

# **Comparative Analysis of Different Activation Methods on Platelet Rich Plasma**

## ***Thesis***

Submitted in Partial Fulfillment for the Master Degree  
in Clinical Pathology

## ***Presented by***

**Noha Mohamed Ahmed El Shishtawy**

M.M., B.Ch

Haematology Resident, Theodor Bilharz Research Institute

## ***Supervised by***

**Prof. Dr./ Ibrahim Youssef Abdel-Messih**

Professor of Clinical Pathology

Faculty of Medicine, Ain Shams University

**Prof. Dr./ Iman Abd El Aziz Khaled**

Professor of Clinical Pathology

Haematology Department

Theodor Bilharz Research Institute

**Assist. Prof. / Mohamed Tarif Mohamed Hamza**

Assist. Professor of Clinical Pathology

Faculty of Medicine, Ain Shams University

Faculty of Medicine

Ain Shams University

**2019**



## Acknowledgments

First and foremost, thanks to **Allah**; to whom I relate any success in achieving any work in my life.

Words stand short when they come to express my gratefulness to my supervisors. I wish to express my deepest gratitude to **Professor Dr. Ibrahim Youssef Abdel-Messih**, Professor of Clinical Pathology, Faculty of Medicine, Ain Shams University, for his effective guidance, unlimited help and support. No words of gratitude would be enough for him. I really have the honor to complete this work under his supervision.

I wish to express my deep appreciation and profound gratitude to **Professor Dr. Iman Abd El Aziz Khaled**, Professor of Clinical and Chemical Pathology, Theodor Bilharz Research Institute, for her indispensable guidance, instructional supervision, continuous support, unlimited help, unlimited patience and close supervision throughout the entire work.

I am greatly honored to express my sincere gratitude and deep appreciation to **Assistant Professor Dr. Mohamed Tarif Mohamed Hamza**, Assistant Professor of Clinical Pathology, Faculty of Medicine, Ain Shams University, for his help and support.

I can't forget to thank with all appreciation **Prof. Dr. Salwa Toema**, Professor of Clinical Pathology, Theodor Bilharz Research Institute, for her encouragement and support.

I would like to thank all the staff members and colleagues of the Haematology Department, **Theodor Bilharz Research Institute** who supported me a lot throughout this entire work.

Last but not least, my deepest appreciation is expressed to all members in my family, especially my **Mother** and **Father** for their co-operation and encouragement.

✍ **Noha M. El-Shishtawy**

**2019**

## **List of Contents**

| <b>Subject</b>                      | <b>Page No.</b> |
|-------------------------------------|-----------------|
| <b>List of Abbreviations .....</b>  | <b>i</b>        |
| <b>List of Tables .....</b>         | <b>iv</b>       |
| <b>List of Figures .....</b>        | <b>v</b>        |
| <b>Introduction .....</b>           | <b>1</b>        |
| <b>Aim of the Work.....</b>         | <b>5</b>        |
| <b>Review of Literature</b>         |                 |
| Platelets .....                     | 6               |
| Platelet Rich Plasma.....           | 30              |
| Platelet Activation.....            | 47              |
| Platelets and Liver.....            | 69              |
| <b>Subjects and Methods .....</b>   | <b>74</b>       |
| <b>Results.....</b>                 | <b>79</b>       |
| <b>Discussion .....</b>             | <b>88</b>       |
| <b>Summary and Conclusion .....</b> | <b>94</b>       |
| <b>Recommendations .....</b>        | <b>98</b>       |
| <b>References .....</b>             | <b>99</b>       |
| <b>Arabic Summary .....</b>         | <b>—</b>        |

---

## List of Abbreviations

---

**Abbrev. Full-term**

---

|                       |                                                  |
|-----------------------|--------------------------------------------------|
| <b>ADP</b>            | : Adenosine diphosphate                          |
| <b>ADSC</b>           | : Adipose tissue derived stem cells              |
| <b>ANOVA</b>          | : Analysis of variance                           |
| <b>bFGF</b>           | : Basic fibroblast growth factor                 |
| <b>BMSC</b>           | : Bone marrow stem cells                         |
| <b>CBD</b>            | : Carboxy-terminal cell binding domain           |
| <b>CIB</b>            | : Calcium- and integrin-binding protein          |
| <b>DAG</b>            | : Diacylglycerol                                 |
| <b>DTS</b>            | : Dense tubular system                           |
| <b>ECM</b>            | : Extracellular matrix                           |
| <b>EDTA</b>           | : Ethylenediaminetetraacetic acid                |
| <b>EGF</b>            | : Epidermal growth factor                        |
| <b>ELISA</b>          | : Enzyme-linked immunosorbent assay              |
| <b>ERK</b>            | : Extracellular signal-regulated kinase          |
| <b>Fg</b>             | : Fibrinogen                                     |
| <b>Fn</b>             | : Fibronectin                                    |
| <b>GMP</b>            | : Guanosine monophosphate                        |
| <b>GP</b>             | : Glycoprotein                                   |
| <b>GPCR</b>           | : G protein-coupled receptor                     |
| <b>GRB</b>            | : Growth factor receptor bound                   |
| <b>HGF</b>            | : Hepatocyte growth factor                       |
| <b>HSC</b>            | : Hematopoietic stem cell                        |
| <b>IAP</b>            | : Integrin-associated protein                    |
| <b>ICAM</b>           | : Intercellular adhesion molecule                |
| <b>IGF</b>            | : Insulin growth factor                          |
| <b>IGFBP</b>          | : Insulin growth factor binding proteins         |
| <b>IL</b>             | : Interleukin                                    |
| <b>IP<sub>3</sub></b> | : Inositol triphosphate                          |
| <b>ITAM</b>           | : Immunoreceptor tyrosine-based activatory motif |
| <b>ITIM</b>           | : Immunoreceptor tyrosine-based inhibitory motif |

|               |                                                                      |
|---------------|----------------------------------------------------------------------|
| <b>LAMP</b>   | : Lysosomal-associated membrane protein                              |
| <b>LIBS</b>   | : Ligand-induced binding site                                        |
| <b>LSECs</b>  | : Liver sinusoidal endothelial cells                                 |
| <b>MAP</b>    | : Mitogen-activated protein                                          |
| <b>MAPK</b>   | : Mitogen-activated protein kinase                                   |
| <b>MCP</b>    | : Monocyte chemotactic protein                                       |
| <b>MK</b>     | : Megakaryocyte                                                      |
| <b>MPT</b>    | : Mitochondrial permeability transition                              |
| <b>MSC</b>    | : Mesenchymal stem cells                                             |
| <b>MVBs</b>   | : Multivesicular bodies                                              |
| <b>NBEAL2</b> | : Neurobeachin-like 2                                                |
| <b>OCS</b>    | : open canalicular system                                            |
| <b>PAC</b>    | : Procaspase-activating compound                                     |
| <b>PADGEM</b> | : Platelet activation dependent granule-external<br>membrane protein |
| <b>PAF</b>    | : Platelet activating factor                                         |
| <b>PAI-1</b>  | : Plasminogen activator inhibitor type I                             |
| <b>PAR</b>    | : Protease activated receptor                                        |
| <b>PDGF</b>   | : Platelet Derived Growth Factor                                     |
| <b>PDI</b>    | : Protein disulfide isomerase                                        |
| <b>PECAM</b>  | : Platelet endothelial cell adhesion molecule                        |
| <b>PF4</b>    | : Platelet factor 4                                                  |
| <b>PG</b>     | : Prostaglandin                                                      |
| <b>PI3K</b>   | : Phosphatidylinositol 3-kinase                                      |
| <b>PKG</b>    | : Protein kinase G                                                   |
| <b>PRGF</b>   | : Plasma rich in growth factors                                      |
| <b>PRP</b>    | : Platelet Rich Plasma                                               |
| <b>PS</b>     | : Phosphatidylserine                                                 |
| <b>PtdIns</b> | : phosphatidylinositol                                               |
| <b>PTKs</b>   | : Protein tyrosine kinases                                           |
| <b>PTPs</b>   | : Protein-tyrosine phosphatases                                      |
| <b>RANKL</b>  | : Receptor-activator of nuclear factor kappa beta<br>ligand          |
| <b>ROS</b>    | : Reactive oxygen species                                            |
| <b>S1P</b>    | : Sphingosine-1-phosphate                                            |

|                        |                                                                    |
|------------------------|--------------------------------------------------------------------|
| <b>SCF</b>             | : Stem cell factor                                                 |
| <b>SD</b>              | : Standard deviation                                               |
| <b>SDF</b>             | : Stromal cell-derived factor                                      |
| <b>SNAP</b>            | : Soluble N-ethylmaleimide-sensitive factor<br>attachment protein  |
| <b>SNARE</b>           | : Soluble N-ethylmaleimide-sensitive factor<br>attachment receptor |
| <b>SOCS</b>            | : Suppressors of cytokine signalling                               |
| <b>STAT</b>            | : Signal transducer and activator of transcription                 |
| <b>STEM</b>            | : Scanning transmission electron microscopy                        |
| <b>TFPI</b>            | : Tissue factor pathway inhibitor                                  |
| <b>TGFβ</b>            | : Transforming growth factor beta                                  |
| <b>TIMPs</b>           | : Tissue inhibitors of metalloproteases                            |
| <b>TLR</b>             | : Toll like receptor                                               |
| <b>TMEM</b>            | : Transmembrane protein                                            |
| <b>TNF</b>             | : Tumor necrosis factor                                            |
| <b>TPO</b>             | : Thrombopoietin                                                   |
| <b>TSP</b>             | : Thrombospondin                                                   |
| <b>TXA2</b>            | : Thromboxane A2                                                   |
| <b>TXA<sub>2</sub></b> | : Thromboxane A <sub>2</sub>                                       |
| <b>TXR</b>             | : Thromboxane A <sub>2</sub> receptor                              |
| <b>VAMP</b>            | : Vesicle- associated membrane proteins                            |
| <b>VAMP</b>            | : Vesicle-associated membrane protein                              |
| <b>VEGF</b>            | : Vascular endothelial growth factor                               |
| <b>Vn</b>              | : Vitronectin                                                      |
| <b>vWF</b>             | : Von Willibrand factor                                            |

## List of Tables

| <i>Table No.</i>  | <i>Title</i>                                                                             | <i>Page No.</i> |
|-------------------|------------------------------------------------------------------------------------------|-----------------|
| <b>Table (1):</b> | Contents of $\alpha$ granules .....                                                      | 12              |
| <b>Table (2):</b> | Synopsis of growth factors contained in platelet rich plasma .....                       | 16              |
| <b>Table (3):</b> | Miscellaneous Clinical Applications For Prp .....                                        | 45              |
| <b>Table (4):</b> | Laboratory results for participants .....                                                | 80              |
| <b>Table (5):</b> | PDGF-AB comparison between different activators of PRP ( <i>P</i> value) .....           | 83              |
| <b>Table (6):</b> | TGF- $\beta$ comparison between different activators of PRP ( <i>P</i> value).....       | 86              |
| <b>Table (7):</b> | Comparison between mean $\pm$ SE of different activators of PRP with resting group ..... | 87              |

## List of Figures

| <i>Figure No.</i>   | <i>Title</i>                                                                                                    | <i>Page No.</i> |
|---------------------|-----------------------------------------------------------------------------------------------------------------|-----------------|
| <b>Figure (1):</b>  | Diagrammatic representation of coronal and longitudinal section of the platelet.....                            | 7               |
| <b>Figure (2):</b>  | Platelet degranulation and action of the released cytokines in the process of formation of new bone tissue..... | 11              |
| <b>Figure (3):</b>  | Cell proliferation triangle, fundamental for tissue engineering. Source adapted from Crane and Everts.....      | 31              |
| <b>Figure (4):</b>  | Stem cell can replicate and differentiate.....                                                                  | 35              |
| <b>Figure (5):</b>  | Stem cells differentiate into different tissues .....                                                           | 35              |
| <b>Figure (6):</b>  | Preparation of PRP using 2 centrifugations.....                                                                 | 37              |
| <b>Figure (7):</b>  | Some clinical applications of PRP. In osteoarthritis (A) and in alopecia areata (B).....                        | 44              |
| <b>Figure (8):</b>  | Platelet activation responses .....                                                                             | 50              |
| <b>Figure (9):</b>  | Structure of the liver lobule (A) and the liver sinusoid (B).....                                               | 70              |
| <b>Figure (10):</b> | PDGF-AB comparison between different activators of PRP .....                                                    | 82              |
| <b>Figure (11):</b> | TGF- $\beta$ comparison between different activators of PRP .....                                               | 85              |

## Introduction

Platelet Rich Plasma (PRP) has been a breakthrough in the stimulation and acceleration of healing for over 20 years, starting by tissue repair in dentistry and expanding its clinical applications to other fields afterwards **(Marx et al., 1998)**. It represents a relatively new biotechnology that is part of growing interest for extensive research in tissue engineering and regenerative medicine nowadays **(Niemeyer et al., 2010)**.

Platelet-rich plasma is autologous plasma with high platelet concentration used to accelerate healing, due to several different growth factors and cytokines. PRP is easy, cheap, safe, and minimally invasive source for autologous growth factors **(Cavallo et al., 2016)**. Platelet rich plasma is prepared by centrifugation with different centrifugal forces, temperature, and time. Accordingly the two-step procedure is most widely used, in which the first step (soft spin) separates plasma buffy coat and cells and a second spin (hard spin) further concentrates platelets with an outcome of platelets exceeding  $400 \times 10^9/\text{L}$ ; unlike platelet poor plasma which gives platelets less than  $10 \times 10^9/\text{L}$  following vigorous centrifugation **(Alves & Grimalt, 2018)**.

Platelets are disk shaped fragments of megakaryocytes originating from stem cells in the bone marrow. Platelets are approximately  $2 \mu\text{m}$  in diameter **(Machlus et al., 2014)** **(Michelson & Alan, 2013)**. The platelets count ranges from

150-400 x 10<sup>9</sup> per litre with a life span of 7-10 days (**Thon et al., 2010**). Although platelets most known function is hemostatic, a process which controls bleeding by forming a clot at the site of a cut or wound by sticking platelets together; other beneficial effects of PRP have been revealed such as cell proliferation, differentiation, improved synthesis of extracellular matrix (ECM), and angiogenesis which results in neovascularization and formation of a new conjunctive tissue necessary for healing (**Marwah et al., 2014**) (**Cervantes et al., 2018**).

Platelets contain alpha ( $\alpha$ ) granules, dense granules, and lysosomes; with  $\alpha$  granules being most abundant. Each type has specific properties concerning both the structure and function. Alpha granules -80 granules per platelet- contain growth factors (such as PDGF, bFGF, SDF 1 $\alpha$ ), angiogenic factors (such as VEGF, angiogenin), necrotic factors (such as TNF  $\alpha$ , TNF  $\beta$ ), hemostatic (such as factor V, VWF, Fibrinogen), and other cytokines (**Blair & Flaumenhaft, 2009**) (**Whiteheart, 2011**). Alpha-Granules function in adhesion of platelets and healing (**Rendu et al., 2001**). The dense granules contain adenosine diphosphate (ADP), adenosine triphosphate (ATP), ionized calcium, and serotonin (**Orkin et al., 2001**). Dense granules function in recruiting other platelets. Lysosomes contain hydrolases that eliminate platelet aggregates (**Rendu et al., 2001**).

The platelets can be activated *in vivo* or *in vitro*. Once activated, platelets show morphological changes from the discoid to round shape, develop pseudopods that spread through the injured tissue, and platelets adhere to one another and to collagen in endothelium forming platelet aggregates. Degranulation of their granules also occur releasing GFs from  $\alpha$ -granules by either fusion with the open canalicular system and extrusion of its contents through small channels in the cell membrane, or by exocytosis, that is releasing  $\alpha$  granules content outside by fusing the granules with the cell membrane (**Zandim et al., 2012**). Additionally, a platelet gel forms through a clotting process by cleavage of fibrinogen. The factors activate an intracellular signal protein that causes the expression of a gene sequence that directs cell migration, proliferation and differentiation, and promote extracellular matrix accumulation by binding to specific cell surface receptors on target cells (eg. Mesenchymal stem cells, osteoblasts, fibroblast, endothelial cells, and epidermal cells) thus provoking tissue repair and regeneration (**Dhurat and Sukesh, 2014**).

Activation may be initiated *in vivo* by thrombin, calcium, collagen, and shear stress. In clinical practice, PRP may be pre-activated using *in vitro* calcium or/and thrombin (bovine or autologous) to induce release of GF from PRP.

However, evidence is lacking for pre-injection activation requirement for therapeutic uses (**Cavallo et al., 2016**).

PRP is prepared for either harvesting platelets for therapeutic purposes or testing platelet function using aggregometer (**Gentile et al., 2010**). Therapeutic clinical applications for PRP have revealed remarkable results in dentistry (**Anitua, 1999**), surgeries (**Patel et al., 2016**) (**Arnalich et al., 2016**), orthopaedics (**Filardo et al., 2012**), rheumatology (**Cieslik-Bielecka et al., 2009**), chronic wounds (**Rozmam et al., 2007**), and muscle injuries (**Hammond et al., 2009**).

In recent years, platelets were discovered to have a positive influence on the liver thus promoting its regeneration (**Murata et al., 2014**) (**Takahashi et al., 2013**), improving liver fibrosis (**Takahashi et al., 2013**) (**Nowatari et al., 2014**) (**Hesami et al., 2014**), and attenuating liver damage (**Hisakura et al., 2011**); as it accelerates liver regeneration and have anti-fibrosis and anti-apoptotic activity. Platelets have a direct impact on hepatocytes, a supportive effect with liver sinusoidal endothelial cells, and a cooperative effect with Kupffer cells which accelerates liver regeneration. Platelets undergo adenosine-cyclic adenosine 5'-monophosphate signaling pathway which deactivates hepatic stellate cells thus applying anti-fibrotic activity. Platelets activate the Akt pathway and up-regulates Bcl-xL, which in turn suppresses caspase-3 activation thus preventing hepatocyte apoptosis (**Takahashi et al., 2013**).

## **Aim of the Work**

**T**o compare different strategies of PRP activation by evaluating content and release of growth factors for further clinical trials to validate the clinical relevance of best method to help regeneration of liver in hepatic patients.

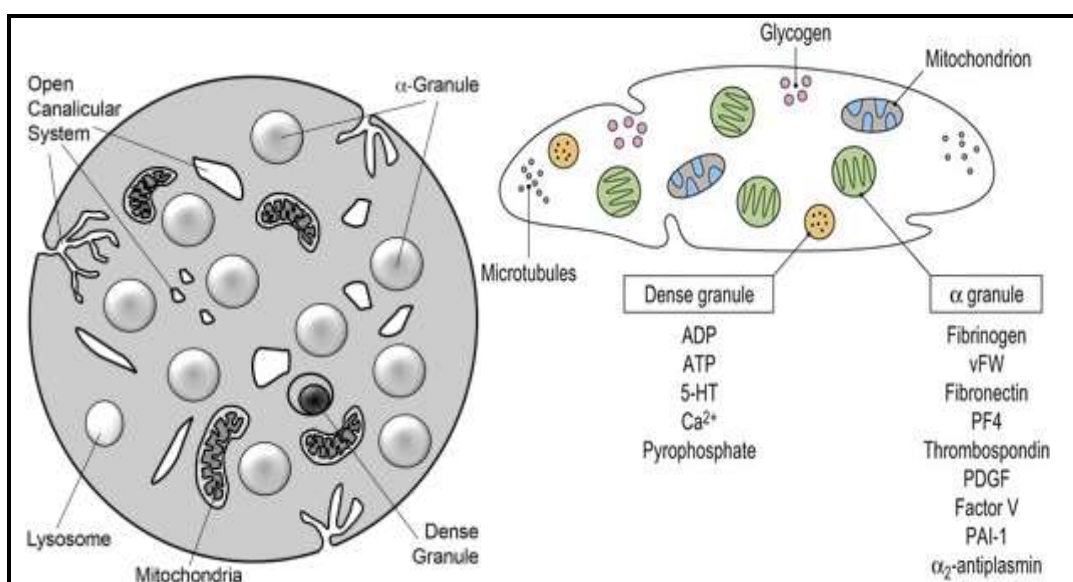
## Chapter 1: Platelets

Platelets, or thrombocytes, are small (approximately 2-3  $\mu\text{m}$  in diameter), rounded or oval disc-shaped, anucleate cells formed from the fragmentation of long proplatelet extensions, that become interwoven through endothelial pores of the bone marrow sinusoids and are fragmented by shear forces releasing platelets from the megakaryocyte. Megakaryocyte is large multinucleated cell from myeloid lineage of hematopoietic stem cells constituting 0.1% of cellular population of the bone marrow (**Junt et al., 2007, Francone et al., 1990**). A single megakaryocyte can give rise to 1,000–3,000 platelets (**Stenberg and Levin; 1989**) before the residual nuclear material is eliminated by macrophage-mediated phagocytosis (**Radley and Haller; 1983**). Differentiation of stem cells to platelets is predominantly driven by thrombopoietin (TPO) signaling through the cMpl receptor, and is supported by additional growth factors such as interleukin-3 (IL-3), stem cell factor (SCF), IL-6, and IL-11 (**Kaushansky, 2006**).

Thrombopoietin is the principal regulator of thrombopoiesis and primate platelet counts can be reduced with an inhibitor against TPO without impairment of primary hemostasis (**Kaushansky; 2006, Tucker et al., 2010**). Platelets count ranges from 150,000 to 350.000/l (**Weibrich**

**et al., 2002).** The efficiency of platelets is variable, affected by size and age of the platelet, with younger and larger platelets exhibiting better hemostatic function than smaller and older ones (**Karpatkin,; 1978, Hartley; 2007**). Platelets contribute to many aspects of host defense including hemostasis and thrombosis, inflammation, angiogenesis, and wound healing (**Franco et al., 2015**).

Although they lack a nucleus, platelets have an extensive cytoskeleton, organelles, specific granules and membrane features (**Weyrich et al., 2009**).



**Figure (1):** Diagrammatic representation of coronal (left) and longitudinal (right) section of the platelet.