

The Potential Hepatoprotective Effect of Quercetin on Cholestatic Liver Injury in Rats

Thesis

*Submitted For Partial Fulfillment of Master Degree in
Basic Medical Science (Physiology)*

Presented by

Gehad Salah Abdel Hamid

M.B.B.Ch.

Faculty of Medicine- Ain Shams University

Under Supervision of

Prof. Dr. Ebtessam Ahmed Abou Shadi

Professor of Physiology

Faculty of Medicine- Ain Shams University

Prof. Dr. Sahar Mohammad El-Agaty

Professor of Physiology

Faculty of Medicine- Ain Shams University

Dr. Noha Abdel-Aziz Nassef

Assistant Professor of Physiology

Faculty of Medicine- Ain Shams University

**Department of Medical Physiology
Faculty of Medicine - Ain Shams University
2019**

بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

□

رَبِّ أَوْزَعْنِي أَنْ أَشْكُرَ نِعْمَتَكَ الَّتِي

أَنْعَمْتَ عَلَيَّ وَعَلَى وَالِدَيَّ وَأَنْ

أَعْمَلَ صَالِحًا تَرْضَاهُ وَأَصْلِحْ لِي فِي

ذُرِّيَّتِي إِنِّي تُبْتُ إِلَيْكَ وَإِنِّي مِنَ

الْمُسْلِمِينَ

□



Acknowledgement

First and foremost, Thanks to **Allah**, to whom I relate any success in achieving any work in my life.

My deep appreciation to **Dr. Ebteessam Abou Shadi**, Professor of Physiology, Ain Shams University for her valuable instructions, unlimited help and great deal of support, her endless patience with me and for her experienced guidance and helpful suggestions that make the completion of this work possible.

I owe special feeling of gratitude to **Dr. Sahar Mohammad El-Agaty**, Professor of Physiology, Ain Shams University for her great help, close supervision, wise opinions, guidance and her continuous encouragement and for her precious effort. Without her support, this work would not have been completed.

I would like to express my deep gratitude and sincere appreciation to **Dr. Noha Abdel-Aziz Nassef**, Assistant Professor of Physiology, Ain Shams University for her sustained support, continued encouragement and for her precious time and effort that made this thesis possible. It was great honor to me to do this thesis under her supervision.

I am also grateful to **Prof. Dr. Bataa Mohammad El-Kafoury**, Professor and Head of Physiology Department, Faculty of Medicine- Ain Shams University, for her unique effort, considerable help throughout this work.

I am also grateful to **Dr. Sayed Abdel Raheem**, Assistant professor of Histopathology, Faculty of Medicine, Al Azhar University, for his contribution in the histological studies.

Last but not least, I dedicate this work to **My Mother and My Husband**, whom without their sincere emotional support, pushing me forward this work would not have ever been completed.

 **Gehad Salah**

Contents

Subject	Page No.
List of Abbreviations	I
List of Tables	IV
List of Figures	VII
Introduction	1
Aim of the Work	4
Review of Literature	5
1. Cholestatic liver injury.....	6
2. Pathophysiology of cholestatic liver injury	8
3. Clinical manifestations.....	13
4. Bile duct ligation as an experimental model of cholestatic liver	15
5. Therapeutic approaches for cholestatic liver disease	16
6. Flavonoids as potential therapeutic agents (Quercetin).....	20
Materials and Methods	25
Results	56
Discussion	89
Summary and Conclusion	101
References	105
Arabic Summary	١

List of Abbreviations

Abb.	Full Term
%BW	Percentage of body weight gain
ABC	ATP-binding cassette transporters
ALP	Alkaline phosphatase
ALT	Alanine aminotransferase
ANOVA	Analysis of variance
AST	Aspartate aminotransferase
BDL	Bile duct ligated-group
BDL-Q	Bile duct ligated quercetin-treated group
BSEP	Bile salt export pump
BW_f	Final body weights
BW_i	Initial body weights
CD18	Cluster of differentiation 18
COX	Cyclooxygenase
DAMPs	Damage associated molecular pattern molecules
ECM	Extracellular matrix
EGFR	Epidermal growth factor receptor
Egr-1	Early growth response factor 1

Abb.	Full Term
GPX	Glutathione peroxidase
GR	Glutathione reductase
GSH	Reduced glutathione
GSSG	Oxidized glutathione
H&E	Hematoxylin & Eosin
HSC	Hepatic stellate cell
ICAM1	Intracellular adhesion molecule 1
LOX	Lipoxygenase
LPS	Lipopolysaccharide
LW	Liver weight
LW%	Liver index
MAPK	Mitogen-activated protein kinase
MPO	Myeloperoxidase
MPT	Mitochondrial permeability transition pore
MRP-2	Multi-drug resistant related protein 2
MTC	Masson's trichrome
NF-κB	Nuclear factor kappa
NO	Nitric oxide
NTCP	Na ⁺ dependent taurocholate transporter
O.D.	Optical density

Abb.	Full Term
OATP2	Organic anion transporter
PDGF	Platelet derived growth factor
ROS	Reactive oxygen species
SAP	Serum amyloid P
SHAM	Sham-operated
SPSS	Statistical Program for Social Science
SW	Spleen weight
SW%	Spleen index
TGF-β1	Transforming growth factor- beta 1
TIMP1	Tissue inhibitor of metalloproteinase 1
TNF-α	Tumor necrosis factor- alpha
TP	Total proteins
UDCA	Ursodeoxycholic acid
α-SMA	Alpha smooth muscle antigen

List of Tables

<i>Table</i>	<i>Title</i>	<i>Page</i>
1	Changes in initial body weight (BW _i), final body weight (BW _f), percentage of body weight gain (% BW), liver weight (LW), liver index (LW%), spleen weight (SW) and spleen index (SW%), in the three study groups	58
2	Changes in serum levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP) and total proteins (TP) in the three study groups	63
3	Changes in liver tissue levels of myeloperoxidase (MPO), glutathione peroxidase (GPX), tumor necrosis factor alpha (TNF- α) and transforming growth factor beta 1 (TGF- β 1), in the three study groups	68
4	Changes in the percentage area of collagen fibers in the three study groups	73
5	Initial body weight (BW _i), final body weight (BW _f), percentage of body weight gain (BW %), liver weight (LW), liver index (LW %), spleen weight (SW) and spleen index (SW %) in sham - operated group	80

<i>Table</i>	<i>Title</i>	<i>Page</i>
6	Initial body weight (BW _i), final body weight (BW _f), percentage of body weight gain (BW %), liver weight (LW), liver index (LW %) spleen weight (SW) and spleen index (SW %) in bile duct-ligated group	81
7	Initial body weight (BW _i), final body weight (BW _f), percentage of body weight gain (BW %), liver weight (LW), liver index (LW %), spleen weight (SW) and spleen index (SW %), in bile duct ligated quercetin- treated group	82
8	Serum levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP) and total proteins (TP), in sham- operated group	83
9	Serum levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP) and total proteins (TP) in bile duct-ligated group	84
10	Serum levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP) and total proteins (TP) in bile duct ligated quercetin-treated group	85

<i>Table</i>	<i>Title</i>	<i>Page</i>
11	Liver tissue levels of myeloperoxidase (MPO), glutathione peroxidase (GPX), tumor necrosis factor alpha (TNF- α) and transforming growth factor beta 1(TGF- β 1) in sham-operated group	86
12	Liver tissue levels of myeloperoxidase (MPO), glutathione peroxidase (GPX), tumor necrosis factor alpha (TNF- α) and transforming growth factor beta 1(TGF- β 1) in bile duct-ligated group	87
13	Liver tissue levels of myeloperoxidase (MPO), glutathione peroxidase (GPX), tumor necrosis factor alpha (TNF- α) and transforming growth factor beta 1(TGF- β 1) in bile duct ligated quercetin-treated group	88

List of Figures

<i>Figure</i>	<i>Title</i>	<i>Page</i>
1	The chemical structure of quercetin	21
2	Aspartate aminotransferase standard curve	31
3	Alanine aminotransferase standard curve	33
4	Changes in initial body weight (BW _i), final body weight (BW _f) and percentage of body weight gain (%BW) in the three study groups	59
5	Changes in liver weight (LW) and liver index (LW%) in the three study groups	60
6	Changes in spleen weight (SW) and spleen index (SW%) in the three study groups	61
7	Changes in serum levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT) in the three study groups	64
8	Changes in serum levels of alkaline phosphatase (ALP) and total proteins (TP) in the three study groups	65
9	Changes in liver tissue levels of myeloperoxidase (MPO), glutathione peroxidase (GPX) in the three study groups	69

<i>Figure</i>	<i>Title</i>	<i>Page</i>
10	Changes in liver tissue levels of tumor necrosis factor alpha (TNF- α), transforming growth factor beta 1(TGF- β 1) in the three study groups	70
11	Changes in the percentage area of collagen fibers in the three study groups	74
12	Photomicrograph of H & E (X200) stained liver sections in the three study groups	75
13	Photomicrograph of H & E (X400) stained liver sections in the three study groups	76
14	Another photomicrograph of H & E (X400) stained liver sections in the three study groups	77
15	Photomicrograph of Masson trichrome (MTC) (X200) stained liver sections in the three study groups	78
16	Photomicrograph of Masson trichrome (MTC) (X400) stained liver sections in the three study groups	79

The Potential Hepatoprotective Effect of Quercetin on Cholestatic Liver Injury in Rats

Abstract

Background: cholestasis is a prevalent health problem associated with liver oxidative stress, inflammation, and fibrosis. Quercetin has been shown to afford a beneficial effect in a variety of liver diseases.

Aim: This study was designed to investigate the potential protective effect of quercetin on liver cholestasis and the possible underlying mechanisms in a rat model of bile duct ligation (BDL).

Design: Experimental study

Methods: This study was carried out on adult male Wister rats which were randomly divided into: Sham, BDL and BDL- quercetin treated (BDL- Q) groups. Quercetin was given by gavage in a dose of 50 mg/kg/day.

Results: Bile duct ligation resulted in a significant increase in serum levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), and liver levels of myeloperoxidase (MPO), tumor necrosis factor alpha (TNF- α), and transforming growth factor beta 1 (TGF- β 1), along with a significant decrease in serum levels of total proteins (TP) and liver glutathione peroxidase (GPX) in BDL group versus sham group. Quercetin treatment significantly lowered serum levels of AST, ALT, ALP, and MPO, TNF- α , and TGF- β 1 in liver tissues associated with a significant increase in serum TP and liver GPX in BDL-Q group versus BDL rats. Histological studies revealed enhancement of inflammation and a significant increase in the percentage area of collagen deposition in BDL versus sham group. These changes were attenuated in BDL-Q group compared to BDL rats.

Conclusions:

Quercetin alleviated cholestasis induced liver injury and improved liver function possibly via attenuating liver oxidative stress, inflammation and fibrosis.

Key words: Cholestasis, quercetin, oxidative stress, inflammation, fibrosis

Introduction

Chronic cholestasis is a prevalent health problem produced by impairment of bile flow, and accumulation of toxic bile acids in hepatocytes, evoking severe liver injury which may progress to cirrhosis and liver failure (**Hirschfield et al., 2010**). It is caused by various aetiologies such as primary biliary cirrhosis, primary sclerosing cholangitis and progressive familial intrahepatic cholestasis (**Ghonem et al., 2015**). Ligation of the common bile duct in rodents represents a well settled experimental model of cholestasis that initiates a complex cascade of pathological events similar to that of human (**Dondorf et al., 2017**). This model enabled the scientists to recognize the pathophysiology of the cholestasis and to develop different treatments to human cholestatic liver diseases.

Recently, evidence has accumulated that the progression of liver injury in cholestasis is heavily dependent on overproduction of reactive oxygen species (ROS) as well as pro-inflammatory cytokines (**Gonzalez-Sanchez et al., 2015**). Exposure of hepatocytes to high concentrations of potentially toxic bile acids initiates hepatocellular injury, followed by a leukocytic phase in which activated neutrophils infiltrate and attack the bile

acid stressed hepatocytes through ROS formation (*Rust et al., 2009; Copple et al., 2010*). Excessive production of ROS has been demonstrated to induce cellular damage (*Cesaratto et al., 2004*) and promote inflammation by upregulating tumor necrosis factor-alpha (TNF- α) signaling pathway, and IL-6 mRNA expression in chronic liver diseases (*Elsharkawy et al., 2005; Ghatak et al., 2011*). Transforming growth factor beta-1(TGF- β 1), a potent profibrotic cytokine, was also activated by oxidative stress (*Poynard et al., 2010*) and in a consequence stimulates the transformation of hepatic stellate cells into myofibroblasts which increase the extracellular matrix formation, promoting liver fibrosis (*Elsharkawy et al., 2005*). During the past decades, the mechanisms of liver cholestasis have been investigated, but few therapeutic strategies are available to efficiently interrupt the progression of liver injury. According to the aforementioned reports, oxidative stress, and inflammation are potential targets for therapeutic intervention in treating cholestatic liver disorders. Therefore, antioxidants might have a relevant benefit and restrain the hepatocellular injury in chronic cholestasis.

Quercetin (3,3',4',5,6-Pentahydroxyflavon), a natural flavonoid compound found in vegetables and fruits, has a