

**Immunological Activity of Snail
Hemocytes as Biomarkers of
Environmental Quality**

Presented by

Mohamed Mohsen Mohamed Mohamed

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Thesis Title: Immunological activity of Snail Hemocytes as Biomarkers of Environmental Quality

Name of candidate: Mohamed Mohsen Mohamed Mohamed

This thesis has been approved for submission by the supervisors:

Signature

1- Prof. Dr. Abdel-Hakim Saad El-Din

Professor of Immunology
Faculty of Science, Cairo University
Cairo, Egypt

2- Prof. Dr. Abdel-Rahman Basther

Professor of Parasitology
Faculty of Science, Cairo University
Cairo, Egypt

3- Prof. Dr. Ragia Ali Charmy

Professor of Immunology
Faculty of Science, Cairo University
Cairo, Egypt

Prof. Dr. Kawther Sayed Abou EL Ala

**Chairman of Zoology Department
Faculty of Science- Cairo University**

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List of abbreviations

| | |
|--------------------|------------------------------------|
| °C | Degrees Celsius |
| AAS | Atomic Absorption Spectroscopy |
| Ab | Antibody |
| BSA | Bovine serum albumin |
| D | Degeneration |
| DC | Digestive Cells |
| DHC | Different Hemocyte Counts |
| DNA | Deoxyribonucleic acid |
| DT | Digestive gland Tubules |
| E.M | Electron Microscope |
| EC | Epithelial Cells |
| ELISA | Enzyme Linked Immunosorbent Assays |
| F | Filaments |
| G | Granulocytes |
| Hsp 70 | Heat shock protein-70 |
| Ig | Immunoglobulin |
| IL-2 | Interleukin-2 |
| IL-6 | Interleukin-6 |
| INF- γ | Interferon gama |
| m | Mitochondria |
| M | Mucous |
| <i>M. edentula</i> | <i>Macoma edentula</i> |
| MF | Muscle Fibers |
| MFO | Mixed Function Oxygenases |
| n | Nucleus |
| PCBs | polychlorinated Biphenyls |

| | |
|---------------------|----------------------------------|
| PHAs | Polycyclic Aromatic Hydrocarbons |
| <i>R.decussatus</i> | <i>Ruditapes decussatus</i> |
| RER | Rough Endoplasmic Reticulum |
| S | Secretion |
| SC | Secretory gland Cells |
| SER | Smooth Endoplasmic Reticulum |
| THC | Total Hemocyte Counts |
| TNF- α | Tumor necrosis factor alpha |
| UV | Ultra violet |
| V | Vacuolation |

Abstract

ABSTRACT

The aim of the present study was to investigate some hematological, histopathological and immunological parameters in the clams, *Ruditapes decussatus* (Veneridae) and *Macoma edentula* (Tellinidae) to serve as a reference for studies in the monitoring of different environmental pollutants through the potential change in the clam defence activities. Two localities of the Lake Timsah exhibiting a similar pattern of seasonal variation in hemocyte abundance and a distinct degree of environmental contamination were selected for a comparative study of *in vitro* hemocyte activities. The results of DNA strand breakage, were expressed as the mean tail DNA content and the mean tail length, The two measured comet parameters in both species, showed a significant differences ($P > 0.05$) between the unpolluted and polluted groups. Hsp 70 activity of *R. decussatus* and *M. edentula* was studied for natural variability related to water pollution, Hsp 70 activity was significantly higher in normal unpolluted than polluted samples. Water pollution was observed to cause significant histopathological changes in the mantle of both snails species by using electron and light microscopic studies. Histopathological changes of mantle from both *R. decussatus* and *M. edentula* are represented in the arrangement or regularity of different cells. Vacuolation and swelling of the gill filaments occurred and distortion and damage of gill filament also occurred. The amount of secretion of glandular cells increased in polluted *R. decussatus*. The secretion was significantly decreased in *M. edentula* and the cells became completely necrotic and vacuolation occurred in most parts of digestive tubules. Later on, digestive gland tubules completely damaged. Polluted *R. decussatus* and *M. edentula* showed a statistically significantly, higher levels of IL-2 than non-polluted samples. The levels of IL-6 in both the polluted *R. decussatus* and *M. edentula* were significantly lower than the non-polluted control. The INF- γ levels in the polluted *R. decussatus* exhibited a non-significant increase as compared with non-polluted animals. However, the levels of INF- γ in the polluted *M. edentula* exhibited significant elevation as compared with non-polluted animals. The polluted *R. decussatus* showed low levels of TNF- α than non-polluted control. Also, the levels of TNF- α in the polluted *M. edentula* were significantly decreased as compared to non-polluted animals. These results suggest that the parallel use of polluted and native bivalve in environmental biomonitoring can improve the characterization of the study area.

Key words: Mollusc bivalve; *Ruditapes decussatus*; *Macoma edentula*; DNA damage; Comet assay; Hemocytes; Mantle; Gills; Digestive gland; Immune function; Biomarkers; Ecotoxicology.

Introduction

INTRODUCTION

The immune system of invertebrates is relatively simple in contrast to that of vertebrates, providing accessible means for monitoring immune function. Unlike vertebrates, invertebrates lack immune specificity and memory, characteristic of the adaptive immunity observed in mammalian subjects. Invertebrates have evolved innate defence mechanisms that can identify and protect against non-self material, with invertebrate immune response centered largely on the multifunctional hemocytes (**Cheng, 1981**). The various parameters that contribute to invertebrate host defence act to provide a multifaceted immune response. It has been suggested that an integrated suite of biomarkers can be used for assessing immunocompetence of invertebrates (**Pipe and Coles, 1995**).

Advances in environmental risk assessment have highlighted the value of biomarkers to allow rapid evaluation of contaminant exposure and effect (**Galloway *et al.*, 2002**). These biomarkers may be molecular, cellular or physiological endpoints that indicate exposure to and/or damage incurred by environmental pollutants providing early-warning indicators of contaminant-induced stress. The use of biomarkers has been recognized as a cost-effective technique in identifying the effects of pollutants on biota (**Galloway *et al.*, 2002**).

The majority of biomarker studies to date have been based on the mussel *Mytilus edulis*, and the American oyster *Crassostrea virginica* (**Wootton *et al.*, 2003**). Bivalve exhibit worldwide distribution and sedentary, filter feeding behavior. They have an open circulatory system that is continually exposed to fluctuating environmental conditions and