

**VON WILLEBRAND FACTOR AS NEW NONINVASIVE PREDICTOR
OF PORTAL HYPERTENSION, DECOMPENSATION AND
MORTALITY IN PATIENTS WITH LIVER CIRRHOSIS**

Thesis

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List of Contents

<i>Title</i>	<i>Page No.</i>
List of Abbreviations.....	ii
List of Tables	iv
List of Figures	v
Introduction	i
Aim of the Work	9
Review of Literature	
von Willebrand Factor	10
Chronic Liver Diseases.....	31
Subjects and Methods	63
Results.....	67
Discussion.....	78
Summary and Conclusion.....	86
Recommendations	90
References	91
Arabic Summary	—

List of Abbreviations

ALT	Alanine aminotransferase
AFP	Alpha feto protien
AMP	Adenosine mono-phosphate
AST	Aspartate aminotransferase
CBC	Complete Blood Count
CK	Cystine knot
CLDs	Chronic liver diseases
CSPH	Clinically significant portal hypertension
CT	Computerize Tomography
DDAVP	Desamino-8-D-arginine vasopressin
ECM	Extracellular matrix
ECs	Endothelial cells
EIA	Enzyme immunoassay
FVIII	Factor VIII
GGT	Glutamyl transpeptidase
GP	Glycoprotein
HBV	Hepatitis B virus
HCC	Hepatocellular carcinoma
HCV	Hepatitis C virus
HDV	Hepatitis D virus
HGV	Hepatitis G virus
HMM	High molecular mass
HMW_vWF	High molecular weight

List of Abbreviations (Cont...)

HSCs	Hepatic stellate cells
HVPG	Hepatic venous pressure gradient
INR	International normalized ratio
IRMA	Immunoradiometric assay
IQR	Inter quartile range
MELD	Model for end stage liver disease
MRI	Magnetic resonance imaging
NASH	Non-alcoholic steatohepatitis
PFV	Portal flow velocity
PH	Portal hypertension
PT	Prothrombin time
PVD	Portal vein diameter
RCO	Restocetin cofactor
TAV	Time averaging flow velocity
TE	More recently, transient elastography
TIPS	Transjugular intrahepatic portosystemic shunt
v WF	von Willebrand factor
v WF-Ag	von Willebrand factor antigen
vWF: Ag II	vWF antigen II
vWFpp	vWF propeptide

List of Tables

<i>Table No.</i>	<i>Title</i>	<i>Page No.</i>
Table (1):	Hepatitis Viruses causing chronic hepatitis:.....	36
Table (2):	Common Patterns of Laboratory Test Abnormalities	42
Table (3):	Scoring of Child-Turcotte-Pugh Score:	56
Table (4):	Interpretation of the Child-Turcotte-Pugh Score:	56
Table (5):	Descriptive Statistics in case group.....	71
Table (6):	Descriptive Statistics in control group.	72
Table (7):	Comparison between case group and control group.	72
Table (8):	Comparison between Patients with and without ascites.	73
Table (9):	Comparison bet. Patients with and without varices:	73
Table (10):	Comparison between patients with and without shrunken liver:	74
Table (11):	Correlation between vWF and studied parameters in cases.	75
Table (12):	Correlation between VWF and studied parameters in control.....	75
Table (13):	Diagnostic Validity Test: between patients with PFV <13 cm/sec & those ≥13 cm/sec.	76

List of Figures

<i>Fig. No.</i>	<i>Title</i>	<i>Page No.</i>
Fig. (1):	Structure and domains of von Willebrand factor (vWF) and Type 2 mutations which are located in specific domains (regions).....	11
Fig. (2):	The structure of vWF factor	13
Fig. (3):	Biosynthesis of vWF.....	14
Fig. (4):	The site of vWF biosynthesis	18
Fig. (5):	Schematic representations of the mechanisms of platelet adhesion.....	22
Fig. (6):	The role of vWF in platelet adhesion and aggregation	25
Fig. (7):	Model of vWF –FVIII complex in plasma.....	27
Fig. (8):	Structure of Hepatitis C virus.	36
Fig. (9):	Portal vein diameteris measured where it crosses anterior to IVC.....	59
Fig. (10):	Portal vein peak velocity which is shown to be 21 cm/sec.	60
Fig. (11):	Portal vein time averaging flow velocity (TAV) measurement show: 16cm/s mean velocity of portal vein flow.....	61
Fig. (12):	ROC curve analysis showing the diagnostic performance of vWF for discriminating patients with PVV<13 from those >13.....	77

INTRODUCTION

Portal hypertension (PH) accounts for the major complications of liver cirrhosis, such as ascites, variceal hemorrhage and decompensation. Early diagnosis of PH is essential for the management of patients with cirrhosis (*Thabut et al., 2011*).

Clinically significant portal hypertension (CSPH), {HVPG (hepatic venous pressure gradient) 10mmHg}, is associated with a higher risk of liver-related mortality, development of varices, and other PH related complications. An HVPG 12 mmHg is associated with a higher risk of bleeding from varices (*Bosch et al., 2006*).

Normally, portal blood flows towards the liver (hepatopetal flow), In normal patients the mean PV (portal velocity) flow rate is 13 to 23 cm/sec but in patients with portal hypertension it tend to decrease, the mean PV (portal vein) velocity may vary depending on the presence and location of spontaneous shunts (*Al-Nakshabandi et al., 2006*).

More recently, transient elastography (TE) was described as a non invasive tool for the diagnosis of PH in patients with liver cirrhosis, but the costs and availability of TE represent limiting factors in smaller hospitals. Thus, the recent Baveno V consensus conference on PH recommended to investigate and

identify further noninvasive markers for PH (*De Franchis et al., 2010*).

Hepatic decompensation is the most important predictor of prognosis and mortality in patients with liver cirrhosis, with several precipitating factors contributing to the first event of decompensation (*D'Amico et al., 2006*).

Endothelial cells dysfunction is considered as an important determinant of the increased intrahepatic vascular resistance in cirrhotic livers (*Iwakiri et al., 2007*).

von Willebrand factor antigen (v WF-Ag) is released by activated endothelial cells (ECs) and therefore represents an indicator of EC activation and plays a crucial role in high shear stress, depending on primary hemostasis. Furthermore, in patients with liver cirrhosis, elevated levels of vWF-Ag are frequently observed (*Lisman et al., 2006*).

Although it is established that VWF-Ag is increased in patients with cirrhosis, no data on the association of vWF-Ag and portal pressure exist (*La Mura et al., 2011*).

Because vWF-Ag plays a crucial role in primary hemostasis and is an indicator of endothelial cells activation and development of thrombotic vascular obliteration, which are all discussed as possible mechanisms leading to PH, it is hypothesized that patients with CSPH have increased vWF-Ag levels, compared to patients without CSPH (*Ferlitsch et al., 2012*)

AIM OF THE WORK

The aim of this work is to evaluate the diagnostic performance of vWF-Ag to detect clinically significant PH correlated by portal vein velocity in patients with liver cirrhosis and to evaluate vWF-Ag levels in the prediction of mortality and decompensation.

VON WILLEBRAND FACTOR

Over the last 20 years, investigators have gained much insight into the structure and function of vWF by studying its molecular and cellular biology. Such studies have helped us understand the clinical manifestations of vWF and develop assays for laboratory diagnosis, the classification of vWD and its variants, and the rationale for therapy (*Colman et al., 2006*).

A) vWF structure

vWF is a large multimeric glycoprotein present in blood plasma and produced by the endothelium (in the Weibel-Palade bodies), megakaryocytes (α -granules of platelets), and subendothelial connective tissue (*Zaverio, 2007*).

The vWF gene has been localized to chromosome 12, contains 52 exons, and has a length of about 178 kb. Transcription of the gene results in mRNA of approximately 8.7 kb (*Baronciani et al., 2003*).

The primary product of the vWF gene is a 2813 amino acid protein made of a signal peptide of 22 amino acids (also called a pre peptide) synthesized by endothelial cells and megakaryocytes. Post translational processing cleaves the pro-vWF peptide into a large pro-peptide of 741 amino acids and a

mature vWF molecule of 270 kDa containing 2050 amino acids (*Castaman et al., 2003*). The pro-peptide (pre-pro-vWF) composed of four types of repeated domains (D1, D2, D', D3, A1, A2, A3, D4, B1, B2, B3, C1, C2 from the N- to C-terminal region) of cDNA which are responsible for the different binding functions of the molecule (Figure 1) (*Franchini and Lippi, 2007*).

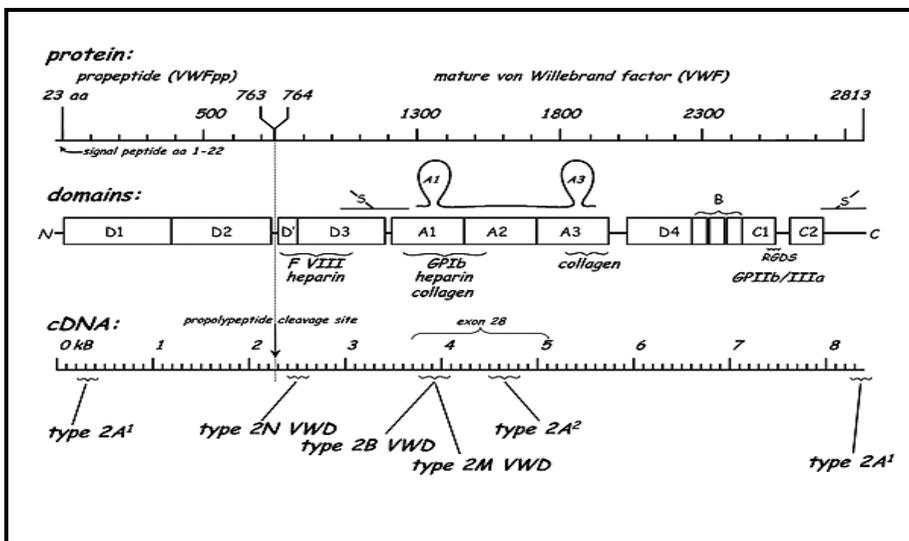


Fig. (1): Structure and domains of von Willebrand factor (vWF) and Type 2 mutations which are located in specific domains (regions) (*Nichols et al., 2008*).

Monomers of mature vWF are assembled into variable sized multimers via disulfide bridging of cysteine residues at both the amino and carboxy-terminal ends. Under physiological conditions, smaller complexes constitutively exit the cell and enter the circulation, where they bind and stabilize the

coagulation factor VIII or undergo reduction and proteolytic cleavage. The size of circulating vWF multimers is also regulated through processing by the metalloprotease ADAMTS13. Large complexes, with up to 100-fold activity of monomeric fragments, are essential for normal platelet adhesion and aggregation after vascular injury in rapid-flow, high shear stress vessels such as arterial capillaries (*Kulkarni et al., 2000*).

The basic vWF monomer (subunit) consists of 2050 amino acid. Every monomer contains a number of specific domains with a specific function:

- The D'/D3 domain, which binds to FVIII.
- The A1 domain also contains the binding site for the platelet glycoprotein GP Ib–IX–V complex, different matrix collagens.
- The A3 domains bind to different matrix collagens (*Sadler et al., 2000*).
- The C1 domain, in which binds to the platelet.
- The C2 domain is close to the carboxy terminal region of vWF and interacts with the activated GP IIb–IIIa complex (Figure 2) (*Ruggeri et al., 1998*).

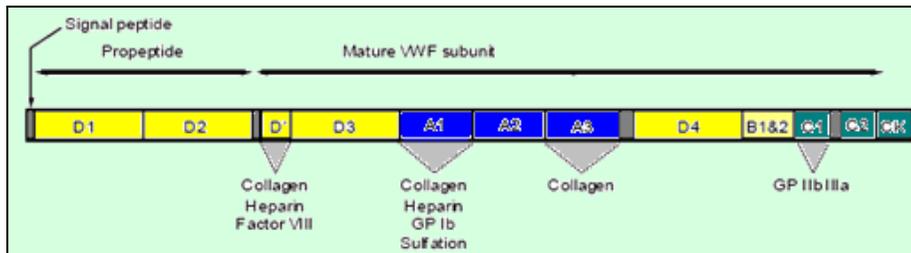


Fig. (2): The structure of vWF factor (*Carol, 2008*).

The building block of vWF multimers is a dimer formed in the endoplasmic reticulum and made up of two single chain pro-vWF molecules, joined through disulphide bonds within their C-terminal region. The pro-vWF dimers are then transported to the Golgi apparatus where they are polymerized into very large molecules, with molecular weights of up to 20, 000 kDa, through disulphide bonds connecting the two N-terminal ends of each dimer (*Franchini and Lippi, 2007*).

B) vWF Biosynthesis:

vWF circulates in plasma at concentration ranging between 5 and 10 $\mu\text{g/ml}$. Some of these molecules are complexed with FVIII, apparently protecting FVIII against degradation (*Volt et al., 2000*).

vWF is synthesized by vascular endothelial cells and megakaryocytes (*Mannucci., 2000*), and stored in the Weibel-Palade bodies of endothelial cells and in α -granules of platelets.

In physiologic conditions, vWF is mainly synthesized by the endothelial cell (Figure 3) (*Mertens et al., 2006*).

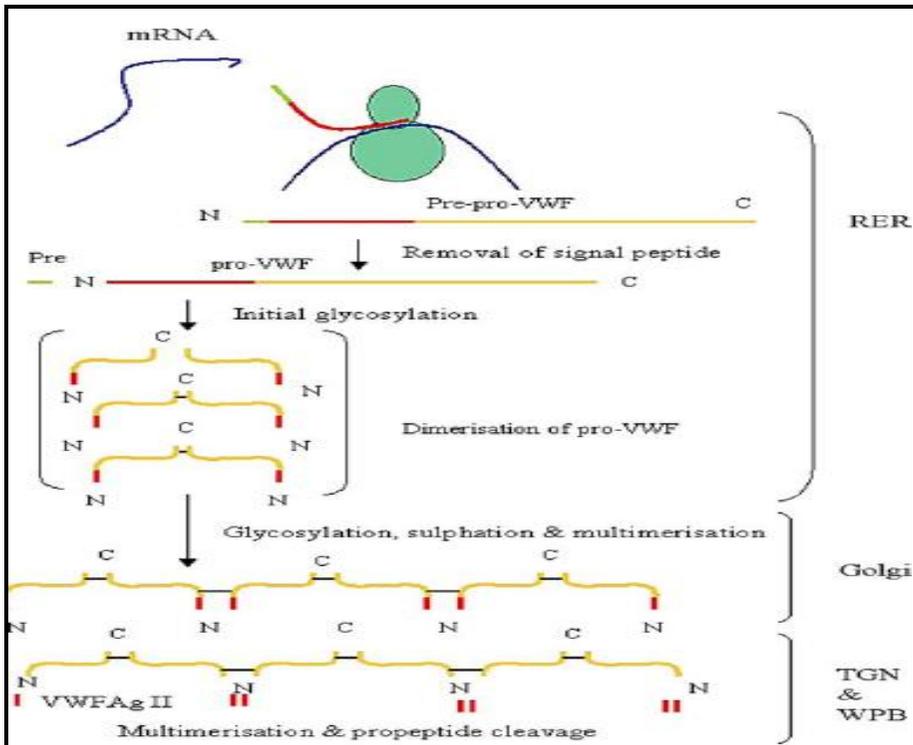


Fig. (3): Biosynthesis of vWF (*Millar and Brown et al., 2006*).

Small vWF multimers are secreted constitutively, and the large multimeric forms are stored in Weibel–Palade bodies of endothelial cells and in platelet α -granules (*Fuchigami, 2008*).

1. Endothelial Synthesis:

The primary translation product of the mRNA is a 2813 residue pre-pro-peptide, known as pre-pro- vWF. This consists