Haemostatic Failure in Critically Ill Patients

An Essay

Submitted For Partial Fulfillment of Master Degree in Intensive Care

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List of Abbreviations

AABB : American Association of Blood BanksACoTS : Acute Coagulopathy of Trauma-Shock

ACS : Acute coronary syndrome **ADP** : Adenosine diphosphate

APLA : Antiphospholipid antibody

APTT : Activated partial thromboplastin time

AT3 : Antithrombin III

ATP : Adenosine triphosphate

BT : Bleeding time

CBC : Complete blood count

CCI : Corrected count increment

CFT : Clot firmness time

DDAVP : 1-deamino-8-D-arginine VasopressinDIC : Disseminated intravascular coagulation

DITP : Drug-induced immune thrombocytopenic purpura

EACA : e-aminocaproic acid
 ELT : Euglobulin lysis time
 FFP : Fresh frozen plasma
 GAGs : Glycosaminoglycans
 GVHD : Graft-vs-host disease

HELLP: Syndrome of haemolytic anaemia, elevated

liver enzymes and low platelets

HIPA : Heparin-induced platelet activationHIT : Heparin-induced thrombocytopenia

ICU : Intensive care unit

INR : International normalized ratio

List of Abbreviations

ISTH : International Society of Thrombosis and Haemostasis

ITP : Immune thrombocytopenic purpura.

JAAM : Japanese Association for Acute Medicine

LMWH : Low molecular weight heparin
 PAI-1 : Plasminogen activator inhibitor
 PCC : Prothrombin Complex Concentrate

PF4 : Platelet factor-4

PFA-100 : Platelet function analyzer-100

PT : Prothrombin time RBCs : Red blood cells

rFVIIa : Recombinant factor VIIa SDPs : Single donor platelets

SIRS : Systemic inflammatory response syndromeTAFI : Thrombin activatable fibrinolysis inhibitor

TCT: Thrombin clotting time

TEG: Thrombelastography

TFPI : Tissue factor pathway inhibitor **tPA** : Tissue plasminogen activator

TTP : Thrombotic thrombocytopenic purpur

TXA : Tranexamic acid

UFH : Unfractionated heparin

uPA : Urokinase-type plasminogen activator

VKA : Vitamin K antagonismVTE : Venous thromboembolismvWD : Von Willebrand disease

vWF : Von Willebrand factor

α2-PI : Alpha2-plasmin inhibitor

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Introduction

aematologic factors, in particular platelets and the coagulation system, play an important role in the pathogenesis of organ failure in the intensive care unit (*Levi and Schultz*, 2011).

Platelets are irregularly shaped, disk-like fragments of the membrane of their precursor cell, the megakaryocyte. Megakaryocytes shed platelets in the bone marrow sinusoids. From there the platelets are released to the blood, where they function in hemostasis. Several factors stimulate megakaryocytes to release platelets, including the hormone thrombopoietin, which is generated and released into the bloodstream when the number of circulating platelets drops. Platelets have no defined nucleus. They are one fourth to one third the size of erythrocytes. Platelets possess physiologically important proteins, stored in intracellular granules, which are secreted when the platelets are activated during coagulation (English, 2003).

The liver plays a major role in haemostasis, as most of the coagulation factors, anticoagulant proteins and components of the fibrinolytic system are synthesized by hepatic parenchymal cells. Additionally, the reticuloendothelial system of the liver

helps to regulate coagulation and fibrinolysis by clearing these coagulation factors from the circulation (*De Sancho and Pastores*, 2005).

Failure of these haematologic systems is common in intensive care patients and may range from isolated thrombocytopenia or prolonged global clotting tests to complex defects, such as disseminated intravascular coagulation. There are many causes for a deranged coagulation in critically ill patients, and each of these underlying disorders may require specific therapeutic management. Hence, a proper differential diagnosis and initiation of adequate (supportive) treatment strategies are crucial to reduce morbidity and mortality in critically ill patients with coagulation abnormalities (*Levi and Schultz*, 2011).

Many critically ill patients in the intensive care unit (ICU) present with failure of haemostatic system, most prominently platelets and the coagulation system. In general, coagulation abnormalities require prompt and proper identification of the underlying cause because many of these disorders may necessitate very different therapeutic management strategies (*Levi and Opal*, 2006).

Haemostasis is the process of forming clots in the walls of damaged blood vessels and preventing blood loss while maintaining blood in a fluid state within the vascular system (*Guyton and Hall, 2006*).

Failure of haemostasis is common in critically ill patients and may be complex and multifactorial in pathogenesis. As haemostatic failure may complicate a wide range of medical, surgical and obstetric disorders, definitive diagnosis and specific therapy can significantly impact on outcome. Frequently, complex tests are required for definitive diagnosis, but the urgency of the situation cannot always wait for the results, and therapy may be initiated on clinical evidence with minimal laboratory information. Consultation with a clinical haematologist is strongly recommended (*Bersten and Soni*, 2009).

The incidence of thrombocytopenia (platelet count $<150\times10^9$ /L) in critically ill medical patients is 35 to 44%.A platelet count of $<100\times10^9$ /L is seen in another 30 to 50% of patients. A prolonged global coagulation time such as the prothrombin time (PT) or the activated partial thromboplastin time (aPTT) occurs in 14 to 28% of intensive care patients (*Levi and Opal, 2006*).

Thrombocytopenia is important because it increases the risk of bleeding, alters plans for care, and serves as a marker of morbidity and mortality. Thrombocytopenia may also be a manifestation of a disease that promotes clotting, such as heparin-induced thrombocytopenia (HIT) or disseminated intravascular coagulation (DIC) (*Strauss et al.*, 2002).

Thrombocytopenia is one of the most common laboratory abnormalities in intensive care unit (ICU) patients. Thrombocytopenia can be a result of increased (nonimmune or immune) platelet destruction, haemodilution, platelet sequestration (as in hypersplenism), or decreased platelet production (*Vanderschueren et al.*, 2000).

The most common causes of acquired coagulation disorders include DIC, trauma, liver disease, vitamin K deficiency and exposure to anticoagulants (*Irwin and Rippe*, 2014).

In general, there may be clinical features suggesting local or generalized failure of the haemostatic system. Clinical history is important, especially with respect to previous bleeding problems, family history, comorbid medical conditions and medications. The nature of surgery or

an invasive intervention may have haemostatic issues that need specific consideration (*Bersten and Soni*, 2009).

In most clinical settings, laboratory haemostatic screening tests are readily available, with near-patient testing techniques continuing to improve. A full blood count, prothrombin time (PT), activated partial thromboplastin time (APTT), fibrinogen level, D-Dimer, thrombin clotting time (TCT) provide a broad screen for most clinically significant haemostatic disorders. Based on these results, and following consultation with a haematologist, further specific tests of haemostasis may be performed (e.g. mixing studies, factor assays, platelet function tests and test of fibrinolytic function) (Rochon and Shore-Lesserson, 2006).

The laboratory investigation of a patient with a potential haemostatic defect depends on the degree of urgency. It may be necessary to administer blood component therapy without a definitive haemostatic defect being established. A detailed clinical history and screening laboratory investigations prior to elective procedures will often avert undiagnosed emergency haemostatic crises (*Bersten and Soni, 2009*).