Screening of GATA-1 Mutation in Patients with Down Syndrome

Thesis

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By

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List of Abbreviations

Abb.	Full term
AFP	Alpha fetoprotein
AMKL	Acute megakaryocytic leukemia
APP	β-amyloid precursor protein
ASD	Atrial septal defect
AVSD	Atrio-ventricular septal defect
Αβ	β-amyloid peptide
CBC	complete blood count
CEP	Congenital erythropoietic porphyria
CH	Congenital hypothyroidism
CHD	Congenital heart disease
CRP	C-reactive protein
DNA	Deoxyribo-nucleic acid
DS	Down syndrome
DSCR	Down syndrome critical region
FISH	Flourescent in situ hybridization
GI	Gastrointestinal
GIT	Gastrointestinal tract
HPLC	high performance liquid chromatography
HSCs	Hematopoietic stem cells

List of Abbreviations

Abb.	Full term
Mb	Megabase
MEP	Megakaryocyte-erythrocyte progenitor
ML-DS	Myeloid leukemia in Down syndrome
MPNs	Myeloproliferative neoplasms
MI	meiosis I
МП	Meiosis Π
NT	Nuchal translucency
PAD	Patent ATRIAL duct
PCR	POLYMERASE chain reaction
PT21	Partial trisomy 21
RNA	Ribonucleic acid
Ss	Sanger sequencing
TAM	Transient abnormal myelopoiesis
TMD	Transient myeloproliferative disease
uE3	Unconjugated estriol
VSD	\$ventricular septal defect
WHO	World health organization
XLT	X-linked thrombocytopenia
XLTT	X-linked thrombocytopenia with thalassemiA
β-hCG	Human chorionic gonadotrophins

Introduction

In children with Down syndrome (DS), the risk of developing acute megakaryocytic leukemia (AMKI) is estimated to be 500 times higher than in children without DS (kanezaki et al., 2010).

Interestingly, neonates with DS are at a high risk 5-10% of developing a hematologic disorder referred to as transient myeloproliferative disease (TMD) (*Hitzler*, 2007). The World Health Organization (WHO) defines TMD as increased peripheral blood blast cells in neonates with DS (*Vardiman et al.*, 2009).

In about 60% of cases, TMD resolves spontaneously within the first 3 months of life (*Massey et al., 2006*). A small proportion of babies with TAM will die from their disease, usually due to liver failure caused by hepatic fibrosis and blast cell infiltration (*Klusmann et al, 2008; Gamis et al, 2011*).

An estimated 20% to 30% of babies with TMD subsequently develop MI-DS (Myeloid leukemia in DS). Thus, TMD is an important clinical problem (*Klusmann et al.*, 2008).

Acquired mutations in exon 2 of the hematopoietic transcription factor *GATA-1* mapped at Xp11.23 are consistently present in the affected cells of children with TMD and MI-DS, leading to expression of N-terminally truncated GATA-1 protein *(Cabelof et al., 2009)*.



GATA-

1 is a transcription factor that comprehensively regulates the gen es that are important for the development of erythroid and megak aryocytic cells. Accumulating evidence supports the notion that d efects in GATA-

1 function are intimately linked to hematopoietic disorders (Shim ızu and Yamamoto, 2012).

GATA-

1 mutation analysis showed that 8.5% of DS neonates had a GA

I mutation detected by Sanger sequencing/denaturing high perfo rmance liquid chromatography (Ss/DHPIC) (Roberts et al., 2013) . Similar to estimates from retrospective studies (5%-10%) (Malinge et al., 2009).

Xu et al. (2003) found the presence of GATA-1 mutations in 21 patients out of 22 patients Down syndrome with TMD and in 12 patients out of 18 patients Down syndr ome with AMKI.

Roberts et al. (2013) suggest that a practical and sensiti ve definition of TMD is the presence of blasts >10% on bloo d smears and a GATA-1 mutation detected by Ss/DHPIC.

AIM OF THE WORK

The aim of this study is to screen DS neonates with peripheral blasts for GATA-1 mutations. This facilitates regular clinical and laboratory follow up and ensures appropriate management of cytopenias that may precede AMI, including the timing of antileukemic therapy, any possibility of use of GATA-1 as a marker for cure.



Review of Literature -

Chapter 1

DOWN SYNDROME

Introduction

Trisomy for human chromosome 21 is the most frequent I iveborn aneuploidy and results in Down syndrome (DS). This is a well recognized syndrome with variable phenotypic expression (Gardiner et al., 2010).

The incidence of trisomy 21 is influenced by maternal age (*Wiesman et al., 2009*), It occurs in one in approximately 691 and 1000 newborns in the USA and Europe, respectively (*Jiang et al., 2015*).

Genotype of DS

Chromosome 21 is the smallest human autosome. It consists of ~ 50 Mb of DNA. The short arm is very small and a ll of the unique genes that have been located to this chromosome have been mapped to the long arm of the chromosome as shown in figure 1 (Kola and Hertzog, 1997).

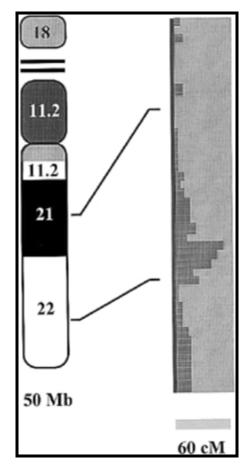


figure (1): Map of the frequency of expressed sequences encoded on different parts of human chromosome 21 (Kola and Hertzog, 1997).

The smallest chromosomal region in common among in dividuals who share a given feature is referred to as a 'Down syndrome critical region' (DSCR). The best-

defined DSCR extends \sim 5 Mb from D21S17 to MX1 in band 21q22.3. This segment contains about 33 conserved genes (O lson et al., 2007).

Mechanisms of DS

1. Nondisjunction:

Nondisjunction occurs in about 95% of people with Down syndrome (*Shin et al.*, 2010).

Nondisjunction is the failure of homologous chromoso mes or sister chromatids to segregate to separate daughter cells during cellular division. When this type of error occurs during meiosis, some of the resulting gametes will have too many or too few chromatids compared with the expected haploid number (aneuploidy) (Middlebrooks et al., 2014).

In~90% of trisomy 21 individuals, the additional chro mosome is maternal in origin, \sim 70% of the maternal errors h ave been found to occur during meiosis I (MI), while the oth er 30% occur during meiosis II (MII) (lamb et al., 1997).

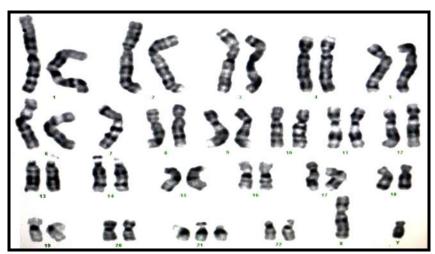


figure (2): Karyotype of a patient with nondisjunction Down syndrome: 47, XY, +21. (Genetics Unit, Ain Shams University)

2. Translocation:

This type accounts for a small percentage of people with Down syndrome (about 3%) (*Shin et al., 2010*).

There is extra chromosome 21 material attached (transloc ated) onto another chromosome. For parents of a child with Do wn syndrome due to a translocation, there may be an increased chance of Down syndrome in future pregnancies. This is becaus e one of the two parents may be a carrier of a balanced transloc ation *(GHR, 2012)*.



figure (3): Karyotype of a patient with translocation Down syndrome: 46, XX, der (14;21), +21 *(Genetics Unit, Ain Shams University).*