LIST OF ABBREVIATIONS

AAT Aspartate aminotransferase

ABA Abscisic acid

ABF₃ Abscisic acid responsive elements-Binding Factor 3 **ABF**₄ Abscisic acid responsive elements-Binding Factor 4

ALDHs Aldehyde dehydrogenase **APX** ascorbate peroxidase

ASA Ascorbic acid

ATP Adenine triphosphate

Ca Calcium

CBB Coomassie brilliant blue

CHCA a-cyano-4-hydroxycinnamic acid
CID Collision induced dissociation

Cl Chloride

CMO Choline mono-oxygenase

CO₂ Carbon dioxide

DHAR Dehydroascorbate reductase
 DHB Dihydroxybenzoic acid
 DNA Deoxyribonucleic acid
 DNA Deoxyribonucleic acid

DTT Dithiothreitol

ECD Electron capture dissociation

ESI Electrospray ionization
ESTs Expressed sequence tags

ETD Electron transfer dissociation
EUI elongated uppermost internode
FAD Flavin adenine dinucleotide

FBP aldolase Fructose-1,6-bisphosphatase aldolase

GAs Gibberellins
GB Glycinebetaine

GPX Glutathione peroxidase

GR Glutathione reductaseGS Glutamine synthetase

GSR Glutathione-disulfide reductase

H Hydrogen

 H_2O_2 Hydrogen peroxide

ha Hectare

HSP Heat-shock proteinsIAA indole-3-acetic acidIEF Isoelectric focusing

IPG Immobilized pH gradientIPG immobilized pH gradient

JA Jasmonic acid
K Potassium
KD Kilo Dalton

LC Liquid chromatography

Li Lithium

MALDI Matrix-assisted laser desorption/ionization

mgMilligramMgMagnesiumminMinutemlMilliliter

mRNA Messenger RNAMS Mass spectrometryMW Molecular weight

Na Sodium

NADPH Reduced nicotinamide adenine dinucleotide phosphate

OEEs Oxygen-evolving enhancer proteins
P5CS Pyrroline-5-carboxylate synthase

PA Picolinic acidpI Isoelectric point

PMSF Phenylmethyl sulfonyl fluoride

PPDK Orthophosphate dikinase

PSII Photosystem II

PTMs Post-translational modifications

RNA Ribonucleic acid

ROS Reactive oxygen species

SA salicylic acid

SAGE serial analysis of gene expression

SAM S-adenosyl-L-methionine

SAMS S-adenosyl-L-methionine synthase

SDS-PAGE Sodium dodecyl sulfate polyacrylamide gel

electrophoresis

SID Surface induced dissociation

SOD Superoxide dismutaseSOS Salt Overly SensitiveSOS Salt overly sensitive

TK Transketolase

TPI Triosphosphate isomeraseas **DMSP** Dimethyl sulfonium propironate

ng Nanogram

PMF Peptide mass fingerprinting

TOF Time-of-flight

2 DE gel Two-dimensional gel electrophoresis

I. INTRODUCTION

Crop plants are often exposed to various biotic and abiotic stresses that greatly reducing the productivity of the crop worldwide. Salinity is a major abiotic stress limiting crop productivity worldwide and factor seriously affecting crop production in different regions, particularly in arid and semi–arid regions and affects more than 800 million hectares of land, equivalent to more than 6% of the total global area of the Earth (Munns and Tester, 2008; Taffouo *et al.*, 2009; Deinlein *et al.*, 2014; Carmelina and Stefania 2016).

Salt stress causes detrimental effects on crop yield by altering plant metabolism, including reduced water potential, ion imbalances and toxicity (**Krishnamurthy** *et al.*, **2016**). Understanding the molecular mechanisms underlying the plant's response to salinity stress will facilitate efforts to develop crop plants with enhanced resistance to high salinity (**Peng** *et al.*, **2014**). Hence, improved salt tolerance of crops has also become an urgent task to reduce the spread of salinity and sustain increases in food production worldwide (**Munns**, **2005**).

Actually, plants have evolved sophisticated mechanisms to cope with salinity stress. The salt signal is primarily perceived through roots, which rapidly respond and transmit signals to the shoot for appropriate changes in function, regulating the transcription and translation of intracellular genes associated with stress response, ultimately generating a series of physiological and biochemical responses in plants (Lv et al., 2016).

Plants have been divided into salt-tolerant 'halophytes' and salt-sensitive 'glycophytes', depending on growth performance of these Plants in saline habitats. Glycophytes display severely subdued growth and even death in