BIOLOGICAL AND MOLECULAR CHARACTERIZATION OF SOME TRICHODERMA ISOLATES WITH HIGHLY ANTIFUNGAL ACTIVITY

By

MOHAMED IBRAHIM ABD ELHAMID

B.Sc. Agric. Sci. (Agronomy), Fac. Agric., Cairo Univ., 2000 M. Sc. Agric. Sci. (Microbiology), Fac. Agric., Cairo Univ., 2010

THESIS

Submitted in Partial Fulfillment of the Requirements for the Degree of

DOCTOR OF PHILOSOPHY

In

Agricultural Sciences (Agricultural Microbiology)

Department of Agricultural Microbiology
Faculty of Agriculture
Cairo University
EGYPT

2018

Format reviewer

Vice Dean of Graduate Studies

APPROVAL SHEET

BIOLOGICAL AND MOLECULAR CHARACTERIZATION OF SOME TRICHODERMA ISOLATES WITH HIGHLY ANTIFUNGAL ACTIVITY

Ph.D. Thesis
In
Agric. Sci. (Agricultural Microbiology)

By

MOHAMED IBRAHIM ABD ELHAMID

B.Sc. Agric. Sci. (Agronomy), Fac. Agric., Cairo Univ., 2000 M. Sc. Agric. Sci. (Microbiology), Fac. Agric., Cairo Univ., 2010

Approval Committee

Dr. ELSHAHAT MOHAMED RAMADAN Professor of Microbiology, Fac. Agric., Ain Shams University.	••••
Dr. MOHAMED ABDELALIM ALIProfessor of Agricultural Microbiology, Fac. Agric., Cairo Unive	
Dr. MOHAMED ZAKARIA SEDIK	

Date: 26/9 /2018

SUPERVISION SHEET

BIOLOGICAL AND MOLECULAR CHARACTERIZATION OF SOME TRICHODERMA ISOLATES WITH HIGHLY ANTIFUNGAL ACTIVITY

Ph.D. Thesis
In
Agricultural Sci. (Agricultural Microbiology)

By

MOHAMED IBRAHIM ABD ELHAMID

B.Sc. Agric. Sci. (Agronomy), Fac. Agric., Cairo Univ., 2000 M. Sc. Agric. Sci. (Microbiology), Fac. Agric., Cairo Univ., 2010

SUPERVISION COMMITTEE

Dr. MOHAMED ZAKARIA SEDIK Professor of Agricultural Microbiology, Fac. Agric., Cairo University.

Dr. ISMAIL MOHAMED ISMAIL Senior Researcher of Microbiology, (AGERI), ARC., Giza. Name of Candidate: Mohamed Ibrahim Abd Elhamid Degree: Ph.D.

Title of Thesis: Biological and Molecular Characterization of Some

Trichoderma Isolates with Highly Antifungal Activity

Supervisors: Dr. Mohamed Zakaria Sedik

Dr. Ismail Mohamed Ismail

Department: Agricultural Microbiology **Approval:** 26 /9 /2018

ABSTRACT

Phytopathogen fungi are posing serious problems to the worldwide cultivation systems of economically important plants. Eco-friendly bio-control agents become necessary to help in resolving this problem. In this study, soil and rhizosphere samples were collected from five locations in Egypt. While forty three Trichoderma isolates were successfully isolated. Of these forty three fungi, fourteen isolates showed antifungal activity, six of them were belonged to Trichoderma harzianum. The other eight isolates included T. asperellum, T. viride, T. hamatum and T. lixii. The effect of identified isolates of Trichoderma spp. on radial growth and the percentage of inhibition toward the different fungal pathogens were studied in dual culture on Petri dishes. The molecular identification of the isolated *Trichoderma* spp. was done using two taxon-selective primers (ITS1 & ITS4) for the internal transcribed spacer (5.8S-ITS) region in the nuclear ribosomal repeat unit. The sequence BLAST analysis of the ITS sequences showed that the isolates were belonging to Trichoderma spp. The phylogenetic relationship was conducted between isolated sequences. Full length of chitinase gene (1.2 Kb) was isolated from two different species and comparatively analyzed between other isolates of high antifungal activity. Select the best medium in terms of growth spore production and biomass yield for the production of *Trichoderma* spp.

Keywords: *Trichoderma*, biocontrol agents, internal transcribed spacer (ITS), chitinase gene, antifungal activity.

DEDICATION

I dedicate this work to whom my heart felt thanks; to my father spirit, to my mother, sisters and my lovely kids (Mohamed and Aya) for support and encouragement they continually offered along the period of my graduation.

ACKNOWLEDGEMENT

At first, I would like to thank **ALLAH** that allowing me to achieve this work, without his bless any great effort is invaluable.

I wish to express my sincere thanks, deep gratitude and appreciation to **Dr. Mohamed Z. Sedik** Professor of Microbiology, Faculty of Agriculture, Cairo University for suggesting the problems, supervision, continued assistance, guidance, great interest, encouragement, following the progress of the work with great interest and continuous criticism through the course of study.

My deepest and sincere graduate to Ismail M. Ismail Senior Researcher of Microbiology, Agricultural Genetic Engineering Research Institute (AGERI), for directing, suggesting the point of research, supervision, continuous encouragement, continuous support for practical parts of this work and constant helping writing the paper and the thesis.

Many thanks are due to all the members of the Laboratory of Molecular Biology and Microbial Biotechnology (MBMB), AGERI for their continuous help, providing facilities and moral support.

CONTENTS

INTRODUCTION
REVIEW OF LITERATURE
1. Fungal diseases and their impact
2. Diseases resistance through genetic engineering
3. Fungal cell wall
4. Cell wall degrading enzymes
5. Mechanisms of biological control activity
a. Competition
b. Mycoparasitism
c. Induced resistance
d. Inactivation of the pathogen's enzymes
e. Secondary metabolites
6. Biodiversity and phylogeny of <i>Trichoderma</i>
7. Characteristics of <i>Trichoderma</i>
8. Trichoderma as industrial workhorses
a. Cellulases and plant cell wall-degrading enzymes
b. Heterologous protein production
c. Food industry
9. ITS as an environmental DNA barcode for fungi
MATERIALS AND METHODS
1. Materials
a. Biological material
b.Chemicals and research tools
c. Bacteriological media
d. Supplements
e. Vectors
f. SDS - PAGE solutions
g. DNA Solutions
h. Trituration Buffer
i. Instruments
j. Computer software
k. DNA Primers
2. Methods
a. Sample collection of <i>Trichoderma</i> from soil

b. Isolation of fungal species from soil
c. Identification of the isolated fungi
d. Screening of <i>Trichoderma</i> spp
e. Polymerase Chain Reaction (PCR)
f. Elution & Purification of PCR fragments
g. Cloning of chitinase PCR fragments
h. Plasmid DNA preparation
i. Electrophoresis of DNA4
j. Visualization and photography4
k. DNA digestion
1. DNA sequencing
m. SDS – PAGE analysis4
n. Electrophoresis of protein gel4
o. Staining and destaining of the protein gel4
p. Study mycelia growth on different culture media
q. Media for inoculums and fermentation
r. Initial inoculum for inoculation of fermenter4
RESULTS
1. Isolation of <i>Trichoderma</i> spp
2. Morphological Characterization 4
3. Antagonistic activity of <i>Trichoderma</i> in dual culture 4
4. Modes of action of <i>Trichoderma</i> spp 5
5. Determination of chitinase activity
6. Protein profile of <i>Trichoderma</i> spp 5.
7. PCR amplification of 5.8S-ITS region 5
8. Sequence analysis of 5.8S-ITS region
9. Phylogenetic analysis
10. Amplification of chitinase genes
11 Cloning of chitingse genes
12 Confirmation of clones
13. Amino acids analysis of chitinase
14. Study mycelia growth on different culture media
15. Media for inoculums and fermentation
DISCUSSION ⁷
SUMMARY ⁷
REFERENCES
ARABIC SUMMARY 8

LIST OF TABLES

No.	Title	Page
1.	Components volume of SDS-PAGE solutions	27
2.	Oligonucleotide sequence of specific primers	32
3.	The source of samples, location and the soil types	33
4.	Governorate and type of isolates	45
5.	Average inhibition percentage of mycelial growth	49
6.	BLAST search results for <i>Trichoderma</i> isolates	62

LIST OF FIGURES

No.	Title	Page
1.	The molecular structure of chitin	8
2.	The internal transcribed spacer (ITS) region	22
3.	Restriction map and general specification of pGEM	27
4.	Antifungal property of Trichoderma harzianum	51
5.	Antifungal property of Trichoderma asperellum	52
6.	Antifungal property of Trichoderma viride	53
7.	Antifungal property of Trichoderma lixii	54
8.	Antifungal property of Trichoderma hamatum	54
9.	Screening of Trichoderma harzianum for chitinase activity	56
10.	Screening of Trichoderma asperellum for chitinase activity	57
11.	Screening of <i>Trichoderma viride</i> for chitinase activity	57
12.	SDS-PAGE of total protein patterns <i>Trichoderma</i> isolates	59
13.	5.8S-ITS region amplified for all <i>Trichoderma</i> isolates	61
14.	Nucleotides sequence of 5.8S-ITS for <i>Trichoderma</i> isolates.	62
15.	Phylogenetic tree for all <i>Trichoderma</i> isolates	63
16.	PCR amplification of chitinase gene	64

17.	Schema showing cloning strategy of PCR amplified Chitinase coding sequence in TA vector	65
18.	Restriction digestion of recombinant clone using restriction enzyme	66
19.	Nucleotides sequence of chitinase gene	67
20.	Deduced amino acid sequence of chitinase gene	68
21.	Schema showing secondary and three dimensional structure of chitinase gene	69
22.	Comparison of percentages of deduced amino acids between chitinase genes	70

INTRODUCTION

One of the ultimate aims of agricultural biotechnology is to feed an expanding world population. At present, world is facing the twin problem of increasing population and decreasing available land for agriculture. The world population has increased by 90% in the past 40 years while food production has increased by only 25% per head. With an additional 1.5 billion mouths to feed by 2020, farmers worldwide will have to produce 39% more grain (Anonymous, 2000).

In agriculture, worldwide, parasites and pathogens of plants are threat to crop production (Sarah and Paul, 2005). An estimated 37% crop loss is due to pests, of which 12% is due to pathogens. The extensive use of fungicides in various parts of the world for years has increased the pollution level in soil and water, and adverse effect on food quality and human health. Apart from this, the chemicals tend to become less efficient due to the development of resistance among the pathogens over time. Hence, it is necessary to look for alternative disease management practices, which include the use of eco-friendly biological control agents and pathogen-resistant crop cultivars.

Biological control means control of disease through some biological agency that is any living microorganism. Biocontrol may be defined as any condition or practice where by survival or activity of pathogen is reduced through the agency of any other living organism with the result that there is reduction in the incidence of disease caused by the pathogen. Biocontrol of soil-borne plant pathogens affecting agricultural plants can be controlled by the use of species of Trichoderma, Aspergillus and Trichothecium. There are some antagonistic bacteria like Bacillus subtilis, Enterobacter aerogenes, Pseudomonas fluorescence and Streptomyces spp. in disease control.

The specific need of complete disease control, which the biological agent seldom offers due to the problems associated with distribution of pathogen propagates in soil. There is a growing concern in recent years, both in developed and developing countries, about the use of hazardous fungicides for controlling plant diseases. Chemical pesticides have already been proven to cause adverse environmental effects and result in health hazards to human as well as other organisms including beneficial natural enemies. So there is need to develop safer and environmentally feasible control alternatives. Biological control, *i.e.*, the use of biological processes to lower inoculum density of the pathogen in order to reduce the disease producing activities thereby reducing crop loss, is a potential nonhazardous alternative (Chet, 1990).

Among different biological control agents, *Trichoderma* spp. has proved effective and selective enough against most of the fungal diseases. *Trichoderma* spp. has evolved numerous mechanisms that are involved in attacking other fungi. These mechanisms include competition for space and nutrients (Elad *et al.*, 1999), mycoparasitism (Haran *et al.*, 1996 and Lorito *et al.*, 1996), production of inhibitory compounds (Sivasithamparam and Ghisalberti, 1998), inactivation of the pathogen's enzymes and induced resistance (Kapulnik and Chet, 2000). Today, more than 50 different *Trichoderma*-based agriculture products can be found as registered in many countries in five continents, and are sold and applied to protect and improve yield of

vegetables, ornamentals and fruit trees. *Trichoderma* is completely safe and in 55 years of research there has never been a recorded adverse reaction on humans and livestock (Lorito, 2005).

As the skeleton of the fungal cell wall mainly contains chitin, glucan and proteins where mycoparasitism and enzymes that hydrolyze these components are one of the main mechanisms accounting for showing antagonistic activity against plant pathogenic fungi. Chitinase, β -1, 3-glucanase and cellulase are important in the hyper-parasitic mechanism. The distribution of chitinases in nature is very common. It has been established that the chitinase producing *Trichoderma* spp. can be effective biocontrol agents against fungal pathogens (Kubicek *et al.*, 2001).

The ultimate goal of this study was isolation, purification and characterization of some local *Trichoderma* isolates conferring antifungal activity conducted to investigate and characterize some environemently friendly alternatives to protect local crops from fungal pathogenes and, to a lesser extent, fungal pests.

The aims of this work were:

- 1. Isolation and screening of *Trichoderma* isolates from different soil samples.
- 2. Screening the effects of *Trichoderma* isolates with high activity against selected important fungal pests.
- 3. Identification of *Trichoderma* isolates on the basis of morphological features, pathogenecity test and biochemical characteristics.

- 4. Identification *Trichoderma* isolates using ITS (Internal transcribed spacer) gene, Cloning and nucleotide sequence analysis of ITS gene.
- 5. Design of oligonucleotide sequences of specific primers to isolate of antifungal gene based on highly conserved domains of different antifungal gene available in nucleotide sequence database.
- 6. Cloning and nucleotide sequence analysis of antifungal coding sequence.
 - 7. Study the chitinase activities of local *Trichoderma* isolates.
- 8. Finding the best solutions for economic production of the *Trichoderma* biomass.