

# **A Cytokine Profile in Response to Stimulation of Peripheral Blood Mononuclear Cells by HCV C200**

## **Thesis**

Submitted for Partial Fulfillment of Master Degree  
in Clinical and Chemical Pathology

**By**

**Anwar Hassan Elsayed**  
M.B., B.Ch. Zagazig University

**Under Supervision of**

**Prof. / Mona Mohamed Rafik**

Professor of Clinical and Chemical Pathology  
Faculty of Medicine – Ain Shams University

**Prof./ Shahera Fathy El Fedawy**

Professor of Clinical and Chemical Pathology  
Faculty of Medicine – Ain Shams University

**Dr./ Khaled Omar Abdallah**

Assistant Professor of Clinical and Chemical Pathology  
Faculty of Medicine – Ain Shams University

**Faculty of Medicine  
Ain Shams University  
2010**

# لمحة عن استجابة المؤثرات الخلوية بعد تحفيز الخلايا الطرفية وحيدة النواة باستخدام ببتيد سي 200 الخاص بالالتهاب الكبدى الفيروسي سى

## رسالة

توطئة للحصول على درجة الماجستير  
فى الباثولوجيا الإكلينيكية والكيميائية

## مقدمة من

الطبيبة/أنوار حسن السيد  
بكالوريوس الطب والجراحة  
جامعة الزقازيق

## تحت إشراف

ا.د /منى محمد رفيق

أستاذ الباثولوجيا الإكلينيكية والكيميائية  
كلية الطب – جامعة عين شمس

ا.د/ شهيرة فتحى الفداوى

أستاذ الباثولوجيا الإكلينيكية والكيميائية  
كلية الطب – جامعة عين شمس

د/خالد عمر عبد الله

أستاذ مساعد الباثولوجيا الإكلينيكية والكيميائية  
كلية الطب – جامعة عين شمس

كلية الطب  
جامعة عين شمس

2011

# **contents**

|   |            |
|---|------------|
| <b>Introduction .....</b>                   | <b>1</b>   |
| <b>Aim of the work .....</b>                | <b>4</b>   |
| <b>Review of literature .....</b>           | <b>5</b>   |
| <b>Subjects and Methods .....</b>           | <b>74</b>  |
| <b>Results .....</b>                        | <b>85</b>  |
| <b>Discussion .....</b>                     | <b>106</b> |
| <b>Conclusion and Recommendations .....</b> | <b>116</b> |
| <b>Summary.....</b>                         | <b>117</b> |
| <b>References.....</b>                      | <b>121</b> |
| <b>Arabic summary .....</b>                 | <b>-</b>   |

## List of Tables

|   | Page                                |
|---|-------------------------------------|
| <b>Table (1):</b> Descriptive Statistics: medians and percentiles of each cytokine in three groups                                  | <b>Error! Bookmark not defined.</b> |
| <b>Table (2):</b> Comparison between the cytokine levels in the studied groups (+ve P.I HCWs, -ve P.I HCWs & Chronic HCV HCWs)..... | <b>Error! Bookmark not defined.</b> |
| <b>Table (3):</b> Comparison between the cytokine levels in chronic HCV HCWs and -ve P.I HCWs:                                      | <b>Error! Bookmark not defined.</b> |
| <b>Table (4):</b> Comparison between the cytokine levels in chronic HCV HCWs and +ve P.I HCWs:                                      | <b>Error! Bookmark not defined.</b> |
| <b>Table (5):</b> Comparison between the levels of cytokines in +ve and -ve P.I HCWs.....   | <b>Error! Bookmark not defined.</b> |
| <b>Table (6):</b> Correlation between levels of cytokines and cellular CD markers among +ve P.I HCWs:                               | <b>Error! Bookmark not defined.</b> |
| <b>Table (7):</b> Correlation between levels of cytokines and cellular CD markers among -ve P.I HCWs:                               | <b>Error! Bookmark not defined.</b> |
| <b>Table (8):</b> Correlation between levels of cytokines and cellular CD markers among Chronic HCV HCWs:                           | <b>Error! Bookmark not defined.</b> |
| <b>Table (9):</b> Correlation between different cytokines within individual groups: .....   | <b>Error! Bookmark not defined.</b> |

# List of Figures

|  | Page                                |
|--|-------------------------------------|
| <b>Fig. (1):</b> Structure of Hepatitis C Virus  | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (2):</b> The HCV genome and gene products  | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (3):</b> Circulating HCV particles can be associated with low- and very-low-density lipoproteins | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (4):</b> Life cycle of hepatitis C virus   | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (5):</b> Global Prevalence of Hepatitis C Virus  | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (6):</b> Sources of Infection for Persons with Hepatitis C in USA                                | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (7):</b> Geographic distribution of hepatitis C viral species                                    | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (8):</b> HCV attenuates innate immune response   | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (9):</b> Secretion of IFN by pDCs after contact with HCV-infected hepatocytes                    | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (10):</b> Crosstalk between NK cells and dendritic cells   | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (11):</b> Principle of the assay   | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (12):</b> Materials provided   | <b>Error! Bookmark not defined.</b> |
| <b>Fig. (13):</b> Comparison between the cytokine levels in the studied groups.                          | <b>Error! Bookmark not defined.</b> |

- Fig. (14):** Regression analysis showing the correlation between IFN-Gama and CD3-<sub>CD8+</sub> among +ve P.I HCWs group**Error!**  
**Bookmark not defined.**
- Fig. (15):** Regression analysis showing the correlation between IFN-Gama and CD3-CD8- among +ve P.I HCWs group**Error!**  
**Bookmark not defined.**
- Fig. (16):** Regression analysis showing the correlation between IFN-Gama and CD3+CD8- among +ve P.I HCWs group**Error!**  
**Bookmark not defined.**
- Fig. (17):** Regression analysis showing the correlation between IL-2 and CD3- CD8- among +ve P.I HCWs group.....**Error!**  
**Bookmark not defined.**
- Fig. (18):** Regression analysis showing the correlation between IL-2 and CD3+CD8- among +ve P.I HCWs group.....**Error!**  
**Bookmark not defined.**
- Fig. (19):** Regression analysis showing the correlation between IL-2 and CD3-CD8+ among +ve P.I HCWs group.....**Error!**  
**Bookmark not defined.**
- Fig. (20):** Regression analysis showing the correlation between TNF-Alfa and CD3-<sub>CD8+</sub> among +ve P.I HCWs group**Error!**  
**Bookmark not defined.**
- Fig. (21):** Regression analysis showing the correlation between TNF-Alfa and CD3+CD8+ among +ve P.I HCWs group**Error!**  
**Bookmark not defined.**
- Fig. (22):** Regression analysis showing the correlation between TNF-Alfa and CD3+<sub>CD8-</sub> among +ve P.I HCWs group**Error!**  
**Bookmark not defined.**
- Fig. (23):** Regression analysis showing the correlation between TNF-Alfa and CD3+<sub>CD8+</sub> among -ve P.I HCWs group**Error!**  
**Bookmark not defined.**
- Fig. (24):** Regression analysis showing the correlation between TNF-Alfa and CD3+<sub>CD8-</sub> among -ve P.I HCWs group**Error!**  
**Bookmark not defined.**

- Fig. (25):** Regression analysis showing the correlation between IFN-Gama and CD3+\_CD8- among chronic group.....**Error!**  
**Bookmark not defined.**
- Fig. (26):** Regression analysis showing the correlation between IFN-Gama and IL-2 among +P.I HCWs group**Error! Bookmark not defined.**
- Fig. (27):** Regression analysis showing the correlation between IFN-Gama and TNF\_Alfa among +ve P.I HCWs group **Error!**  
**Bookmark not defined.**
- Fig. (28):** Regression analysis showing the correlation between IL-2 and TNF\_Alfa among +ve P.I HCWs group.....**Error!**  
**Bookmark not defined.**
- Fig. (29):** Regression analysis showing the correlation between IL-2 and TNF\_Alfa among -ve P.I HCWs group.....**Error!**  
**Bookmark not defined.**
- Fig. (30):** Regression analysis showing the correlation between IL-2 and IFN-Gama among -ve P.I HCWs group.....**Error!**  
**Bookmark not defined.**

## List of Abbreviations

|                                |   |
|--------------------------------|---|
| <b>aa</b>                      | : amino acid                                      |
| <b>Ag</b>                      | : Antigen   |
| <b>ALT</b>                     | : Alanine aminotransferase                        |
| <b>APCs</b>                    | : Antigen presenting cells                        |
| <b>ARF</b>                     | : Alternate reading frame                         |
| <b>ARFP</b>                    | : Alternate Reading Frame Protein                 |
| <b>AST</b>                     | : <a href="#">Aspartate transaminase</a>          |
| <b>bDNA</b>                    | : Branched DNA                                    |
| <b>CD</b>                      | : Cluster of differentiation                      |
| <b>CDC</b>                     | : Centers for disease control and prevention      |
| <b>CFSE</b>                    | : Carboxyfluorescein diacetate succinimidyl ester |
| <b>CMI</b>                     | : Cell mediated immunity                          |
| <b>CTL</b>                     | : cytotoxic T lymphocytes                         |
| <b>D</b>                       | : Domain  |
| <b>DCs</b>                     | : Dendritic Cells                                 |
| <b>DNA</b>                     | : deoxyribonucleic acid                           |
| <b>dsRNA</b>                   | : Double stranded RNA                             |
| <b>E</b>                       | : Envelope glycoprotein                           |
| <b>EIA</b>                     | : Enzyme immunoassay                              |
| <b>ELISA</b>                   | : Enzyme-Linked Immunosorbent Assay               |
| <b>ELISPOT</b>                 | : Enzyme-linked immunospot                        |
| <b>ER</b>                      | : Endoplasmic reticulum                           |
| <b>F</b>                       | : Frameshift                                      |
| <b>HCC</b>                     | : Hepatocellular carcinoma                        |
| <b>HCV</b>                     | : Hepatitis C Virus                               |
| <b>HCWs</b>                    | : Health care workers                             |
| <b>HLA</b>                     | : Human leukocyte antigens                        |
| <b>HS</b>                      | : Highly Significant                              |
| <b>HVR</b>                     | : Hypervariable region                            |
| <b>IFN<sub>I</sub></b>         | : Type I interferon                               |
| <b>IFN-<math>\alpha</math></b> | : Interferon alpha                                |
| <b>IFN-<math>\beta</math></b>  | : Interferon beta                                 |
| <b>IFN-<math>\gamma</math></b> | : Interferon-gamma                                |
| <b>IgG</b>                     | : Immunoglobulin G                                |



|                    |  |
|--------------------|--|
| <b>IKK</b>         | : Inhibitor of $\kappa$ B kinase               |
| <b>IL-2</b>        | : Interleukine-2                               |
| <b>IL-2R</b>       | : IL-2 receptors                               |
| <b>IPS-1</b>       | : IFN- $\beta$ promoter stimulator protein 1   |
| <b>IRES</b>        | : Internal ribosomal entry site                |
| <b>IRF</b>         | : Interferon regulatory factor                 |
| <b>ISDR</b>        | : IFN- $\alpha$ sensitivity determining region |
| <b>ISGF</b>        | : IFN stimulated gene factor                   |
| <b>ISGs</b>        | : Interferone stimulation genes                |
| <b>ISREs</b>       | : IFN-stimulated response elements             |
| <b>JAK</b>         | : Janus tyrosine kinase                        |
| <b>KIR</b>         | : Killer inhibitory receptors                  |
| <b>LFTs</b>        | : <a href="#">Liver Function Tests</a>         |
| <b>MDCs</b>        | : Myeloid DCs                                  |
| <b>MHC</b>         | : Major histocompatibility complex             |
| <b>MICA/B</b>      | : MHC class-I related chain A/B                |
| <b>NK</b>          | : Natural killer cells                         |
| <b>NKT</b>         | : Natural killer T cells                       |
| <b>NS</b>          | : Non Significant                              |
| <b>NS proteins</b> | : non structural proteins                      |
| <b>NTPase</b>      | : Nucleoside triphosphatase activities,        |
| <b>PBMCs</b>       | : Peripheral blood mononuclear cells           |
| <b>PCR</b>         | : Polymerase chain reaction                    |
| <b>PD-1</b>        | : Programmed death 1                           |
| <b>PDCs</b>        | : Plasmacytoid DCs                             |
| <b>PD-L</b>        | : programmed death ligand                      |
| <b>PI</b>          | : Proliferation index                          |
| <b>PIAS</b>        | : Protein inhibitor of activated STAT1         |
| <b>PKR</b>         | : dsRNA-dependent Protein Kinase R             |
| <b>PP2A</b>        | : Protein phosphatase 2A                       |
| <b>QPCR</b>        | : Quantitative PCR                             |
| <b>RdRp</b>        | : RNA-dependent RNA polymerase                 |
| <b>RFU</b>         | : relative fluorescence units                  |
| <b>RIBA</b>        | : Recombinant immunoblot assay                 |
| <b>RIG-I</b>       | : Retinoic acid-inducible gene I               |
| <b>RNA</b>         | : Ribonucleic acid                             |
| <b>RT</b>          | : Reverse Transcriptase                        |

|                  |   |
|------------------|---|
| <b>S</b>         | : Significant   |
| <b>Sig</b>       | : Significant   |
| <b>SOCS</b>      | : Suppressor of cytokine signaling  |
| <b>SP</b>        | : Signal peptidase  |
| <b>SPSS</b>      | : Statistical Package for Social Sciences   |
| <b>SR-BI</b>     | : Scavenger receptor B type I   |
| <b>STAT</b>      | : Signal transducer and activator of transcription  |
| <b>T regs</b>    | : <a href="#">Regulatory T cells</a>  |
| <b>TBK1</b>      | : TNF receptor– associated factor family member–associated NF-κB activator–binding kinase–1   |
| <b>TCR</b>       | : T cell receptor   |
| <b>Th cells</b>  | : T helper cells  |
| <b>TLR</b>       | : Toll like receptor  |
| <b>TMA</b>       | : Transcription mediated amplification  |
| <b>TMD</b>       | : Transmembrane domain  |
| <b>TNF-α</b>     | : Tumor necrosis factor-alfa  |
| <b>TRIF</b>      | : Toll–IL-1 receptor domain–containing adaptor inducing IFN-β<br>NF-κB nuclear factor kappa B |
| <b>TYK2</b>      | : Tyrosine kinase 2   |
| <b>UTR</b>       | : Untranslated regions  |
| <b>γGT</b>       | : Gamma Glutamyl Transferase  |
| <b>2'-5' OAS</b> | : 2'-5'Oligoadenylate synthetase  |
| <b>5-PL</b>      | : Five-Parameter Logistic curve   |



## Introduction

Hepatitis C virus (HCV) infection has become a global health problem with around 170–190 million infected people worldwide (*Berenguer, 2007*).

Spontaneous viral clearance of HCV is observed in only 15–40% of patients with acute hepatitis whereas, persistent infection is established despite evidence of immune regulation and is associated with progression to cirrhosis and hepatocellular Carcinoma (*Shiffman, 2003*).

Hepatitis C virus has structural proteins like core protein and envelope glycoproteins E1 and E2, which mediate entry into cell by binding to CD81 as well as the non-structural (NS) proteins, which have essential functions in viral replication. NS3 contains protease, RNA helicase and nucleoside triphosphatase (NTPase) activities, all of which are essential to viral replication. NS4A acts as cofactor for N3 protease activity. NS4B has a role in the formation of HCV RNA replication complex (*Inoue et al., 2007*).

Strong and persistent cell mediated immune responses have been reported in HCV seronegative individuals with documented exposure to HCV in the absence of detectable viral RNA (*Post et al., 2004*).

HCV-specific CD4<sup>+</sup>T cell responses persist in acutely infected individuals who permanently cleared the virus, and disappear in patients whose viraemia subsequently recurred (*Chang et al., 2001*).

All patients with self limited disease had a significant CD4<sup>+</sup> T-cell proliferation to HCV specific C200 peptide, running parallel with the antigen-stimulated secretion of IL-2 and IFN-  $\gamma$  but not with IL-4 and IL-10, indicating predominant Th1 response (*Semmo and Klenerman, 2007*).

HCV-specific CD8<sup>+</sup>T cell responses that are associated with spontaneous viral clearance tend to be multi-specific and polyclonal (*Lauer et al., 2005*).

Because of the high variability of HCV, escape mutations in CD8<sup>+</sup>T cell epitopes are common and they are expected to play a major role in chronic viral persistence (*Ray et al., 2005*).

HCV-specific CD8<sup>+</sup>T cells have both cytolytic and non cytolytic effector function. The non cytolytic function is mediated by production of Interferone gamma (*Guidotti et al., 2001*).

The imbalance between T helper cell Th1 (IL-2, IL-12, IFN- $\gamma$ ) cytokines and Th2 (IL-10) cytokines affect the outcome of HCV infection. The defect in both IL-12 and IFN- $\gamma$  production may contribute to persistence of HCV infection (*Sarih et al., 2000*).

IL-2 was capable of both pushing semi-effector CTL to complete its effector cell program and restoring the HCV core-dependent inhibitory effect (*Accapezzato et al., 2004*).

TNF- $\alpha$  triggers a partially overlapping set of antiviral defense mechanisms and serum level of TNF- $\alpha$  reflects the progression of inflammation (*Akyüz et al., 2005*).

Interferon- gamma (IFN- $\gamma$ ) is a key cytokine and can inhibit HCV replication (*Lanford et al., 2003*).

## **Aim of the work**

The aim of the work is to detect a panel of cytokines IL2, IFN- $\gamma$  and TNF- $\alpha$  of cell culture supernatant from unstimulated and stimulated peripheral mononuclear cells (PBMCs) by HCV specific C200 peptide.