Potential Beneficial Effects of Verapamil on Liver fibrosis in Rats

Thesis

Submitted in partial fulfillment of Master Degree in Pharmacology

Presented by

Mohamed El-Sayed Mahmoud

M.B., B.Ch.- Master Degree in Anesthesia Clinical Demonstrator of Pharmacology Faculty of Medicine Cairo University

<u>Under Supervision of</u>

Prof. Dr. Nadia Ali Gouda

Professor of Pharmacology Faculty of Medicine Cairo University

Dr. Amani Nabil Shafik

Assistant Professor of Pharmacology Faculty of Medicine Cairo University

Dr. Mostafa Mahmoud Khodeir

Lecturer of Pathology Faculty of Medicine Cairo University

Faculty of Medicine Cairo University

2009

Acknowledgement

I am extremely grateful to my God ALLAH the most gracious, most merciful for supporting me in this thesis.

I would like to express my deepest gratitude to **professor Dr. Nadia Ali Gouda**, Professor of Pharmacology, Faculty of Medicine, Cairo University for her encouragement and support during this work.

I would like to express my appreciation to **Dr. Amani Nabil Shafik**, Assistant Professor of Pharmacology, Faculty of Medicine, Cairo University.

I would like to express my gratitude to **Dr. Mostafa Mahmoud Khodeir**, Lecturer of Pathology, Faculty of Medicine, Cairo University for his help during this work.

I would like to express my gratitude to **Dr. Dalia Ahmed Mohamed**, Assistant Professor of Public health and community medicine, Faculty of Medicine, Cairo University for participation in statistics of this work.

No words can describe my sincere gratitude to all stuff members of pharmacology department, faculty of Medicine, Cairo University for their support and encouragement.

I would like to express my deepest gratitude to my family.

Contents

	Page
 List of Abbreviations 	
• List of Tables	
• List of Figures	
Introduction and aim of the work	
Review of litrature	
 Liver fibrosis Management of liver fibrosis Verapamil Silymarin Models of induction of liver fibrosis 	Page 1 Page 23 Page 28 Page 38 Page 42
Materials and Methods Results Discussion Summary References Arabic summary	Page 50 Page 56 Page 72 Page 82 Page 84

List of Abbreviations

ALT	Alanine aminotransferase
Apo A1	Apolipoprotein A1

As Arsenic

ASMA Alpha-smooth muscle actin
AST Aspartate aminotransferase
Bcl-xL B-cell lymphoma-extra large
CBDL Common bile duct ligation
CCL21 C-C chemokine ligand
CCl₄ Carbon tetrachloride

CINC Cytokine-induced neutrophil chemoattractant

CTGF Connective tissue growth factor
DILI Drug-induced liver injury
DMN Dimethylnitrosamine
ECM Extracellular matrix proteins
EGF Epidermal growth factor

ET-1 Endothelin-1

FGD Fibroblast growth factor

GSH Glutathione
HA Hyalouronic acid
Hap Haptoglobin
HCV Hepatitis C virus

HGF Hepatocyte growth factor HSCs Hepatic stellate cells

ICAM-1 Intercellular adhesion molecule 1

IFN-γ Interferon-γ

IGF-I & II Insulin growth factors I & II

IL-6 Interleukin-6

LBP Lipopolysaccharide-binding protein
mAb Monoclonal anti-alpha-actinin antibody
MCP-1 Monocyte chemoattractant protein-1
MIP-2 Macrophage inflammatory protein-2

Matrix-metalloproteinases MMP N-acetyl-ρ-benzoquinone imine **NAPQI** Neural cell adhesion molecule **NCAM** NS3 Nonstructural protein 3 NS5 Nonstructural protein 5 **ODNs** Oligodeoxynuleotide Platelet-activating factor PAF **PBHF** Primary biliary hepativ fibrosis **PDGF** Platelet derived growth factor

PIIINP N-terminal propeptide of type III collagen

PPARy Peroxisome proliferators-activated nuclear receptors-y

ROS Reactive oxygen species

SOCs Store-operated calcium channels

SREBP-1c Sterol regulatory element binding protein-1c

TAA Thioacetamide

TFOsTriplex-forming oligonuleotideTGF-αTransforming growth factor-αTGF- $β_1$ Transforming growth factor- $β_1$ TIMPsTissue inhibitor of metalloproteinases

TNF- α Tumor necrosis factor- α VCAM Vascular cell adhesion α -2-M α -2-Macroglobulin

List of Tables

Table		Page
Table 1	Animal grouping & design of the work	Page 52
Table 2	Architectural Changes, Fibrosis/Cirrhosis Scoring System	Page 55
Table 3	Histopathological changes (grade of fibrosis) in the different groups studied	Page 66
Table 4	Serum levels of ALT (Mean \pm SD) in the different groups studied	Page 67

List of figures

Figure		Page
Figure 1	Portal fibrosis	Page 5
Figure 2	Intralobular fibrosis	Page 6
Figure 3	Mild central fibrosis "chicken wire fibrosis"	Page 6
Figure 4	Sclerosing hyalin necrosis	Page 7
Figure 5	Secondary biliary cirrhosis	Page 7
Figure 6	Primary biliary cirrhosis	Page 8
Figure 7	Cellular mechanisms of liver fibrosis	Page 13
Figure 8	α _{IC} Subunit of the L-Type Calcium Channel in Humans	Page 29
Figure 9	Chemical structure of verapamil	Page 31
Figure 10	The three structural components of silymarin	Page 38
Figure 11	Mean Serum level of ALT in groups I, II, III & IV	Page 68
Figure 12	Mean Serum level of ALT in groups IV, V, VI, VII, VIII & IX	Page 68
Figure 13	Mean Serum level of ALT in groups VII after 10 weeks	Page 68
Figure 14	Section in rat liver showing no liver scarring	Page 69
	(Ishak fibrosis score = 0) (Hematoxylin/eosin $x200$)	
Figure 15	Section in rat liver showing no liver scarring	Page 69
	(Ishak fibrosis score = 0)(Masson trichrome x200)	
Figure 16	Section in rat liver showing minimal liver scarring around	Page 69
	some blood vessels (minimal septal fibrosis)	
	(Ishak fibrosis score = 1) (Hematoxylin/eosin x200)	
Figure 17	Section in rat liver showing minimal liver scarring around	Page 70
	some blood vessels (minimal septal fibrosis)	
	(Ishak fibrosis score = 1) (Masson trichrome x400)	
Figure 18	Section in rat liver showing liver scarring around most of	Page 70
	blood vessels (septal fibrosis) (Ishak fibrosis score = 2)	
	(Hematoxylin/eosin x200)	D =0
Figure 19	Section in rat liver showing liver scarring around most of	Page 70
	blood vessels (septal fibrosis)	
F: 00	(Ishak fibrosis score = 2) (Masson trichrome x400)	D 51
Figure 20	Section in rat liver showing liver scarring that forms	Page 71
	bridges between blood vessels (porto-portal bridging)	
F: 21	(Ishak fibrosis score = 3) (Hematoxylin/eosin x200)	Daga 71
Figure 21	Section in rat liver showing liver scarring that forms	Page 71
	bridges between blood vessels (porto-portal bridging)	
Figure 22	(Ishak fibrosis score = 3) (Masson trichrome x 200)	Page 71
Figure 22	Section in rat liver after 3 rd injection at 2 nd week of pig	rage /I
	serum injection showing increased expression for α -SMA	
	in perisinusoidal and perivascular cells.	
	(DAB chromogen and hematoxylin counter stain x 200)	

Introduction

Hepatic fibrosis, or scarring of the liver, is the final stage of a wound healing response that invariably follows chronic liver injury of any etiology. The point that the response to recurrent injury in the liver is similar in multiple different types of liver disease underscores the value of identifying common regulatory components of the fibrotic response because such components theoretically could be targeted without respect to cause of disease. However, it is important to emphasize that, at least in theory; different pathologic patterns of fibrosis (biliary, perisinusoidal, pericentral) can occur and thus may merit different types of therapies. Fibrosis develops with different spatial patterns and is a consequence of different prevalent mechanisms according to the diverse causes of parenchymal damage. Indeed, fibrosis, observed as a consequence of chronic viral infection is initially concentrated within and around the portal tract, while fibrosis secondary to toxic/metabolic damage is located mainly in the centrolobular areas. In addition, it is increasingly evident that different cell types are involved in the deposition of fibrillar extracellular matrix during active hepatic fibrogenesis: hepatic stellate cells are mainly involved when hepatocellular damage is limited or concentrated within the liver lobule, whereas portal myofibroblasts and fibroblasts provide a predominant contribution when the damage is located in the proximity of the portal tracts. Progressive fibrosis ultimately leads to a gross disruption of the normal liver architecture (cirrhosis) and subsequent impairment of liver function.

The 'activated' hepatic stellate cell (HSC) is the pivotal cell type involved in the development of liver fibrosis, simultaneously secreting excessive scar proteins (collagens) and potent inhibitors of collagen degradation –

tissue inhibitors of matrix metalloproteinases (TIMPs). Resolution of fibrosis is characterized by matrix degradation and is associated with HSC apoptosis and the increased activity of matrix metalloproteinase enzymes (MMPs).

Activation of stellate cells is accompanied by a marked increase in proteins that are characteristic of contractile cells (i.e. such as smooth muscle actin and smooth muscle myosins). Stellate cell contraction is important in the injured liver because it may contribute to the collapse and shrunken state of cirrhotic livers, and because it also appears to play a role in portal hypertension. Thus, stellate cell contractility, although not directly related to fibrosis, is an important physiologic target.

Currently, the most effective treatment for hepatic fibrosis is removal of the causative agent.

Since the HSC plays a central role in the development of liver fibrosis, this cell is a major target.

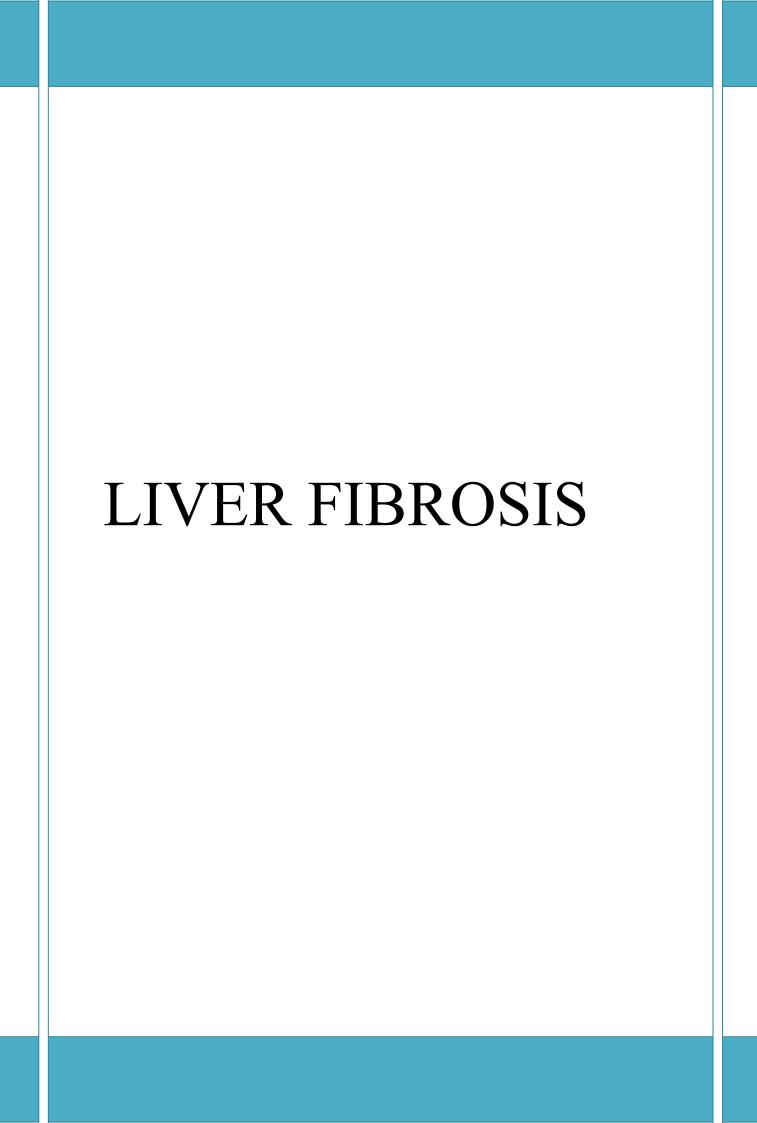
Verapamil, a phenylalkylamine calcium channel blocker, can prevent energy metabolism disorder in hepatocytes induced by calcium overloading through blockage of voltage-dependent calcium channels and Ca²⁺ inflow. Verapamil as an angiotenic, can increase local blood circulation and oxygen supply, inhibit lipid peroxidation, antioxidation, and thereby limit hepatonecrosis *in vivo*.

The pig serum model induces liver fibrosis without severe inflammation. Because we would like to mainly focus on the effect of verapamil on activated HSC, we employed this model in the present study.

Aim of the Work

The aim of the present work is to:

- 1- To investigate the effect of variable doses of a calcium channel blocker 'VERAPAMIL" as a prophylactic and therapeutic antifibrotic drug in experimental liver fibrosis induced in rats by pig serum.
- 2- To compare verapamil to standard prophylactic therapy silymarin.
- 3- To demonstrate the beneficial effect of combining verapamil to the standard prophylactic therapy silymarin.



Liver fibrosis

Hepatic fibrosis is the wound-healing response of the liver to repeated injury (Iredale, 2003). Liver fibrosis results from chronic damage to the liver in conjunction with the accumulation of extracellular matrix protein (ECM) proteins, which is a characteristic of most types of chronic liver diseases. The accumulation of ECM proteins distorts the hepatic architecture by forming a fibrous scar, and the subsequent development of nodules of regenerating hepatocytes defines cirrhosis. Cirrhosis produces hepatocellular dysfunction and increased intrahepatic resistance to blood flow, which result in hepatic insufficiency and portal hypertension, respectively (Gines et al., 2004).

Liver fibrosis was thought to be a passive and irreversible process due to the collapse of the hepatic parenchyma and its substitution with a collagen-rich tissue (Schaffner and Klion, 1968; Popper and Uenfriend, 1970). Currently, it is considered a model of the woundhealing response to chronic liver injury (Albanis and Friedman, 2001). Early clinical reports in the 1970s suggested that advanced liver fibrosis is potentially reversible (Soyer et al., 1976). However, liver fibrosis received little attention until the 1980s, when hepatic stellate cells (HSCs), formerly known as lipocytes, Ito cells, or perisinusoidal cells, were identified as the main collagen-producing cells in the liver (Friedman et al., 1985). This cell type, first described by von Kupffer in 1876, undergoes a dramatic phenotypic activation in chronic liver diseases with the acquisition of fibrogenic properties (Geerts, 2001). Methods to obtain HSCs from both rodent and human livers were rapidly standardized in the 1980s (Otto and Veech, 1980; Friedman et al., 1992), and prolonged culture of HSCs on plastic was widely accepted as a model for the study of activated HSCs (Rockey et al., 1992). Key

signals that modulate HSCs' fibrogenic actions besides HSCs, are portal myofibroblasts and cells of bone marrow (Forbes et al., 2004; Ramadori and Saile, 2004). Rapid and slower fibrosers are identified, and genetic and environmental factors influencing fibrosis progression are known (Bataller et al., 2003). Since the demonstration, in the 1990s, that even advanced liver fibrosis is reversible, However, the most effective therapy for treating hepatic fibrosis to date is still to remove the causative agent (Bataller and Brenner, 2001). A number of drugs are able to reduce the accumulation of fibrous tissue in experimental models of chronic liver injury. Renin-angiotensin system blockers and antioxidants are the most promising drugs, although their efficacy has not been tested in humans.

The onset of liver fibrosis is usually insidious, and most of the related morbidity and mortality occur after the development of cirrhosis (**Poynard et al., 2000**). In the majority of patients, progression to cirrhosis occurs after an interval of 15–20 years. Cirrhosis is also a risk factor for developing hepatocellular carcinoma. Liver fibrosis progresses rapidly to cirrhosis in several clinical settings, including repeated episodes of severe acute alcoholic hepatitis, subfulminant hepatitis, and fibrosing cholestasis in patients with HCV reinfection after liver transplantation (**Berenguer et al., 2003**).

Classifications of Liver Fibrosis:

There are 2 main types, congenital and acquired liver fibrosis. The congenital type is a genetic disorder, which causes polycystic liver diseases, while the acquired type has many different categories and is mainly caused by liver cell injuries (**Desmet, 1992; Friedman, 2000; Hui and Friedman, 2003**).

Causatively, liver fibrosis can be classified as:

- 1. Viral hepatitis fibrosis: Usually caused by chronic hepatitis B, C, and D. About 15% of HBV and 85% of HCV infected persons will develop chronic hepatitis and lead to fibrosis. In which, the liver shows peri-portal area inflammation, piecemeal necrosis and fibrosis. With such large population being affected, this is the most important category of the liver fibrosis (**Arias and Irwin, 1994**)
- 2. Parasitic infestation fibrosis: This kind of liver fibrosis is mainly happening in developing countries and is caused by schistosomiasis. The recurrent infection and the eggs of schistosoma accumulated in the liver can cause liver fibrosis and cirrhosis (**Rockey**, 2005).
- 3. Alcoholic fibrosis: It is mainly caused by the oxidized metabolite of alcohol, acetaldehyde. Its incidence is positively related to the amount of alcohol consumped. Alcoholic fibrosis causes two morphological changes in the liver: fatty liver and cellular organelles deterioration. The fibrosis first appears around the center veins (central hyaline sclerosis) associated with liver parenchymal inflammation and the fibrosis gradually expands to the whole liver (**Arias and Irwin, 1994**).

4. Biliary fibrosis: There is primary and secondary biliary fibrosis. Primary biliary hepatic fibrosis (PBHF) is an autoimmune disorder in which chronic bile retention caused the liver fibrosis (**Dufour et al., 1997**). It affects female around the age 40 to 60. In serum tests there is elevated gamma globulin and serum is positive for the anti-mitochondria antibody. Pathological studies found that the fibrosis mainly around the micro-bile ducts and peri-portal area fibrosis and inflammation. Secondary biliary fibrosis happens following the obstruction of the bile ducts, which causes peri-portal inflammation and progressive fibrosis (**Arias and Irwin, 1994**).

- 5. Metabolic fibrosis: It is not common. Wilson's disease and hemochromatosis are the main disorders that cause metabolic fibrosis. The former is a genetic disorder and causes copper metabolism disorder and deposits in the liver. The latter is an iron metabolic disorder and causes hemoglobin deposits in the liver (**Rockey**, 2005).
- 6. Intoxication fibrosis: When long-term contact with liver-toxic substances, such as carbon tetrachloride, organophosphorus, dimethyl nitrosamine, thioacetamide, or taking liver toxic medications, such as isoniazid, thiooxidizing pyrimidine, wintermin, tetracycline, acetaminophen etc. can all cause various degrees of liver cell injuries, necrosis, bile retention, or allergic inflammation and cause liver fibrosis (Rockey, 2005).
- 7. Malnutritional fibrosis: This type is mainly caused by insufficient or imbalanced nutritional intake (Rockey, 2005).
- 8. Cardiogenic fibrosis: Chronic congestive heart failure can cause long lasting liver vein stagnation. Which causes ischemic degeneration of the

liver cells. In this type of liver fibrosis, the connective tissue hypertrophy starts at the center of the liver lobule and gradually expands to rest of the lobule (Giallourakis et al., 2002).

Pathologically, Liver fibrosis can be classified as:

1. Portal area fibrosis: There is fibroblasts proliferation and fibers expansion from the portal areas to the lobule. Finally, these fibers connected to form bridging septa. This kind of fibrosis is mainly seen in viral hepatitis and malnutritional liver fibrosis (Figure 1) (**Klatt, 2006**).

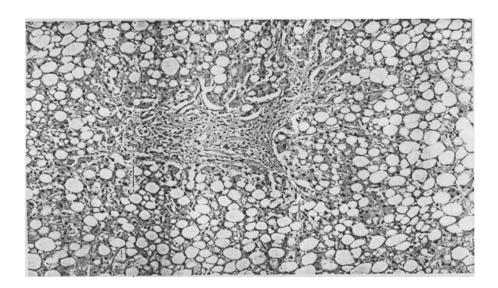


Figure 1: Portal fibrosis:Fibrous expansion of portal fields with fibrosis extending along the terminal centracinar portal veins assuming the appearance of "stellate" fibrosis. Expansion and branching of normal portal fields must not be confused with pathological portal fibrosis which contains inflammatory cell (JANIS AND FRIENDS, 2008).

2. Intra-lobular fibrosis: In which, there is almost no fibroblast found in normal lobule. When large numbers of liver cells degenerate and undergo necrosis, the reticular fiber frame collapses and becomes thick collagen fibers. At the same time, intra lobule fibrotic tissue proliferates and surrounds the liver cells (Figure 2) (Klatt, 2006).