



The effect of *Lactobacillus acidophilus* as a probiotic against *Pseudomonas aeruginosa* growth and biofilm formation

Thesis

Submitted for Partial Fulfillment of Master *Degree in Basic Medical Sciences*
(*Medical Microbiology & Immunology*)

Presented by

Shaimaa Adel Mohammed El-badri Soliman

M.B.B.Ch.

Faculty of medicine-Ain Shams University

Under supervision of
Prof. Dr.Marwa Saad Fathi

Professor of Medical Microbiology and Immunology
Faculty of Medicine-Ain Shams University

Dr. Amira Esmail Abdel Hamid

Lecturer of Medical Microbiology and Immunology
Faculty of Medicine-Ain Shams University

Assist.Prof. Dr. Hanaa Moohamed Abd Allah El Gendy

Assistant Professor of Anesthesia, Intensive Care and Pain Management
Faculty of Medicine-Ain Shams University

Faculty of Medicine
Ain Shams University

2019

Acknowledgment

*Thanks first and last to **ALLAH** for his guidance, support and care in every step throughout my life.*

*I have the greatest pleasure to express my deepest appreciation to **Assist Prof. Dr. Marwa Saad Fathi**, Assistant Professor of Medical Microbiology and Immunology, Faculty of Medicine, Ain Shams University for her unlimited help, guidance, suggestions and supervision, as well as her kindness and continuous advice to ensure that this work would reach an efficient level.*

*I wish to express my profound gratitude to **Dr. Amira Esmail Abdel Hamid**, Lecturer of Medical Microbiology and Immunology, Faculty of Medicine, Ain Shams University for her kind help and assistance, valuable supervision, support, precious opinions and contributive comments that served much in the construction of this work.*

*Also, I want to express my great thanks to **Assist Prof. Dr. Hanaa Mohamed Abd Allah El Gendy**, Assistant Professor of Anesthesia, Intensive Care and Pain management, Faculty of Medicine Ain Shams University, for her guidance and help.*

My greatest thanks to all my colleagues in the department of Medical Microbiology and Immunology for their cooperation and advice.

*Finally I would like to acknowledge with gratitude the support, continuous encouragement and love of **my family: the soul of my father, my gorgeous mother** the origin of my success and **my great husband and son Adam**.*

List of Contents

	<i>Page No.</i>
• List of Abbreviations	I
• List of Figures	IV
• List of Tables	VIII
• Abstract & key words	IX
• Introduction	i
• Aim of the Work	iii
• Review of Literature:	
○ Chapter (I): An overview on bacterial biofilm	1
○ Chapter (II): <i>Pseudomonas aeruginosa</i> as strong biofilm producer	26
○ Probiotics & <i>Lactobacillus acidophilus</i>	38
• Patients and Methods	53
• Results	69
• Discussion	88
• Summary	95
• Conclusion	96
• Recommendations	97
• References	98
• Arabic Summary	1

☞ List of Abbreviations ☞

A	AHLs	Acylated homoserine lactones
	AIDs	Acquired immunodeficiency syndrome
	AME	Aminoglycoside modifying enzymes
	AMPs	Antimicrobial Peptides
	ATPase	Adenosine triphosphatase
C	°C	Degrees Celsius
	<i>C.albicans</i>	<i>Candida albicans</i>
	CF	cystic fibrosis
	CFU	Colony forming unit
	CTAs	Cancer-testis antigens
D	Dept	Department
	DNA	Deoxyribonucleic acid
	DNAses	Deoxyribonucleases
E	eDNA	Extracellular DNA
	EDTA	Ethylenediaminetetraacetic acid
	EPS	Extracellular polymeric substances
	<i>E-coli</i>	<i>Escherichia coli</i>
	ESBLs	extended-spectrum β -lactamases
F	FAO	Food and Agriculture Organization
G	GALT	Gut-associated immune cells
	<i>G. vaginalis</i>	<i>Gardenella vaginalis</i>
	Gm	Gram
H	HAI	Health care associated infections
	H₂O₂	Hydrogen peroxide
	HDL	High density lipoprotein
	HGT	Horizontal gene transfer

	Hrs	Hours
	HSL	Homoserine lactone
I	ICUs	Intensive care units
	IgA	Immunoglobulin A
	IL	Interleukin
	IFN-γ	Interferon gamma
k	KD	Killo Dalton
L	LAB	Lactic acid bacteria
	<i>L.acidophilus</i>	<i>Lactobacillus acidophilus</i>
	LDL	low density lipoprotein
M	MBC	minimum bactericidal concentration
	MDEPs	multidrug efflux pumps
	MDR	Multi drug resistance
	MMRS	methyl- directed mismatch repair system
	Mg	Milligram
	μg	Microgram
	MIC	Minimum inhibitory concentration
	Min	Minute
	ml	Milliliter
	μL	Microliter
	Mub	Mucus-binding protein
	MTP	Microtitre plate
N	<i>N. gonorrhoeae</i>	<i>Nisseria gonorrhoeae</i>
O	Opr	Outer membrane protein
P	<i>P.aeruginosa</i>	<i>Pseudomonas aeruginosa</i>
	PVC	polyvinylchloride
Q	QS	Quorum sensing
	QSI	Quorum sensing inhibitors

R	ROS	radioactive oxygen species
S	<i>S. agalactiae</i>	<i>Streptococcus agalactiae</i>
	<i>S.aureus</i>	<i>Staphylococcus aureus</i>
T	T_{reg}	Regulatory T cells
	Th2	T helper 2
u	UC	Urinary catheter
W	WHO	World Health Organization

❧ List of Figures ❧

<i>Figure No.</i>	<i>Title</i>	<i>Page No.</i>
Figure (1):	a A model of a bacterial biofilm attached to a solid – surface.b The major matrix components.....	5
Figure (2):	Different phases of biofilm formation	6
Figure (3):	Quorum sensing	11
Figure (4):	Biofilm and antibiotic resistance	15
Figure (5):	Antibiotics and persister cells in biofilm	16
Figure (6):	Strategies aimed at disrupting biofilm formation	25
Figure (8):	Gram negative bacterial envelop	32
Figure (9):	The 3 proteins components of MDEPs.....	33
Figure (10):	MexAB-oprM in <i>P. aeruginosa</i>.....	33
Figure (11):	Mechanisms of HGT.....	37
Figure (12):	Structure of cell envelope of <i>L.acidophilus</i>	42
Figure (13):	<i>Show</i> different mechanism of actions of probiotics Lactobacilli.....	48
Figure (14):	A plate of CLED agar medium cultured with urine sample	54
Figure (15):	Inoculated plate of MacConkey's agar showing non – lactose fermenting colonies.....	55
Figure (16):	Exopigment production on Nutrient agar slope, plate and Non-lactose fermenting colonies onMacConkey's agar	55
Figure (17):	Biochemical reactions of <i>P.aeruginosa</i>	56
Figure (18):	Rose pink colonies on macConky.....	57

Figure (19): Mucoid pink colonies on macConky.....	57
Figure (20): Yeast cells of <i>C. albicans</i>	57
Figure (21): <i>C. albicans</i> showing positive germ tube test.....	57
Figure (22): Template for applying antimicrobial discs.....	58
Figure (23): A plate of MHA streaked for antibiotic susceptibility testing of <i>P. aeruginosa</i> isolate.....	59
Figure (24): A plate of MHA streaked for antibiotic susceptibility testing of <i>P. aeruginosa</i> isolate which was multi drug resistant, but sensitive to Aztreonam	60
Figure (25): Lyophilized <i>L.acidophilus</i> ATCC 4356 & MRS agar plat.....	60
Figure (26): Illustrated instruction for culture methods of the lyophilized <i>L.acidophilus</i> ATCC 4356.....	61
Figure (27): Microscopic examination of a Gram stained film of <i>L.acidophilus</i>	61
Figure (28): Cell-free supernatant filtered with 0.20 μ Porous membranes	62
Figure (29): different diameters of the growth inhibitory zone of <i>L. acidophilus</i> on <i>P. aeruginosa</i>	63
Figure (30): MTP after washing with distilled water	64
Figure (31): MTP showing the effect of <i>L.acidophilus</i> against biofilm formations of different <i>P.aeruginosa</i> strains	65
Figure (32): Strains of <i>P.aeruginosa</i> were 1st allowed to grow in wells for 24 h and form biofilms	67.
Figure (33): After adding cell-free supernatant of <i>L.acidophilus</i> to the 2 nd 3 wells in all rows and incubation for another 24 hours.....	67
Figure (34): Shows the effect of <i>L.acidophilus</i> against preformed biofilm of different <i>P.aeruginosa</i> strains after staining	68

Figure (35): Gender distribution among the patients having UTI caused by <i>P. aeruginosa</i>	70
Figure (36): Antimicrobial resistance pattern of the isolated <i>P. aeruginosa</i> strains	76
Figure (37): High statistical significance in resistance to Gentamycin in patient taking immunosuppressive drugs.....	79
Figure (38): The percentage of <i>P.aeruginosa</i> isolated strains inhibited by <i>L.acidophilus</i>	81
Figure (39): Degree of biofilm formation by the isolated <i>P.aeruginosa</i> strains.....	82
Figure (40): The effect of <i>L.acidophilus</i> cell- free supernatant on biofilm forming <i>P.aeruginosa</i> isolated strains as illustrated by decrease in OD	83
Figure (41): The effect of <i>L.acidophilus</i> cell- free supernatant on biofilm forming <i>P.aeruginosa</i> isolated strains as illustrated by decrease in biofilm forming ability.....	84
Figure (42): The effect of <i>L.acidophilus</i> cell free supernatant on the preformed biofilms of <i>P.aeruginosa</i> strains as illustrated by the decrease in OD	85
Figure (43): The effect of <i>L.acidophilus</i> cell free supernatant on the preformed biofilms of <i>P.aeruginosa</i> strains as illustrated by decreasing the biofilm forming ability	85
Figure (44): Difference in OD after mixing with <i>L.acidophilus</i> with <i>P. aeruginosa</i> isolates before biofilm growth and after adding on preformed <i>P.aeruginosa</i> biofilm	86
Figure (45): Difference in biofilm forming ability of <i>P. aeruginosa</i> isolates after adding <i>L.acidophilus</i> cell free supernatant before and after biofilm formation by <i>P. aeruginosa</i> isolates	86
Figure (46): Relation between biofilm forming ability of <i>P. aeruginosa</i> isolates and age of the patients	87

Figure (47): Relation between biofilm forming ability of *P. aeruginosa* isolates and the duration of stay in ICU..... 88

Figure (48): Relation between biofilm forming ability of *P. aeruginosa* isolates and the application of urinary catheter..... 88

❧ List of Tables ❧

<i>Table No.</i>	<i>Title</i>	<i>Page No.</i>
Table (1):	Examples of drug affected by up regulation of genes of efflux pump as described in	36
Table (2):	Criteria for selection of probiotic strains.....	40
Table (3):	Effect of different Lactobacilli strains on Inflammatory cytokines.....	44
Table (4):	Nutritional benefits & potential health of foods prepared with probiotic bacteria.....	49
Table (5):	Health benefits by different probiotic strains.....	52
Table (6):	Identification of isolated organisms	56
Table (7):	Antibiotic discs used for antibiotic susceptibility testing for isolated <i>P. aeruginosa</i> strains and their zones diameter breakpoints	59
Table (8):	Demographic data of the patients	70
Table (9):	Clinical data of the patients included in this study	71
Table (10):	Frequency of <i>P.aeruginosa</i> and other organisms isolated from the 135 urine samples.....	72
Table (11):	Organisms identified in samples showing mixed infections together with <i>P. aeruginosa</i>	73
Table (12):	The relation between the application of urinary catheter and occurrence of UTI by <i>P.aeruginosa</i>	73
Table (13):	The relation between prior antibiotic intake and occurrence of UTI by <i>P.aeruginosa</i>	74
Table (14):	The relation between immunosuppressive drugs intake and occurrence of UTI by <i>P.aeruginosa</i>	74

Table (15):	Antibiotic susceptibility testing results of isolated <i>P.aeruginosa</i> strains	75
Table (16):	Statistical analysis between antibiotic susceptibility and prior antibiotics intake.....	77
Table (17):	Statistical analysis between antibiotic susceptibility and immunosuppressive drugs intake	78
Table (18):	Statistical analysis between antibiotic susceptibility and the application of urinary catheter.....	80
Table (19):	Effect of <i>L. acidophilus</i> on the growth of <i>P. aeruginosa</i> by agar well diffusion	81
Table (20):	Degree of biofilm formations among different isolated <i>P.aeruginosa</i> strains	82
Table (21):	The effect of <i>L.acidophilus</i> cell free supernatant on biofilm formation by <i>P.aeruginosa</i> strains	83
Table (22):	The effect of <i>L.acidophilus</i> on the preformed biofilm of <i>P.aeruginosa</i> strains.....	84
Table (23):	Relation between the biofilm forming ability of <i>P.aeruginosa</i> isolated strains and some demographic and clinical data.....	87

Abstract

The emergence of antibiotic-resistant microorganisms as *Pseudomonas aeruginosa* has pushed efforts to find safe alternatives of antibiotics as probiotics which include Lactobacilli strains with a large antibacterial and anti-biofilm effects against different pathogenic strains. Antibiotic susceptibility test and antibiogram were done for 35 isolated *P. aeruginosa* strains from ICU patients followed by assessment of *Lactobacillus acidophilus* as antibacterial and antibiofilm formation/removal against them by using different methods. The results showed that among the 35 *P. aeruginosa* strains, 1 (2.8%), 2 (5.7%), and 32 (91.4%) were PDR, XDR, and MDR, respectively. The effective antibiotics against them was Aztreonam. *L.acidophilus* with ATCC 4356, showed powerful inhibition and anti-biofilm effects against all isolated *P. aeruginosa* strains. It significantly inhibits biofilms by 68.52% with mean optical density reading 0.72 ± 0.44 and removes already formed biofilms by 43.8 % with mean optical density reading 1.28 ± 0.65 . It was concluded that *L.acidophilus* can be used in bio-control of different antibiotic resistant biofilm producing *P.aeruginosa* strains.

Key words:

Antibiotic resistance. *Lactobacillus-acidophilus*. Probiotic. *Pseudomonas-aeruginosa*

Introduction

Bacterial biofilms are complex structured communities of bacterial cells enclosed in a self-produced polymer matrix that is attached to a surface. Biofilms protect and allow bacteria to survive and thrive in hostile environments. Bacteria within biofilms can withstand host immune responses and are much less susceptible to antibiotics and disinfectants when compared to their planktonic counterparts. Diseases associated with biofilms require great methods for their prevention, diagnosis and treatment (*Tremblay et al., 2014*).

P. aeruginosa causes a threatening diseases world wide especially in compromised immunity . it's one of the most common causes of recurrent UTI. The emergence of multidrug-resistant (MDR), extensively drug-resistant (XDR), and pan-drug-resistant (PDR) strains of *P. aeruginosa* that are treated difficulty with current antibiotics causes wide efforts for finding an alternative approach for treatment (*Gomila ,et al., 2013 ; Ha D, O'Toole ,2015*)

Probiotics are defined as live nonpathogenic microorganisms that when administered in adequate amounts confer a health benefit on the host, The use of probiotics especially *L.acidophilus* as safe and natural live microorganisms against other microorganisms are considered as an alternative to antibiotics (*Hill, et al.,2014; Chatterjee , et al., 2015*)

As a probiotic, *L.acidophilus* can be active as microbial barriers against pathogens by competition with them for binding sites, enhancement of the host's immune responses and production of antimicrobial substances as hydrogen peroxide , organic acids like lactic, formic, acetic, propionic and butyric acids together with proteinaceous

compounds as bacteriocins, bacteriocin-like components (*Marianelli et al, 2010*).

L.acidophilus has potent activity of biofilm inhibition/removing, in addition to its antibacterial effect. This proposes it as potential probiotics for bio-control of antibiotic-resistant *P. aeruginosa* strains as alternative to antibiotics (*Shokri et al ., 2018*).