GRAM NEGATIVE INFECTIONS IN THE INTENSIVE CARE UNIT

Essay

Submitted for Partial Fulfillment of Master Degree in Intensive Care

By

Rashad Mohamed El Attar

(M.B, B.CH.)

Under Supervision of

Prof. Dr. Mervat Mohamed Marzook

Professor of anesthesia and intensive care Faculty of Medicine - Ain Shams University

Dr. Fady Adib Abdel Malek

Lecturer of anesthesia and intensive care Faculty of Medicine - Ain Shams University

Dr. Hany Ahmed Abdel Kader

Lecturer of anesthesia and intensive care Faculty of Medicine - Ain Shams University

Faculty of Medicine
Ain-Shams University

2012

LIST OF CONTENTS

| Acknowledgement | |
|---|----|
| List of Contents | |
| List of Tables | |
| List of Figures | |
| List of Abbreviations | |
| Introduction and Aim of the Work | 1 |
| Review of Literature | |
| Epidemiology of gram negative sepsis | 3 |
| Pathophysiology of gram negative sepsis | 8 |
| Diagnosis of gram negative sepsis | 32 |
| Treatment of gram negative sepsis | 37 |
| Preventions of gram negative sepsis | 72 |
| Summary | 78 |
| References. | 81 |
| Arabic summary | |

LIST OF TABLES

| Table no. | Table content | page |
|-----------|--|------|
| 1 | Biologic effects of Proinflammatory cytokines such as TNF and IL-1 | 10 |
| 2 | Laboratory Indicator Of Sepsis | 34 |
| 3 | Vasopressors used in the treatment of septic shock | 45 |
| 4 | Methods of source control for common ICU infections | 48 |

LIST OF FIGURES

| Figure no. | Title | page |
|------------|--|------|
| | | |
| 1 | Sepsis and SIRS effects. | 12 |
| | Pathways of complement system | |
| 2 | activations | 14 |
| | Comparison of the Gram positive and | |
| 3 | Gram negative bacterial cell walls | 18 |
| | Schematic representation of the | |
| 4 | structure of lipopolysaccharide | 19 |
| | Schematic representation of the inner | |
| 5 | and outer bacterial membrane of gram negative bacteria | 20 |
| | | |

List of Abbreviations

LIST OF ABBREVIATIONS

| ALI | acute lung injury |
|----------------------------|---|
| ALT | alanine aminotransferase |
| APACHE | Acute physiology and chronic health evaluation |
| APC | Activated protein C |
| AQP4 | aquaporin 4 |
| ARDS | acute respiratory distress syndrome |
| AST | aspartate transaminase |
| BBB | Blood Brain Barrier |
| BP | Blood pressure. |
| C1 | complement fragment 1 |
| C5a | complement fragment 5a |
| C5aR | complement fragment 5a receptor |
| C5b-9complexes | terminal or late acting complement proteins |
| CCR2 | chemokine receptor 2 |
| CDC | Centers for Disease Control and Prevention |
| CD14, MD2, toll-like | cell surface receptors |
| receptors 2 and 4, and Fc- | |
| gamma receptors II and III | |
| cGMP | cyclic guanosine monophosphate |
| cNOS/ NOS3 | Constitutive nitric oxide synthase |
| CVP | central venous pressure |
| CVVH | Continuous venovenous hemofiltration |
| DIC | disseminated intravascular coagulation |
| ED | Emergency Department |
| EDTA | Disodium edetate |
| eNOS | endothelial nitric oxide synthase |
| ESBL | Extended spectrum beta-lactamases |
| ESICM | European Society of Intensive Care Medicine |
| ESKAPE | Enterococcus faecium, Staphylococcus aureus, |
| | Klebsiella pneumoniae, Acinetobacter baumannii, |
| | Pseudomonas aeruginosa and Enterobacter spp |
| G-CSF | granulocyte colony-stimulating factor |

List of Abbreviations

| GI | gastrointestinal |
|------------------|--|
| HAI | Hospital acquired infections |
| HCW | health care worker |
| ICAM-1 | intercellular adhesion molecule-1 |
| ICU's | intensive care units |
| IIT | intensive insulin therapy |
| IL-1 | Interleukin-1. |
| IM | inner membrane |
| iNOS/ NOS2 | Inducible nitric oxide synthase |
| IPP | inspiratory plateau pressure |
| ISF | International Sepsis Forum |
| LBP | Lipid-binding protein |
| LMWH | low molecular weight heparin |
| LPS | lipopolysaccharide |
| MAC | membrane attack complex |
| MAP | mean arterial pressure |
| MDR | multi-drug resistant |
| MODS | Multiple organ dysfunctions |
| MOF | multiple organ failure |
| MRSA | methicillin-resistant Staphylococcus aureus |
| MyD88 | myeloid differentiation factor 88 |
| nAChR | nicotinic acetylcholine receptor |
| NF- b | Nuclear factor- b. |
| NNIS | National Nosocomial Infections Surveillance. |
| nNOS/ NOS1 | Neuronal nitric oxide synthase |
| NO | Nitric oxide. |
| NOD | nucleotide-oligomerization domain |
| NOS | nitric oxide synthase |
| OM | outer membrane |
| OMPs | outer membrane proteins |
| OMVs | Outer membrane vesicles |
| PAMPs | pathogen-associated molecular patterns |
| PCO ₂ | Partial pressure of CO ₂ |
| PECAM | platelet endothelial cell adhesion molecule |
| PEEP | Positive end expiratory pressure |

List of Abbreviations

| PMNs | Polymorphonuclear leukocytes |
|--|--|
| Ppao | pulmonary artery occlusion pressure |
| PRRs | Pattern recognition receptors |
| PS | Polysaccharide |
| PT | prothrombin time |
| PTT | partial thromboplastin time |
| RCTs | Randomized controlled trials. |
| rhAPC | recombinant human activated protein C |
| rhMFG-E8 | Recombinant Human Milk Fat Globule Epidermal |
| | Growth Factor 8 |
| RIG-I | retinoic-acid-inducible gene I. |
| ROS | reactive oxygen species |
| SARS | severe acute respiratory syndrome |
| SBT | Spontaneous breathing trials |
| SCCM | Society of Critical Care Medicine |
| ScvO ₂ and SvO ₂ | Central or mixed venous oxyhemoglobin saturation |
| SIRS | systemic inflammatory response syndrome |
| SNP | single nucleotide polymorphism |
| SSC | Surviving Sepsis Campaign |
| SVR | systemic vascular resistance |
| TIR | Toll-IL-1 resistance |
| TLRs | Toll-like receptors. |
| TNF | tumor necrosis factor alpha |
| TRIF | TIR-domain-containing adaptor molecule 1 |
| UFH | unfractionated heparin |
| VCAM-1 | vascular cell adhesion molecule-1 |
| VRE | vancomycin-resistant enterococci |
| WHO | World Health Organization |



Thanks first and last to **ALLAH** as we owe Him for His great care, support and guidance in every step in our life.

I would like to express my deep gratefulness to Professor Dr. Mervat Mohamed Marzook, Professor of anesthesia and intensive care Faculty of Medicine - Ain Shams University, for her concern, her guidance and continuos encouragement. I consider myself very fortunate to work under her supervision.

I am grateful to Dr. Fady Adib Abdel Malek Lecturer of anesthesia and intensive care Faculty of Medicine - Ain Shams University for his generous and continous help and valuable suggestions as well as his desire to achieve perfection. His great help was very impressive and her meticulous reading and enormous assistance should be appreciated.

I am also grateful to Dr. Hany Ahmed Abdel Kader Lecturer of anesthesia and intensive care Faculty of Medicine - Ain Shams University for his generous and continous help and valuable suggestions as well as his desire to achieve perfection. His great help was very impressive and her meticulous reading and enormous assistance should be appreciated.

I must also express my thanks to all my family and my colleagues who helped me in this essay.

Introduction

Infections caused by gram negative bacilli typically occur in the lungs, the urinary tract, at surgical sites and in the blood stream. They are considered a significant cause of morbidity and mortality. The gram negative bacilli most commonly responsible for infection in humans are Enterobacter *species*, Klebsiella pneumoniae, Pseudomonas aeruginosa, Escherichia coli, Acinetobacter species, *and* Serratia marcescens. Depending on the site of infection and the patient's comorbid conditions, mortality related to infections caused by gram-negative bacilli ranges from 20 % to 60 % (Sarah et al., 2011).

In prevalence study evaluating the extent and patterns of infection among more than 13 000 patients in ICU's around the world, 51% of patients had documented infection and 71% were receiving antimicrobial agents; gram-negative infections accounted for 63% of infections (Vincent et al., 2009).

Infections caused by gram-negative bacteria have features that are of particular concern. These organisms are highly efficient at up-regulating or acquiring genes that code for mechanisms of antibiotic drug resistance, especially in the presence of antibiotic selection pressure (Boucher et al., 2009).

The aim of the study

The aim of this essay is to cast light on the problem of gram negative infections in the intensive care unit with emphasis on their prevalence, pathogenesis, diagnosis, treatment and available strategies for their prevention.

Epidemiology of gram negative sepsis

bacilli were the gram negative to the 1980's, predominant organisms associated with nosocomial blood stream infections in the United States. Since then, gram-positive aerobes (e.g., coagulase negative staphylococci, staphylococcus aureus, and enterococcus) and Candida species have increased in relative importance. In 1975, hospitals that participated in the National Nosocomial Infections Surveillance (NNIS) System reported that gram negative bacilli were responsible for 55 % of bacteremia hospital wide. By 2003, NNIS hospitals reported that gram negative bacilli were responsible for 24 % of nosocomial bacteremia in intensive care units (ICUs) (Gaynes and Edwards, 2005).

However, a surveillance study of hospital wide primary health care associated blood stream infections in a large tertiary care hospital in the United States found a significant increase in the rate of gram negative bacillary blood stream infections in 2003 which may represent either a reemergence of gram-negative infections over the past five years or changes in patient demographics in that institution (Albrecht et al., 2006).

The proportion of gram negative organisms identified in blood stream infections increased significantly from 15.9 % in 1999 to 24.1 % in 2003. No specific gram negative species contributed disproportionately to the increase and with few exceptions there were no significant increases in antimicrobial resistance to explain this trend. This study excluded bloodstream infections that were community-acquired or were secondary to infection at another site. In Latin America, and some areas of Europe and

the Far East, the proportion of bacteremia caused by gramnegative bacilli is greater than that identified in the United States this geographic difference in etiology of bacteremia is illustrated by the following studies: (**Biedenbach et al., 2004**).

A prospective study of bloodstream infections in hematologic malignancy patients from Brazil hospitalized between January 2000 and June 2001 identified gram-negative bacteria in 81 of 133 (61 %) episodes (**velasco et al., 2003**). In contrast, studies from North America have generally shown a predominance of gram-positive bacteria (**Gaynes and Edwards, 2005**).

A prospective multicenter study in Italy conducted between 1999 and 2000 found 46 % of bacteremia was due to gram negative bacilli in this study; gram negative bacilli were the etiologic agent in 42 % of hospital-acquired bacteremia and 55 % of community-acquired bacteremia (luzzaro et al., 2002).

A retrospective review of 238 episodes of bacteremia in patients 65 years of age in the United States identified gramnegative bacilli as the etiologic agent in 36 % of cases; 81 % of patients were admitted with bacteremia from home (greenberg et al .,2005). The rate of gram-negative bacteremia in patient's 65 years is even higher in nursing home residents as illustrated by a retrospective review of 169 episodes of bacteremia identified between January 1997 and April 2000 Of the 132 episodes of monomicrobial bacteremia, 78 (59 %) were caused by gram-negative organisms. Thus, even within North America, there are subpopulations of patients in whom gramnegative bacteremia occurs more frequently than does gramnegative bacteremia occurs more frequently than does gramnegative

positive bacteremia (mylotte et al., 2002).

Risk factors

Most hospitalized patients with gram negative bacteremia have at least one comorbid condition a study of 326 episodes of gram negative bacteremia at two universities-affiliated medical centers identified comorbid conditions in 315 (97 %) Conditions identified in this and other studies included:

- Hematopoietic stem cell transplant
- Liver failure
- Serum albumin <3 mg/dL
- Solid organ transplant
- Diabetes
- Pulmonary disease
- Chronic hemodialysis
- HIV infection
- Hematologic malignancy
- Treatment with glucocorticoids
- Elderly

(Thomsen et al., 2005).

In addition to these risk factors, combat injured military personnel and patients injured during natural disasters involving trauma in water are also at increased risk for infections caused by gram-negative bacilli (Kallman et al., 2006).

Source of infection

The source of infection differs depending on the patient population studied as illustrated in the following studies:

In the retrospective review of nursing home residents noted above, the following were identified as the most common sources of infection in the cases of gram-negative bacteremia:

Gram negative infections in ICU

| • | Urinary tract | 81 % |
|---|------------------------|------|
| • | Gastrointestinal tract | 9 % |
| • | Respiratory tract | 4 % |
| • | others | 6 % |

In a retrospective review of gram-negative bacteremia in the elderly (eg, >70 years), the most common sources of infection in community-acquired infection were:

| • | Urinary tract | 50 % |
|---|------------------------|------|
| • | Gastrointestinal tract | 27 % |
| • | Respiratory tract | 10 % |
| • | Skin and soft tissue | 8 % |
| • | Others | 5 % |

A similar distribution was noted for hospital-acquired infection (Mylotte et al., 2002).

The source of infection differs in patients admitted to the ICU; a retrospective study found the following most common sources in this patient population:

| • | Respiratory tract | 49 % |
|---|-------------------------|------|
| • | Central venous catheter | 16 % |
| • | Gastrointestinal tract | 4 % |
| • | Urinary tract | 2 % |
| • | others | 29 % |

Identifying the most likely source of infection impacts on empiric antibiotic treatment because the most likely microbiologic agent responsible may differ by source of infection (Sligl et al., 2006).

Causative organisms

Data from the National Nosocomial Infections Surveillance (NNIS) System stated that the most common gram negative species isolated during gram-negative bacteremia included:

| Enterobacter species | 18.5 % |
|---|--------|
| • Klebsiella pneumoniae | 17.8 % |
| • Pseudomonas aeruginosa | 14.4 % |
| • Escherichia coli | 13.6 % |
| Acinetobacter species | 10.1 % |
| Serratia marcescens | 9.7 % |

(Gaynes and Edwards, 2005).

The order of most common gram negative bacilli in blood stream infections in community acquired cases differs from those responsible for hospital acquired cases. This is illustrated by the Italian study noted above which identified the following list of the most common gram negative organisms isolated during community acquired gram negative bacteremia

| (Luzzaro et al., 2002). | | |
|-------------------------|-------|--|
| • Enterobacter species | 3.7 % | |
| • Proteus mirabilis | 4.2 % | |
| • K. pneumonia | 5.4 % | |
| • P. aeruginosa | 7.9 % | |
| • E. coli | 76 % | |