Screening, Characterization and Biochemical Studies for Novel Polyhydroxybutyrate (PHB) Producing and/or degrading bacteria.

Thesis submitted for Partial Fulfillment of Master degree of Science (Biochemistry)

Submitted by

Eman Mohamed Mostafa Fath El-Din

B. Sc. Faculty of Science Cairo University

Supervised by

Dr. Mervat Elsayed Mohammed

Professor assistant of Biochemistry Faculty of Science Cairo University

Dr. Moataz fouad Abd El-Ahany

Development of gastric vaccines Holding campany of bioproduct and Vaccines

Faculty of Science
Cairo University
2010

Screening, Characterization and Biochemical Studies for Novel Polyhydroxybutyrate (PHB) Producing and/or degrading bacteria.

Thesis Submitted for Partial Fulfillment of Master Degree of Science (Biochemistry)

Submitted by

Eman Mohamed Mostafa Fath El-Din

B. Sc. Faculty of Science Cairo University

Supervised by

Dr. Mervat Elsayed Mohammed

Professor assistant of Biochemistry Faculty of Science Cairo University

Dr. Moataz fouad Abd El-Ahany

Development of gastric vaccines Holding campany of bioproduct and Vaccines

Faculty of Science
Cairo University
2010

APPROVAL SHEET FOR SUBMISSION

Title of M.Sc. the Thesis: Screening, Characterization and Biochemical Studies for Novel Polyhydroxybutyrate (PHB) Producing and/or degrading bacteria.

Name of Candidate: Eman Mohamed Mostafa Fath El-Din

This thesis has been approved for submission by the supervisors:

1. Dr. Mervat Elsayed Mohammed

Professor Assistant of Biochemistry, Faculty of Science, Cairo University.

Signature:

2. Dr. Moataz fouad Abd El-Ahany

Development of gastric vaccines, Egyptian Organization For Biological Product and Vaccines

Signature:

Prof. Dr. Mohamed M. Shakry

Chairman of Chemistry Department Faculty of Science-Cairo University

Acknowledgement

This work would not have been possible without the help of many people.

First, I wish to express my deepest gratitude and my great respect to my **Prof. Dr. Sayed Kamel Goda**, Professor of Biochemistry and Genetic Engineering, Faculty of Science, Cairo University.

I would like to express my deep gratitude and appreciation to him for his unlimited assistance, continuous support and meticulous supervision through the work. I am also grateful for his tremendous help in solving problems in this work.

I wish to express my profound gratitude to my supervisor **Dr. Mervat Elsayed**, Professor Assistant of Biochemistry, Faculty of Science, Cairo University, who showed me the rest of the way and supported me to accomplish this work successfully.

I wish to express my profound gratitude to my supervisor **Dr. Moataz fouad Abd El-Ahany**, Development of gastric vaccines, Holding campany of bioproduct and Vaccines, who helped me in the most important parts of this work and supported me to accomplish this work successfully.

All my thanks to my family they are always praying for me to finish this work successfully

Finally, I would like to express my thanks to all my friends in Biochemistry laboratory, Faculty of Science, Cairo University and to my friends in Holding Company of bioproduct and Vaccines.

ABSTRACT

Name : Eman Mohamed Mostafa Fath El-din

Title of M.Sc. Thesis:

Screening, Characterization and Biochemical Studies for Novel Polyhydroxybutyrate

(PHB) Producing and/or degrading bacteria.

Degree : Master degree in Science

In this study five Polyhydroxybutyrate (PHB)-degrading bacterial strains were isolated from soil samples collected from different agricultural sites in Egypt. These strains designated as MEM 1, MEM 2, MEM 3, MEM 4 and MEM 5. They were able to use Polyhydroxybutyrate [1 g/l] as a sole carbon source in the growth media. MEM1 shows the highest activity of degradation of the PHB. It was therefore chosen for further studies. MEM1 Isolated strain was preliminarily identified by API ® 20 NE as *Alcaligenes faecalis*. PCR amplification and partial sequence analysis of 16S rRNA genes identified MEM1 isolate as *Alcaligenes faecalis* strain ST1. The degradation potential of strains was examined and detected using Optical density, measured by a spectrophotometer. The level of succinic acid (one of PHB degradation products) in the culture broth was utilized as indirect indicator for the ability of growing bacteria to assimilate polymer. Attempts were made to amplify the gene involved in PHB degradation (depolymerase gene) by PCR amplifications. PCR product with the right size was formed. The DNA sequencing and alignment analysis indicated that the fragment very unlikely to contain the depolymerase gene, Further study is being done to clone the new depolymerase from the isolated strain.

Key Words:

Polyhydroxybutyrate(PHB), degradation, Alcaligenes faecalis and depolymerase.

signature:

Supervisors: 1- Dr. Mervat Elsayed Mohammed

2- Dr. Moataz Fouad A.Ghany

Prof. Dr. Mohamed M. Shakry

Chairman of Chemistry Department Faculty of Science-Cairo University

CONTENTS

		<u>Page</u>
Table of cor	ntents	i
List of Table	es	V
List of Figu	res	vi
List of Abbi	reviations	viii
Chapter 1	l: Introduction and Aim of the work	
1.1.	Polymers	1
1.1.1	Natural polymeric materials	1
1.1.2.	Synthetic polymer and industrial application	1
1.1.2 .1	Economic Importance	1
1.2.	Disadvantage of the synthetic polymers (Environmental	2
	pollution)	
1.3.	Methods of Solid Waste Disposal.	2
1.3.1.	Landfilling And Incineration.	2
1.3.1.1.	Landfilling.	2
1.3.1.2.	Incineration	3
1.3.2.	Biosynthetic and Biodegradable Polymers	4
1.3.2.1.	Biosynthetic polymer	4
1.3.2.2.	Biodegradation of Polymers	5
1.4.	Polyhydroxyalkanoates	6
1.4.1.	Properties of PHAs.	9
1.4.2.	PHB (Poly-3-hydroxybutyrate)	10
1.4.2.1.	PHB Properties.	10
1.4.2.1.1.	Chemical Structure of PHB.	11
1.4.2.1.2.	Properties of PHB.	11
1.4.2.1.3.	Biodegradation of PHB	12

1.4.2.2.	PHB depolymerase
1.4.2.3.	Acceleration of biodegradation (Bioaugmentation)
1.4.2.4.	Bioremediation.
1.4.2.5.	Methods of Production of PHB
1.4.2.6.	Application of PHB
1.4.2.7.	PHB and PP
1.5.	Aim of work
Chapter	2: Materials and Methods
2.1.	Materials and Equipments
2.1.1.	Chemicals
2.1.2.	Molecular biology standards and kits
2.1.3.	Equipments
2.1.4.	Bacterial strains
2.1.5.	Media
2.1.6.	DNA solutions
2.2.	METHODS
2.2.1.	Collection of Soil samples.
2.2.2.	Screening for Isolation of Poly-β-Hydroxybutyrate (PHB)-
	degrading bacterial Strain
2.2.2.1.	Enrichment of organisms which can degrade Poly-β-
	Hydroxybutyrate (PHB)
2.2.2.2.	The ability of isolated strains to grow on PHB as the sole
	carbon source.
2.2.2.3.	Isolation of Poly-β-Hydroxybutyrate (PHB) degradating
	microorganism
2.2.2.4.	Bacterial strains
2.2.3.	Culture properties

2.2.3.1. Optical density of isolated strains	
2.2.3.2. Identification of one of the PHB degra	adation products
(succinic acid)	
2.2.4. Characterization and identification of bacterial	l isolates
2.2.4.1. Morphological characterization of bacterial iso	olates
2.2.4.2. Identification of bacteria strain by Biochemica	al test(API ® 20
NE)	
2.2.4.3. Molecular Identification of bacterial isolate	
2.2.4.3.1. Rapid preparation of genomic DNA	from bacterial
colonies	
2.2.4.3.2. Polymerase chain reaction (PCR) amplification	on of 16S rRNA
genes of MEM 1 isolate	
2.2.4.3.3. Agarose Gel Electrophoresis	
2.2.4.3.4. Purification and partial sequence analysis	of the PCR
products	
2.2.4.3.5. Partial sequence analysis of the PCR products	
2.2.5. Primer design for PCR amplification of PHB of	degrading gene
; the gene responsible for PHB degradation	
2.2.5.1. PCR Amplification of partial sequence An	nalysis of PHB
degrading gene (Depolymerase gene)	
Chapter 3: Results	
3.1. Isolation of Polyhydroxybutyrate (PHB) deg	rading bacterial
strains	
3.2. Culture Properties	
3.2.1. Optical density of isolated strains	
3.2.2. Identification of one of the PHB degrad	
(succinic acid)	
3.3 Characterization and identification of isolated b	pacteria
3.3.1 Morphological characterization	

	:	<u>Page</u>
3.3.2	Biochemical Characterization.	44
3.3.3	Molecular Identification of the bacterial isolates	45
3.3.3.1.	PCR amplification of 16S rRNA gene of MEM 1 isolate	45
3.3.3.3.	Partial sequence analysis of 16S rRNA gene	47
3.4.	Primer Design and PCR amplification of Poly-β-	52
	Hydroxybutyrate (PHB) degrading gene : the gene responsible for PHB degradation	
3.4.1.	Agarose Gel electrophoreses of Amplified PCR product: Poly-	55
	β-Hydroxybutyrate (PHB) degrading gene	
3.4.2.	Partial sequence analysis of amplified Poly-β-Hydroxybutyrate	56
	(PHB) degrading gene	
Chapte	er 4: Discussion	59
English	<u>Summary</u>	65
Referen	<u>ices</u>	67
Arabic	summary	

List of Tables

		<u>Page</u>
Table 3.1	Optical density of MEM 1 bacterial isolate in presence and	37
	absence of PHB polymer	
Table 3.2	Optical density of MEM 2 bacterial isolate in presence and	38
	absence of PHB polymer.	
Table 3.3	Optical density of MEM 4 bacterial isolate in presence and	39
	absence of PHB polymer	
Table 3.4	Optical density of MEM 5 bacterial isolate in presence and	40
	absence of PHB polymer	
Table 3.5	Morphological characteristics of bacterial isolates	43
Table 3.6	NCBI Blast sequence alignment of MEM 1 isolate	50
Table 3.7	Characteristics of selected primers used in PCR	52
	amplification of partial nucleotide sequence of gene	

List of Figures

		<u> </u>
Fig. 1.1	Chemical structures of poly(3-hydroxybutyrate) (PHB), poly(e-caprolactone) (PCL). poly(butylene succinate) (PBS) and poly(L-	
	lactide) (L-PLA)	
Fig. 1.2	Biosynthesis of PHA from fatty acid (-oxidation in R. rubrum	
Fig.1.3	Biosynthesis of PHA from "denovo fatty acid synthesis" in pseudomonas group-II	
Fig. 1.4	Chemical structures of P3HB.	
Fig. 1.5	The pathway of PHB degradation with enzymes	
Fig. 3.1	Optical density of MEM 1 bacterial isolate in presence and absence of PHB polymer.	
Fig. 3.2	Optical density was measured daily in culture contain MEM 2 bacteria	
Fig. 3.3	Optical density was measured daily in culture contain MEM 4 bacteria	
Fig 3.4	Optical density was measured daily in culture contain MEM 5 bacteria	
Fig. 3.5	Indirect identification of PHB degradation using MEM 1, MEM 2 and MEM 5 isolates	
Fig. 3.6	API 20 NE Characterization of MEM 1 isolate	
Fig.3.7	Extraction of bacterial genomic DNA	
Fig.3.8	Amplification of 16S rRNA gene	
Fig.3.9	Sequence chromatogram of 16S rRNA gene	
Fig.3.10	Partial Sequence of Amplified 16s rRNA gene	
Fig.3.11	Sequence alignment of amplified 16S rRNA gene of MEM 1 isolate with that of <i>Alcaligenes faecalis</i> strain	
Fig.3.12	Binary alignment of nucleotide sequences of depolymerase gene from Alcaligenes faecalis and Burkholderia pseudomallei	
	strains	

		<u>Page</u>
Fig.3.13	Amplification of PHB degrading gene	55
Fig.3.14	Sequence chromatogram of amplified fragment of degrading gene	57
	from MEM 1 by using DE-f primer.	
Fig.3.15	Partial nucleotide sequence of PCR product from MEM 1 isolated	58
	bacteria	
Fig 3.16	Deduced amino acid sequence isolated from Alcaligenes faecalis	58
	strain ST1	

List of Abbreviations

° C : Degree centigrade

Abs : Absorbance
bp : Base pair
Da : Daltons

DE : depolmerase

DNA : Deoxyribonucleic acid

EBI : European bioinformatics institute

EDTA : Ethylene Diamin Tetra Acetic Acid

F : Forward
Fig. : Figure
g/gm : Gram

IUPAC : International Union of Pure and Applied Chemistry

J. : JournalKb : KilobaseKg : Kilogram

Kp : Kilo base pair

L : Liter

MPa : millipascal

mcl-PHA : medium-side-chain PHA

min : minute

Mix : mixture

ml : milliliter

MSM : minimal salt media

MT : Metric Ton

NCBI : national center for biotechnology information

nm : nanometer
no : Number

O.D. : Optical density

PBS : poly(butylene succinate)

PCL : Polycaprolactone

PES : poly(ethylene succinate)

PCR : Polymerase Chain Reaction

PHA : Polyhydroxyalkanoate

PHAs : Polyhydroxyalkanoates

PHB : polyhydroxybutyrate

P(HB-HV) : poly(hydroxybutyrate/hydroxyvalerate)

PHV : hydroxyvalerate L-PLA : poly(L-lactide)

PP : Polypropylene

ppm : Part per million

R, Rev. : Reverse

Rpm : revolutions per minute

rRNA : Ribosomal ribonucleic acid

Rs : reduce, reuse and recycle

scl-PHA : short-side-chain PHA

SDS : Sodium Dodecylsulphate

SDS-PAGE : Sodium Dodecylsulphate-Polyacrylamide Gel

Sec : Second sp. : Species

TBE : Tris Borate buffer

Tg : glass transition temperature

Tm : Melting temperature

Tris : Trishydroxymethylaminomethane

TSA : Trypticase soy agar media

UPVC : Unplasticized polyvinyl chloride

UV : ultra violet

w/v : Weight to volume

wt : weight

μl : micro liter

