

# **New Guidelines for Platelet Therapy in ICU**

*An Essay*

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In Intensive Care

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بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

قَالُوا سُبْحَانَكَ لَا عِلْمَ لَنَا إِلَّا مَا  
عَلَّمْتَنَا إِنَّكَ أَنْتَ الْعَلِيمُ الْحَكِيمُ

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# List of Contents

<i>Subject</i>	<i>Page No.</i>
List of Abbreviations .....	
List of Tables .....	
List of Figures.....	
Introduction.....	1
Aim of the Work.....	3
Chapter (1): Pathophysiology of Platelet Disorders .....	4
Chapter (2): Diagnostic approach for thrombocytopenia in ICU.....	23
Chapter (3): Therapeutic Aspects for Thrombocytopenic Patients in ICU.....	39
Summary .....	56
References .....	58
Arabic Summary .....	

## List of Abbreviations

<b>Ab</b>	: Antibodies
<b>ADAMTS13</b>	: A Disintegrin And Metalloproteinase with Thrombospondin Motifs
<b>ADP</b>	: Adenosine diphosphate
<b>ADT</b>	: Adult therapeutic dose
<b>APACHE</b>	: Acute Physiology and Chronic Health Evaluation
<b>Aps</b>	: Apheresis platelets
<b>aPTT</b>	: Activated partial thromboplastin time
<b>ATP</b>	: Adenosine triphosphate
<b>ATR</b>	: Allergic transfusion reactions
<b>°C</b>	: The degree Celsius
<b>Ca<sup>2+</sup></b>	: Calcium
<b>CBC</b>	: Complete blood count
<b>CCI</b>	: Corrected count increment
<b>CFU</b>	: Colony-forming unit
<b>CLEC-2</b>	: C-type lectin receptor
<b>DIC</b>	: Disseminated intravascular coagulation
<b>DITP</b>	: Drug-Induced Thrombocytopenia
<b>DNA</b>	: Deoxyribonucleic acid
<b>DTI</b>	: Direct thrombin inhibitors
<b>ELISA</b>	: The enzyme-linked immunosorbent assay
<b>FcR<math>\gamma</math></b>	: Fc receptor $\gamma$ chain
<b>FDP</b>	: Fibrin degradation products
<b>FGF-4</b>	: Fibroblast growth factor 4
<b>fl</b>	: Femtoliter
<b>FNHTR</b>	: Febrile non-hemolytic transfusion reactions
<b>GAS6</b>	: The growth arrest–specific gene 6
<b>GM-CSF</b>	: Granulocyte macrophage-colony, stimulating factor
<b>GP</b>	: Glycoprotein
<b>GPIb<math>\alpha</math></b>	: Glycoprotein Ib $\alpha$

## **List of Abbreviations** *(Cont...)*

<b>GPVI</b>	: Glycoprotein VI
<b>HBV</b>	: Hepatitis B Virus
<b>HCV</b>	: Hepatitis C Virus
<b>HELLP</b>	: Syndrome of haemolysis, elevated liver enzymes and low platelets
<b>HIT</b>	: Heparin-induced thrombocytopenia
<b>HIV</b>	: Human immunodeficiency virus
<b>HLA</b>	: Human leukocyte antigen
<b>HNA</b>	: Human Neutrophil Antigens
<b>HPA</b>	: Human platelet antigen
<b>HUS</b>	: Haemolytic uraemic syndrome
<b>ICU</b>	: Intensive care unit
<b>IgA</b>	: Immunoglobulin A
<b>IL</b>	: Interleukin
<b>INR</b>	: International normalized ratio
<b>ISTH</b>	: The International Society on Thrombosis and Haemostasis
<b>ITP</b>	: Idiopathic thrombocytopenic purpura
<b>IVIG</b>	: Intravenous immunoglobulin
<b>L</b>	: Liter
<b>LDH</b>	: Lactate dehydrogenase
<b>LDH</b>	: Lactate dehydrogenase
<b>MAHA</b>	: Microangiopathic hemolytic anemia
<b>MODS</b>	: Multiple Organ Dysfunction Scores
<b>NAT</b>	: Nucleic acid amplification techniques
<b>NO</b>	: Nitric oxide
<b>OCS</b>	: Open canalicular system
<b>PAR</b>	: Protease-activated receptors
<b>PCs</b>	: Platelet concentrates
<b>PCV</b>	: Packed cell volume
<b>PDGF</b>	: Platelet-derived growth factor
<b>PG</b>	: Prostaglandin

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## **List of Abbreviations** *(Cont...)*

<b>PLC<math>\gamma</math>2</b>	: Phospholipase C $\gamma$ 2
<b>PMPs</b>	: Platelet-derived Microparticles
<b>PTP</b>	: Posttransfusion purpura
<b>RNA</b>	: Ribonucleic acid
<b>SAPS</b>	: Simplified Acute Physiology Scores
<b>SDF</b>	: stromal cell derived factor
<b>SH2</b>	: Src homology 2
<b>TM</b>	: Thrombotic microangiopathy
<b>Tpo</b>	: Thrombopoietin
<b>TRALI</b>	: Transfusion related acute lung injury
<b>TTP</b>	: Thrombotic thrombocytopenic purpura
<b>TXA2</b>	: Thromboxane A2
<b>VKAs</b>	: Vitamin K antagonists
<b>VWF</b>	: von Willebrand factor

## List of Tables

<i>Table No.</i>	<i>Title</i>	<i>Page No.</i>
<b>Table (1):</b>	Categories of Thrombocytopenia.....	24
<b>Table (2):</b>	Diagnostic score for the diagnosis of overt DIC.....	26
<b>Table (3):</b>	Comparison of the features of TTP and hemolytic uremic syndrome (HUS).....	27
<b>Table (4):</b>	Comparison of DIC, TTP-HUS, and HELLP Syndromes.....	28
<b>Table (5):</b>	The 4Ts pretest probability of heparin- induced thrombocytopenia.....	31
<b>Table (6):</b>	Thrombocytopenia (TP) in ICU patients in relation to the time course of platelet count.....	38



## List of Figures

<i>Figure No.</i>	<i>Title</i>	<i>Page No.</i>
<b>Figure (1):</b>	Platelets structure .....	9
<b>Figure (2):</b>	Schematic overview of the main platelet receptors and effectors involved in platelet signaling. ....	13
<b>Figure (3):</b>	Schematic representation of platelet receptors and agonists that mediate platelet activation through tyrosine kinase signaling .....	14
<b>Figure (4):</b>	Platelet count courses in ICU patients. ....	21

# Introduction

Platelets are the second most abundant cells, after red blood cells, in the blood circulation. They are produced from their precursor megakaryocytes in the bone marrow. The major physiological role of platelets is to accumulate at sites of damaged blood vessel endothelium and initiate the blood clotting process. Platelet adhesion, activation, and subsequent aggregation at sites of vascular injury are critical to the normal arrest of bleeding (*Li et al., 2012*).

Thrombocytopenia is a very frequent disorder in the intensive care unit. Many etiologies should be searched, and therapeutic approaches differ according to these different causes (*Van der Linden et al., 2012*).

Thrombocytopenia presents a multifaceted pathogenic mechanism comprising hemodilution, increased platelet consumption, increased platelet destruction, decreased platelet production, increased platelet sequestration, and laboratory artifacts (*MarcoSchulke et al., 2012*).

The many comorbidities in the severely ill patient make thrombocytopenia very common (40%) in intensive care unit patients. The risk of bleeding is high with severe thrombocytopenia and is enhanced in intensive care patients with mild or moderately low platelet counts when additional factors are present that interfere with normal hemostatic mechanisms (eg,

platelet function defects, hyperfibrinolysis, invasive procedures, or catheters). Even if not associated with bleeding, low platelet counts often influence patient management and may prompt physicians to withhold or delay necessary invasive interventions, reduce the intensity of anticoagulation, order prophylactic platelet transfusion, or change anticoagulants due to fear of heparin-induced thrombocytopenia. One approach to identify potential causes of thrombocytopenia that require specific interventions is to consider the dynamics of platelet count changes (*Andreas et al., 2010*).

Platelet transfusions have greatly reduced the incidence of major haemorrhagic complications associated with the management of haematological and oncological disorders. The use of platelet transfusion continues to increase and platelets are an essential component in the management of selected patients with thrombocytopaenia. They need to be used judiciously as they are not free from risk. Platelet transfusions are not indicated in all causes of thrombocytopenia and maybe contraindicated in certain conditions. Hence, the cause of thrombocytopenia should be established before making a decision about platelet transfusion (*Paul et al., 2012*).

## **Aim of the Work**

To focus on different causes of platelet disorders in critically ill patients and decide whether or not to support the platelet count by platelet transfusion.

## Chapter (1): **Pathophysiology of Platelet Disorders**

### **Pathophysiology of platelet disorders**

#### **Physiology of platelets**

##### **1. Origin:**

Platelets originate from the cytoplasm of bone marrow megakaryocytes. Platelets lack genomic DNA but contain megakaryocyte derived messenger RNA (mRNA) and the translational machinery needed for protein synthesis (*Sharathkumar et al., 2008*).

Platelet production, or thrombopoiesis, is a complex process that can be schematically represented as consisting of four main steps. The first step is the production of the thrombopoietic stimulus, which drives the generation of megakaryocytes and ultimately platelets. Although a number of cytokines (ie, IL-3 [interleukin 3], IL-6 [interleukin 6], IL-11 [interleukin 11], GM-CSF [granulocyte macrophage-colony stimulating factor]) and chemokines (ie, SDF [stromal cell derived factor] and FGF-4 [fibroblast growth factor 4]) contribute to this process, thrombopoietin (Tpo) is widely recognized as the most potent known stimulator of platelet production (*Kaushansky et al., 2008*).

Platelet production is tightly regulated by the hormone thrombopoietin (TPO), which is secreted by the liver at a constant rate (constitutive secretion) and acts on hematopoietic progenitor cells, bone marrow megakaryocytes, and platelets by binding to its receptor c-Mpl. Binding of TPO to c-Mpl leads to the differentiation of hematopoietic progenitor cells down the megakaryocyte lineage, maturation of megakaryocytes, and ultimately increased platelet production (*Arnold et al., 2009*).

Once TPO is bound to c-Mpl, it is internalized, degraded, and removed from the circulation. Levels of free TPO are therefore controlled by the number of circulating platelets and the megakaryocyte mass, when platelet levels and megakaryocyte numbers are low, free TPO levels are high and more platelets are produced; when platelet levels are high, circulating TPO levels are reduced (*Arnold et al., 2011*).

## **2. Structure:**

Circulating platelets are discoid in shape, with dimensions of approximately 2.0–4.0 by 0.5  $\mu\text{m}$ , and a mean volume of 7–11 fl. Their shape and small size enables the platelets to be pushed to the edge of vessels, placing them in the optimum location to constantly survey the integrity of the vasculature. Platelets circulate in a concentration of 150,000–450,000 cells/mL. Of the total body platelets, about 70% stay in circulation while the remaining 30% are continually but transiently sequestered in the spleen. Platelets remain in circulation for an average of 10 days. Most platelets are

removed from the circulation by the spleen and liver after senescence, but a constant small fraction is continually removed through involvement in maintenance of vascular integrity (*Sharathkumar et al., 2008*).

On peripheral blood smears stained with Wright-Giemsa stain, platelets appear as small, granular staining cells with a rough membrane, and are normally present as 3-10 platelets per high power oil-immersion field. Despite their simple appearance on the peripheral blood smear, platelets have a complex structure (Figure 1) Platelet internal structure has been divided into four zones:

- Peripheral zone
- Sol-gel zone
- Organelle zone
- Membrane zone (*Sharathkumar et al., 2008*).

The **peripheral zone** includes the outer membranes and closely associated structures. The platelet has a surface connected system of channels called the open canalicular system (OCS). The walls of the OCS are included in this zone. The OCS provides access to the interior of the platelet to plasma substances, and an outlet channel for platelet products. The release of platelet products through the OCS after platelet activation is called “the release reaction” (*Sharathkumar et al., 2008*).

The membranes of the platelet are rich in platelet receptors, which determine its specific cellular identity. These receptors are constitutively expressed on the platelets and

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