



**Faculty of Scien** 

# Bacteriophages targeting antibiotic resistant pathogenic bacteria in infected broiler chickens

A Thesis
Submitted for the Degree of Doctor of Philosophy in
Microbiology (Virology)

By
Mayada Mahmoud Abd El-Samee

M.Sc. (2014)

**Under Supervision** 

Prof. Dr.

#### **Ahmed Barakat Barakat**

Professor of Virology
Microbiology Department
Faculty of Science - Ain Shams University

Dr.

#### Ahmed Abd El-Rahman Askora

Assistant Prof. of Molecular Biology
Botany Department
Faculty of Science
Zagazig University

Prof. Dr.

#### Sayed Emam Hassan

**Professor of Pharmacology Animal Health Research Institute** 

Dr.

#### Omar El-Farouk Rabie

Lecturer of Microbiology Microbiology Department, Faculty of Science Ain Shams University





# Bacteriophages targeting antibiotic resistant pathogenic bacteria in infected broiler chickens

### A Thesis Submitted for the Degree of Doctor of Philosophy in Microbiology (Virology)

By
Mayada Mahmoud Abd El-Samee
M.Sc. (2014)



كلية العلوم قسم الميكروبيولوجي

أسم الطالبة: مياده محمود عبد السميع محمد

عنوان الرسالة: استهداف البكتيريا المقاومة للمضادات الحيوية والممرضة

لدواجن التسمين باستخدام البكتيريوفاج

الدرجة العلمية: دكتوراة الفلسفة في العلوم

#### لجنة الإشراف:

* * *		
السادة المحكمين	الوظيفة	التوقيع
أ.د/ أحمد بـركـات	أستاذ الفيروسات -كلية العلوم - جامعة عين شمس	
أ.د/ السيد إمام حسن	استاذ الفار ماكولوجى - بمعهد بحوث صحة الحيوان	
أ.م.د/ أحمد عبدالرحمن عسكورة	استاذ مساعد البيولوجيا الجزيئية - كلية العلوم — جامعة الزقازيق	
د/ عمر الفاروق ربيع السيد	مدرس الميكروبيولوجي -كلية العلوم - جامعة عين شمس	
لجنة التحكيم:		
السادة المحكمين	الوظيفة	التوقيع
أ. د/ محمد توفيق شعبان محمد	أستاذ الميكروبيولوجي وعميد كلية العلوم - جامعة المنوفية	
أ.د/ محمد السيد محمد محمد	أستاذ ورئيس قسم الأمراض المشتركة – كلية الطب البيطرى - جامعة الزقازيق	
أ.د/ أحمد بسركات بسركسات	أستاذ الفيروسات -كلية العلوم - جامعة عين شمس	
أ.م.د/ أحمد عبدالرحمن عسكورة	استاذ مساعد البيولوجيا الجزيئية - كلية العلوم – جامعة الزقازيق	

الدراسات العليا:

ختم الإجازة:

اجيزتُ الرسالة بتاريخ: ..../.../٢٠١٨ موافقة مجلس الكلية : ..../.../٢٠١٨

موافقة مجلس الجامعة: ..../١٨/ ٢٠١٨



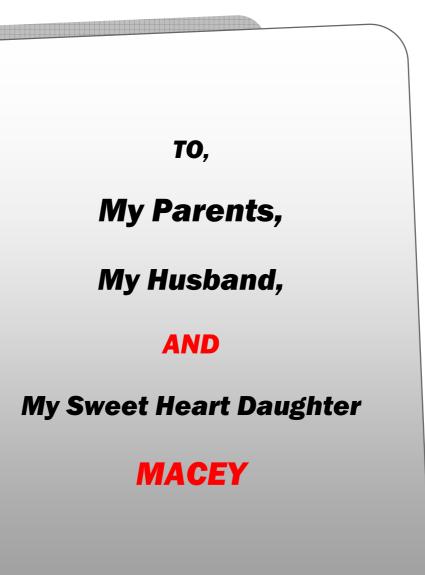
# وقل اعملوا فسيرى الله عملوك ورسوله والمؤمنون

سورة التوبة الآية (١٠٥)









## ACKNOWLEDGMENT

First and foremost, Alhamdulillah, the divine intervention in this academic work. With all my heart, I would like to express my deepest gratitude and sincere thanks to Prof. Dr. Ahmed Barakat Prof. of Virology, Microbiology Department, Faculty of Science, Ainshams University his competent supervision, valuable guidance, great encouragement, help and precious advices throughout this work as he gave me an ideal example of what a university professor should be in his morals, relations and communications with others. I would like to express my deepest gratitude and sincere appreciation to Prof. Dr. Sayed Emam Hassan, Professor of Pharmacology Animal Health Research Institute for his support, continuous efforts, valuable advices and devoting his vast experience to provide me with the best possible piece of advice in this work.

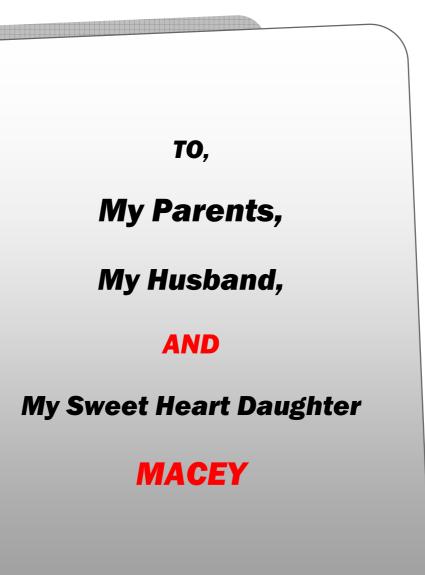
Ahmed Askora, Associate Professor of molecular Biology, Botany Department, Faculty of science, Zagzaig University, Who never ceased until this thesis is structured, and for giving me the opportunity to work on this interesting thesis. I cannot find the words that describe his continuous support and guidance, his unlimited enthusiasm and the many helpful discussions were the cornerstones of this thesis work. I also want to thank him for the enjoyable working atmosphere he created in his group; it has been a real pleasure to work with him.

I would also like to extend my gratitude to Dr. Omar El-Farouk Rabie, Lecturer of Microbiology, Microbiology Department, Microbiology Department, Faculty of Science, Ainshams University, for help and continuous useful advice.

My deep thanks are also extended to the staff members of Microbiology Department, Faculty of Science, Ainshams University.

Last but not least, my heartful thanks to my mother and my father who ecourge me to fulfill my postgraduate studies. Also my heartful thanks to my dear husband who supported me and dealt with my absence and busy studying with a smile.

Special deepest heart full thanks to my love of my life MACEY sweet little daughter whose presence was a great motivation for me to complete my work.



#### **Abstract**

In recent years, the use of bacteriophages as antimicrobial agents controlling pathogenic bacteria in poultry has appeared as a promising new alternative strategy in the face of growing antibiotic resistance, which has caused problems in many fields including medicine, veterinary medicine, and agriculture. Thus, this study was conducted to investigate the prevalence and antimicrobial resistance of different pathogenic bacteria isolated from broiler chickens and the use of their bacteriophages an alternative means of controlling these pathogens. Total numbers of 86 bacterial strains were isolated from broiler chickens samples during this study. Gram-negative bacteria accounted for 87.3% (75 strains) and Gram-positive represented 12.7% (11strains). Major species were Salmonella, E.coli, Proteus spp. Staphylococcus aureus (4%) and Bacillus (1.5%). The most prevalent Salmonella serovars were S. typhimurium (7.5%), S. enteritidis (5.0%), and S. kentucky (3.0%) while the prevalence of E. coli and Proteus sp. was (14.5% and 7.5%) respectively. Antimicrobial resistance profiles of these isolates were determined using the phenotypic agar disc diffusion method. The genes encoding resistance in the resistant strains were screened using PCR. The results showed that bacterial isolates were resistant to at least 3 tested antimicrobial. The PCR results indicated that the tested pathogenic isolates contained antimicrobial resistance gene (blaTEM), which is known to confer resistance. Moreover, the mecA gene was examined in methicillin resistant S. aureus (MRSA) isolates. This study extended to isolate six bacteriophages of different plaques morphology and size targeting these multidrug resistant bacteria. The isolated phages showed variations in their abilities to infect and lysis the target pathogen. These phages were selected for characterization. Bacteriophages active against Salmonella serovars named (Salmacey1, Salmacey2, and Salmacey3), Proteus (Protmacey), S. aureus (Staphmacey), and Bacillus

#### **Abstract**

(Bacmacey). The electron micrographs of negatively stained preparations of these phages revealed that they belong to Myoviridae, Podoviridae, Siphoviridae families. The results of host range assay revealed that these Salmonella bacteriophages were polyvalent and thus capable of infecting different strains of Salmonella serovars, Citrobacter freundii ,Enterobacter and *E.coli.* Bateriophages (Protmacey, Staphmacey, Bacmacey) were restricted to only their specific hosts. Only the isolated bacteriophages were thermostable, inbetween temperature ranges of 30-70°C, and the activity of isolated phages was rapidly decreased toward acidity compared to pH7. The one step growth curve for these phages were determined and the results of burst sizes and latent periods were determined. Molecular analyses indicated that these phages contained double-stranded DNA genomes. The lytic activities of the phages against the most multidrug resistant serovars S. Kentucky, Proteus sp., Staph. aureus and B.cereus as host strain was evaluated. The results showed that these bacteriophages were able to completely kill the growth of tested bacteria in vitro. These results suggest that phages have a high potential for phage application to control Salmonella serovars, Proteus sp., Staph. aureus and B.cereus isolated from broilers in Egypt.

**Keywords**: Food borne pathogens, broilers, multi drug resistant, phages

#### **Table of Contents**

1. Introduction
2. Review of literature6
2.1. Pathogenic bacteria in poultry6
2.1.1. Prevalence of <i>Salmonella</i> in poultry
2.1.2. Sources of <i>Escherichia coli</i> infection for poultry
2.1.3. Prevalence of <i>Proteus</i> spp. in poultry
2.1.4. Prevalence of <i>Staphylococcus</i> infection in poultry
2.1.5. Sources of <i>Bacillus</i> infection in poultry
3.2. The use of antimicrobials in the poultry industry
4.2. Antimicrobial resistance in bacterial poultry pathogens22
4.2.1. Antibiotics resistance of <i>Salmonella</i> in chickens
4.2.2. <i>S. aureus</i> resistance to antibiotics
5.2. Bacteriophage therapy to combat multidrug resistant bacterial infections in poultry
5.2.1. History and discovery of Bacteriophage
5.2.2. Biology, structure and classification of Bacteriophages27
5.2.3. Life Cycle of Bacteriophage
6.2. Bacteriophages for control pathogenic bacteria in poultry and application of phages in biocontrol
7.2 Requirements for selecting a bacteriophage for food applications
8.2. The advantages of phage therapy over antibiotics
3. Materials and methods46
<b>3. I. Materials</b>
3.1. Media
3.2 Media used for biochemical characterization of bacterial isolates 51

3.1.2. Buffers and working solutions	54
3.II. Methods	55
3.1. Samples collection and transportation:	55
3.2. Isolation and identification of pathogenic bacteria	55
3.2.1 Enrichment.	55
3.2.2. Isolation and detection of <i>Salmonella</i>	56
3.2.3. Isolation and detection of <i>E.coli</i>	56
3.2.4 Isolation and detection of <i>Staphylococcus aureus</i>	56
3.2.5. Isolation and detection of <i>Bacillus cereus</i>	56
3.3. Antimicrobial susceptibility testing	57
3.3.1 Screening of resistance genes	57
3.3.2 Detection of antimicrobial resistance genes	59
3.3.3. Detection of antimicrobial resistance genes in <i>S. aureus</i>	59
3.3.3.1. Amplification of the methicillin resistance ( <i>mecA</i> ) gene	59
3.4. Isolation of Bacteriophages.	59
3.5. Detection of bacteriophages.	60
3.5.1. Plaque assay method	60
3.5.2. Bacteriophages purification and propagation	60
3.5.3. Preparation of high titer stock of the isolated bacteriophages	61
3.6. Characterization of isolated bacteriophages	61
3.6.1. Morphological characteristics (Electron microscopy)	62
3.6.2. Determination of host ranges and cross infectivity of the isol phages	
3.6.3 Adsorption assays.	62
3.6.4. One Single-step growth experiments.	63
3.6.5. Effect of different temperatures on the phage stability	64
3.6.6. Effect of pH on the phage stability	64

3.6.7. Effect of chloroform on phages	64
3.7. Molecular characteristics	64
3.7.1. Isolation and characterization of nucleic acids	64
3.7.2. Restriction digestion of isolated bacteriophages genomic DNA	65
3.8. Evaluation of the lytic activity of isolated phages against isopathogenic bacteria	
4. Results	66
4. Isolation and identification of possible pathogenic bacteria from b chickens.	
4.4.1 Salmonella isolation and identification	66
4.4.2 <i>E. coli</i> isolation and identification	67
4.4.3 <i>Proteus</i> isolation and identification	68
4.4.4 Staphylococcus aureus isolation and identification)	69
4.4.5 Bacillus cereus isolation and identification	69
4.4. Prevalence of pathogenic bacteria isolated from broiler chickens	70
4.3. Antimicrobial resistance patterns of the isolated bacteria	72
4.3. 1. Antimicrobial resistance patterns of <i>Salmonella</i> serovars	72
4.3.2 Antimicrobial resistance patterns of <i>E. coli</i> and <i>Proteus</i>	75
3.3.3 Antimicrobial resistance patterns of <i>S. aureus</i>	77
4.3.4 Antimicrobial resistance patterns of <i>B. cereus</i>	78
4.3. Molecular detection of some antimicrobial resistance genes	80
4.1. Detection of (mecA) gene Gene in S. aureus	82
4.5 Isolation of Bacteriophages	83
4.5.1. Characterization of Salmonella bacteriophages	87
4.5.2. Morphological characterization (Electron microscopy)	87
4.5.3. Host range of Salmonella phages	88
4.5.4. One step growth curve of the isolated phages	90