

Evaluation of Lactic Acid Bacteria Isolated from the Honey Bee *Apis mellifera* L. for the Control of the American Foulbrood Disease

A Thesis
Submitted in Partial Fulfillment of the Requirements for the Degree of Master of Science
(Entomology)

By

Fatma Mahmoud El-Sayed Mohammed (B.Sc. Entomology)

Supervisors

Prof. Dr. Akila Mohammed El-Shafei

Professor of Entomology Entomology department - Faculty of Science - Ain Shams University

Prof. Dr. Ahmed Saad Abou Zeid

Professor of Entomology Entomology department - Faculty of Science - Ain Shams University

Dr. Shireen Ahmed Mahmoud Ma'moun

Lecturer of Entomology Entomology department - Faculty of Science - Ain Shams University

> Department of Entomology Faculty of Science Ain Shams University

Biography

Name: Fatma Mahmoud El-Sayed Mohammed.

<u>Degree awarded:</u> B.Sc. (Entomology).

Department: Entomology.

Faculty: Science.

<u>University:</u> Ain Shams University.

Date of Graduation: July 2013.

Date of Appointment: April 2014.

Occupation: Demonstrator, Department of

Entomology, Faculty of Science, Ain

Shams University.

Date of registration

For the M.Sc. Degree: April 2015.

Evaluation of lactic acid bacteria isolated from the honey bee *Apis mellifera* L. for the control of the American foulbrood disease

Board of Supervision:

Prof. Dr. Akila Mohammed El-Shafei

Professor of Entomology Faculty of Science Ain Shams University

Prof. Dr. Ahmed Saad Abou Zeid

Professor of Entomology Faculty of Science Ain Shams University

Dr. Shireen Ahmed Mahmoud Ma'moun

Lecturer of Entomology Faculty of Science Ain Shams University

Approval Sheet

Evaluation of lactic acid bacteria isolated from the honey bee *Apis mellifera* L. for the control of the American foulbrood disease

Approved By:			
Prof. Dr			
Prof. Dr	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • •	
Prof. Dr	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	
Prof. Dr	• • • • • • • • • • • • • • • • • • • •	•••••••	
	,	Committee in c Date: /	0 ,

<u>ACKNOWLEDGEMENTS</u>

First of all, thanks to "ALLAH" to whom I relate any success in achieving any work in my life.

My deepest appreciation and respect with sincere thanks go to **Prof. Dr.** Akila El Shafei, Professor of Entomology for suggesting the problem, her excellent ideas, energetic guidance, constructive discussion and close supervision.

I am also particularly indebted to **Prof. Dr. Ahmed Saad Abou Zeid,** Professor of Entomology for his generous assistance and guidance.

My deepest appreciation and respect with sincere thanks go to **Dr. Shireen Ahmed Mahmoud Ma'moun**, lecturer of Entomology for suggesting the problem, her excellent ideas, energetic guidance, constructive discussion and close supervision.

Finally, I wish to express my deep thanks to **Dr. Rasha**Mohamed Ahmed Farag, Researcher in Apiculture Research

Department, Plant Protection Institute for her help.

Special thanks go to all members of the Entomology Department, Faculty of Science, Ain Shams University for their encouragement and help during my work.

Finally, ALLAH was the only one who made this work possible.

Abstract

The purpose of this study is to investigate a new applicable, reliable, promising and highly effective treatment for American Foulbrood (AFB) disease in honey bee colonies. Some of the honey bee gut microbial diversity in worker bees using 560 amplicon assays of the 16S rRNA gene were investigated. The presence of nine novel anaerobic lactic acid bacterial (LAB) flora were reported within honey bee gut. Four of the LABs are carefully related to four different strains of Lactobacillus plantarum species. Two are closely identical to two different strains of *Lactobacillus kunkeei* species. One is closely related to a strain of Lactobacillus pentosus species. The last two are matching two different strains of Lactobacillus sp.. A strong inhibitory effect of the honey bee stomach LAB flora on AFB bacterial pathogen, Paenibacillus larvae larvae (P. l. larvae) growth in vitro were demonstrated. The individual LAB phylotypes showed different inhibition zones ranges from 0.4 to Artificial infection was accompanied by the administration of a mixture of five of the most effective endogenous LABs, previously tested for their inhibitory effect on agar plates in vitro. It was observed that the honey bee endogenous LAB inhibited P. l. larvae in an in vivo system. LAB mixture added to the larval food in honey bee colonies significantly reduced the number of infected larvae when pooled data from all experiments were analyzed (P $\simeq 0.000$, P < 0.001). Confidence intervals analysis on the effect of the time LAB was added to the colony didn't show significant different from adding LAB to the food on first or second day post infection and throughout the feeding period. Both in vitro and in vivo studies demonstrated that the LAB microbiota in Apis mellifera inhibit one important honey bee pathogen, the bacterial brood pathogen

P. l. larvae that is cause of the brood disease AFB. The results point to new avenues for the prophylactic or therapeutic treatment of honey bee diseases.

Keywords:

Honey bee, *Apis mellifera*, American Foulbrood, Lactic Acid Bacteria, LAB identification, Probiotic treatment and AFB control.

Contents

Conte	ents	Pages
ACK	NOLEDGMENT	I
Abstı	act	II
List o	of Tables	IV
List o	of Figures	\mathbf{V}
List o	of Abbreviation	XVI
I.	Introduction	1
II.	Literature Review	6
1.	Biology of Honey bees	6
2.	American Foulbrood (AFB) disease; Diagnosis	
	and treatment	8
3.	Lactic Acid Bacteria (LAB); A symbiotic micro-	
	flora in the gut of honey bees	17
4.	Lactic Acid Bacteria (LAB); as a control agent of	
	American Foulbrood disease	24
III.	Materials and Methods	30
1.	Maintenance of the honey bee, Apis mellifera,	
	colonies	30
2.	American Foulbrood (AFB) bacterial pathogen,	
	Paenibacillus larvae larvae; Isolation and	
	preparation	31
2	2.1. Source of the bacterial pathogen	31

pathogen P. l. larvae 31 2.3. Identification and characterization of the bacterial pathogen 32 2.3.1. Morphological tests 33 2.3.2. Microscopical tests 33 2.3.3. Biochemical tests 33 3. The probiotic lactic acid bacteria (LAB); Isolation and identification 34 3.1. Isolation of LAB 34 3.2. Identification of Lactic acid bacteria; DNA preparation and manipulation 35 3.2.1. DNA isolation from LAB cultured colonies 35 3.2.2. PCR amplification of 16S-rRNA gene 36 3.2.3. Agarose gel electrophoresis 38 3.2.4. Sequence analysis 39 4. Lactic acid bacteria (LAB) inhibition bioassays against P. l. larvae bacterial spores 40 5. Field experiment 40 5.1. Colonies and treatments 41	2.2. Isolation and Cultivation of the bacterial	
bacterial pathogen 32 2.3.1. Morphological tests 33 2.3.2. Microscopical tests 33 2.3.3. Biochemical tests 33 3. The probiotic lactic acid bacteria (LAB); Isolation and identification 34 3.1. Isolation of LAB 34 3.2. Identification of Lactic acid bacteria; DNA preparation and manipulation 35 3.2.1. DNA isolation from LAB cultured colonies 35 3.2.2. PCR amplification of 16S-rRNA gene 36 3.2.3. Agarose gel electrophoresis 38 3.2.4. Sequence analysis 39 4. Lactic acid bacteria (LAB) inhibition bioassays against P. l. larvae bacterial spores 40 5. Field experiment 40	pathogen P. l. larvae	31
2.3.1. Morphological tests	2.3. Identification and characterization of the	
2.3.2. Microscopical tests	bacterial pathogen	32
2.3.3. Biochemical tests	2.3.1. Morphological tests	33
3. The probiotic lactic acid bacteria (LAB); Isolation and identification	2.3.2. Microscopical tests	33
and identification	2.3.3. Biochemical tests	33
3.1. Isolation of LAB	3. The probiotic lactic acid bacteria (LAB); Isolation	
3.2. Identification of Lactic acid bacteria; DNA preparation and manipulation	and identification	34
preparation and manipulation	3.1. Isolation of LAB	34
3.2.1. DNA isolation from LAB cultured colonies	3.2. Identification of Lactic acid bacteria; DNA	
colonies	preparation and manipulation	35
3.2.2. PCR amplification of 16S-rRNA gene 36 3.2.3. Agarose gel electrophoresis	3.2.1. DNA isolation from LAB cultured	
3.2.3. Agarose gel electrophoresis	colonies	35
3.2.4. Sequence analysis	3.2.2. PCR amplification of 16S-rRNA gene	36
4. Lactic acid bacteria (LAB) inhibition bioassays against <i>P. l. larvae</i> bacterial spores	3.2.3. Agarose gel electrophoresis	38
against <i>P. l. larvae</i> bacterial spores	3.2.4. Sequence analysis	39
5. Field experiment	4. Lactic acid bacteria (LAB) inhibition bioassays	
	against P. l. larvae bacterial spores	40
5.1. Colonies and treatments	5. Field experiment	40
	5.1. Colonies and treatments	41
5.2. Monitoring larval mortality 42	5.2. Monitoring larval mortality	42
5.3. Data analysis	5.3. Data analysis	43
	5. Field experiment	4

5.4. Checking colonies and Culturing of the	
probiotic LAB and the pathogen P. l. larvae	
from honey bees	44
5.5. Field efficacy	45
IV. Results	46
1. Isolates of the American foulbrood (AFB)	
bacterial pathogen; Paenibacillus larvae larvae	46
1.1. Identification and characterization of <i>P. l.</i>	
larvae, AFB bacterial pathogen	46
1.1.1. Morphology of <i>P. l. larvae</i> bacterial	
colony	46
1.1.2. Microscopic identification of <i>P. l. larvae</i>	
bacterium	47
1.1.3. Biochemical identification of <i>P. l. larvae</i>	
bacteriam	48
2. Isolates of the probiotic lactic acid bacteria	
(LAB)	49
2.1. Identification of Lactic acid bacteria; PCR	
amplification and sequencing results	52
2.1.1. PCR amplification of 16S-rRNA gene	52
2.1.2. Sequencing results	53
	33
3. Lactic Acid Bacteria (LAB) inhibition bioassay	76
results	76

4.	Field experiment	79
۷	1.1. Monitoring larval mortality	79
۷	1.2. Data analysis	80
۷	4.3. Checking colonies and Culturing of the	
	probiotic LAB and the pathogen P. l. larvae	
	from honey bees	85
۷	4.4. Field efficacy	92
v.	Discussion	93
1.	Lactic Acid Bacteria (LAB); A symbiotic micro-	
	flora in the gut of honey bees	93
2.	Lactic Acid Bacteria (LAB); as a control agent of	
	American Foulbrood disease	103
VI.	Conclusion	117
VII.	Recommendation	119
VIII.	Summary	121
IX.	References	125
X.	Arabic summary	160

List of Tables

Tables	Pages
Table 1: Inhibition zones formed by the nine-lactic acid bacteria	
(LAB) probiotics against <i>P. l. larvae</i> bacterial spores	79
Table 2: Numbers of survived and dead honey bee larvae and	
capped cells in the four groups of the experiment at zero time	
(before infection) and seventh day post infection and treatment	
with LAB probiotics (G1: -ve control, G2: +ve control, G3:	
treatment on the first day post infection and G4: treatment on	
second day post infection)	80
Table 3: Survive * Treatment Crosstabulation of expected count	
of dead and survived brood larvae on seventh day post	
experiment (G2: +ve control, G3(P): treatment on the first day	
post infection and G4(PA): treatment on second day post	
infection	81
Table 4: Survive * Treatment Crosstabulation of percent within	
treatment of dead and survived brood larvae on seventh day post	
experiment (G2: +ve control, G3(P): treatment on the first day	
post infection and G4(PA): treatment on second day post	
infection)	82
Table 5: Person's Chi-Squared Test	82

List of Figures

Figures	Pages
Figure (1): Colonies of hybrid Carniolian honey bees;	
Apis mellifera carnica, in the apiary yard of the	
Apiculture Research Department, Plant Protection	
Research Institute	30
Figure (2): P. l. larvae streaking growth on J-agar plate.	46
Figure (3): Morphological characters of P. l. larvae	
Colony	47
Figure (4): Paenibacillus larvae larvae vegetative cells	
stained with crystal violet stain	47
Figure (5): Paenibacillus larvae larvae spore cells	
stained with crystal violet stain	47
Figure (6): Holst milk test (a) P. l. larvae spores	
cleared skimmed milk. (b) control	48
Figure (7): Catalase test: bubbly foam is not indicated	49

Figure (8): Nine cultured MRS agar plates with the	
selected twelve white, small and rounded colonies of	
Lactic Acid Bacteria (Lab 1 to 12)	50
Figure (9): Twelve Anaerobic subcultures of Lactic	
Acid Bacteria (Lab 1 to 12) isolated on MRS agar media	51
Figure (10): 16S rRNA amplified region (560 bp) from	
the twelve LAB subcultured colonies (Lanes named	
from L1 to L12 respectively)	52
Figure (11): Forward nucleotide sequences of the 513	
bp lactic acid bacteria (Lab 1) PCR amplicon, of the 16S	
ribosomal RNA gene	55
Figure (12): BLAST server results of the lactic acid	
bacteria (Lab 1) forward PCR sequence and its	
alignment statistics with its first match, Lactobacillus	
plantarum strain LB11 16S ribosomal RNA gene,	
partial sequence	56

Figure (13): Forward nucleotide sequences of the 539	
bp lactic acid bacteria (Lab 2) PCR amplicon, of the 16S	
ribosomal RNA gene	57
Figure (14): BLAST server results of the lactic acid	
bacteria (Lab 2) forward PCR sequence and its	
alignment statistics with its first match, Lactobacillus	
sp. strain 86056 16S ribosomal RNA gene, partial	
sequence	58
Figure (15): Forward nucleotide sequences of the 528	
bp lactic acid bacteria (Lab 5) PCR amplicon, of the 16S	
ribosomal RNA gene	59
Figure (16): BLAST server results of the lactic acid	
bacteria (Lab 5) forward PCR sequence and its	
alignment statistics with its first match, Lactobacillus	
plantarum strain CAG18b 16S ribosomal RNA gene,	
partial sequence	60