



Pathological Studies on The Protective Effect of Ginger Extract and Ginger Nanoparticle on Acetaminophen Toxicity in Rats

Submitted by

Alaa Fouad Ali

B.v.sc (2012), M.Sc. (2016), Faculty of Veterinary Medicine

Cairo University

For Ph.D. Degree of Veterinary Medical Science

Pathology (General, Special and Postmortem)

Under Supervision of

Prof. Dr. Adel Mohamed Bakeer

Department of Pathology, Faculty of Veterinary Medicine Cairo University

Prof. Dr. Sherein Saied Abdelgayed

Department of Pathology
Faculty of Veterinary Medicine
Cairo University

Prof. Dr. Osama Samir Zaky El-Tawil

Department of Toxicology and Forensic Medicine, Faculty of Veterinary Medicine Cairo University بسم الله الرحمن الرحيم اقالُواْ سُبْحَانَكَ لاَ عِلْمَ لَنَا إِلاَّ مَا عَلَّمْ تَنَا إِلاَّ مَا عَلَّمْ تَنَا إِنَّكَ أَنتَ الْعَلِيمُ الْحَكِيمُ" عَلَّمْ تَنَا إِنَّكَ أَنتَ الْعَلِيمُ الْحَكِيمُ" البقرة - (32)



Supervision sheet

Prof. Dr. Adel Mohamed Baker

Professor of Pathology
Department of Pathology
Faculty of Veterinary Medicine
Cairo University

Prof. Dr. Sherein Saied Abdelgayed

Professor of Pathology
Department of Pathology
Faculty of Veterinary Medicine
Cairo University

Prof. Dr. Osama Samir Zaky El-Tawil

Professor of Toxicology and Forensic Medicine Department of Toxicology and Forensic Medicine Faculty of Veterinary Medicine Cairo University Cairo University
Faculty of Veterinary Medicine
Department of Pathology

Name: Alaa Fouad Ali Bakr Date of Birth: 30/3/1991

Nationality: Egyptian

Specialization: Pathology (General, Special, Postmortem).

Title: Pathological Studies on The Protective Effect of Ginger Extract and Ginger

Nanoparticles on Acetaminophen Toxicity in Rats

Degree: Ph.D. of Veterinary Medical Sciences, Pathology (General, Special and Postmortem)

Supervisors:

Prof. Dr. Adel Mohamed Bakeer, Professor of Pathology, Department of Pathology, Faculty of Veterinary Medicine, Cairo University.

Prof. Dr. Sherein Saied Abdelgayed, Professor of Pathology, Department of Pathology, Faculty of Veterinary Medicine, Cairo University.

Prof. Dr. Osama Samir Zaky El-Tawil, Professor of Toxicology and Forensic Medicine, Department of Toxicology and Forensic Medicine, Faculty of Veterinary Medicine, Cairo University.

Abstract

Acetaminophen (APAP) is widely used analgesics all over the world. However, hepatotoxicity and nephrotoxicity are the most remarkable features of acetaminophen overdose. Ginger is a medicinal plant that has immuno-modulatory, anti-tumorigenic, anti-inflammatory, anti-apoptotic, antihyperglycemic, anti-lipidemic and anti-oxidant activities. Recently, ginger extract in form of nanoparticles (NPs) have been investigated in liver protection against alcohol-induced liver damage. We conducted *in vivo* and *in vitro* studies to compare between the protective efficacy of ginger extract and ginger nanoparticles (GNPs) against acetaminophen-induced toxicity from histological and biochemical aspects. In vivo study, adult male Sprague Dawley rats were received ginger extract and GNPs orally at dose of 120 mg/kg (3times/week) and acetaminophen at dose of 375mg/kg (1/10LD₅₀) daily for three months. Serum Alanine aminotransferase (ALT) and aspartate aminotransferase (AST) as liver functionality indicator were determined. In addition, kidneys functions were assessed by evaluating urea and uric acid levels in serum. Moreover, oxidative stress at the cell level was evaluated by determining malondialdehyde content (MDA) and catalase enzyme (CAT) activity. Meanwhile, histopathological changes in liver and kidney tissues were observed. The present study indicates that liver and kidney biochemical markers are improved in rat pretreated with ginger extract and ginger nanoparticles as well as the activities of cell oxidative stress are statistically significant diminished. In

addition, their histological structure of liver and kidney tissues show very little changes. While rats treated with GNPs demonstrate normal biochemical and oxidative stress marker levels and histological structure relative to ginger extract treated rat.

While *in vitro* study, we use ginger extract and ginger nanoparticles at concentration of 60 µg/ml against hepatotoxicity caused by acetaminophen (APAP) at concentration of 0.1 mg/ml using primary isolated rat hepatocytes. Cytotoxicity was determined by assessing cell viability and leakage of cytosolic enzymes, such as (ALT& AST). Oxidative stress was investigated by measuring levels of CAT and MDA. The cytopathological alterations were studied by light microscope. Exposure of isolated rat hepatocytes to APAP caused cytotoxicity and oxidative injury, manifested by loss of cell viability and significant increase in ALT and AST leakages. As well as, APAP caused progressive depletion of CAT content and increase intracellular MDA accumulation, in addition to alteration in cellular morphology. Pretreatment of hepatocytes with either GE or GNPs ameliorated the hepatotoxicity, oxidative stress and enzymatic leakage induced by APAP. However, GNPs were more effective relative to ginger extract pretreated hepatocytes. From both studies we concluded that, ginger nanoparticles have more protective activities relative to ginger extract.

Key Words

Acetaminophen - ginger extract - ginger nanoparticles - oxidative stress- histopathology- isolated hepatocytes.

Dedication

To my dear parents,

To my sisters and brothers,

To my daughter Malika and my son Ali,

To my husband Mohamed and his family

Thanks a lot, I appreciate your great efforts

Acknowledgment

I would like to give special thanks to **Prof. Dr. Adel Mohamed Bakeer**, Professor of Pathology, Pathology Department, Faculty of Veterinary Medicine, Cairo University for his kind supervision, guidance, continuous support, and encouragement. I am honored, thankful and fortunate to be a student of Prof. Dr. Adel Mohamed Bakeer.

I would like to express my appreciation to **Prof. Dr. Sherein Saied**Abdelgayed Professor of Pathology, Pathology Department, Faculty of

Veterinary Medicine, Cairo University for her kind supervision, guidance,

continuous support, encouragement and for suggesting the point of this thesis.

It is pleasuring to express my great thanks to **Prof. Dr. Osama Samir**Zaky El-Tawil, Professor of Toxicology, Forensic Medicine and Veterinary

Regulations Department of Toxicology and Forensic Medicine, Faculty of

Veterinary Medicine, Cairo University, for nominating the idea behind this

study, drawing the plan of work, and kind advice.

Also, I would like to express my gratitude to every member in the Pathology Department, Faculty of Veterinary Medicine, Cairo University for their help, cooperation and support.

CONTENTS

Chapter	Item	Page
number		
1	Introduction	1
2	Review of Literature	3
3	Published research	36
	Assessment of Ginger Extract and Ginger Nanoparticles Protection Activity against Acetaminophen-induced Hepatotoxicity and Nephrotoxicity in Rats.	36
	In Vitro Hepatoprotective Effect of Ginger (Extract and Nanoparticles) Against Acetaminophen Induced Hepatotoxicity in Isolated Rat Hepatocytes.	56
4	Discussion	77
5	Conclusion	83
6	Summary	84
7	References	88
8	Appendix	103
Arabic summary		

LIST OF ABBREVIATIONS

ALT	Alanine aminotransferase
APAP	Acetaminophen (Paracetamol)
AST	Aspartate aminotransferase
ATN	Acute tubular necrosis
BUN	Blood urea nitrogen
CAT	Catalase
Ccl4	Carbon tetrachloride
Cd	Cadmium
Cox-2	Cyclo-oxygenase
COX-3	cyclo-oxygenase
CYP 2E1	Cytochrome 2E1
DNA	Deoxyribonucleic acid
DPPH	2,2-diphenlyl-1-picrylhydrazyl
GE	Ginger extract
GNPs	Ginger nanoparticles
GSH-Px	Glutathione peroxidase
Gst pi	Glutathione S-transferase Pi
H&E	Hematoxylin and eosin
HRS	Hepatorenal syndrome
IHC	Immunohistochemistry
IL-1β	Interleukin-1β
IL-33	Interleukin-33
iNOS	inducible nitric oxide synthase
MDA	Malondialdehyde
NAC	N-Acetylcysteine
NAPQI	N-acetyl-para-benzoquinone imine
NO	Nitric oxide
Pb	Lead
RNA	Ribonucleic acid
ROS	Reactive oxygen species
RSTI	Repeated supratherapeutic ingestion
SGOT	Serum glutamate oxaloacetate transaminase
SGPT	Serum glutamate pyruvate transaminase
SOD	Superoxide dismutase

SPSS	Statistical package for social science
TBARS	Thiobarbituric acid reactive species
TNF-α	Tumor necrosis factor α
TPA	12-O- tetradecanoylphorbol-13-acetate
Vd	volume of distribution

LIST OF FIGURES

First Published Paper Ginger Extract and Ginger Nanoparticles; Characterization and Applications		
Fig (1)	Showing the structure of 6-gingerol, 8-gingerol and 10-gingerol	22
Fig (2)	The structure of 6-gingerol and 6- shogaol	22
	Review	"
Fig (1)	Rumack-Matthew Nomogram	27
	Second Published Paper	,,
Assessn	nent of Ginger Extract and Ginger Nanoparticles Protection Activity aga Acetaminophen-induced Hepatotoxicity and Nephrotoxicity in Rats	inst
Fig (1)	Showing the mean values of serum AST (a&b) and ALT (c&d) in all groups.	47
Fig (2)	Showing the mean values of liver CAT (a&b) and MDA (c&d) in all groups	48
Fig (3)	Showing the mean values of urea (a&b) and uric acid (c&d) in all groups	49
Fig (4)	(a, b and c) Liver of G4 rat (after 6 weeks) showing (a) vacuolar degeneration of hepatocytes with dilatation of sinusoid (X400). (b) centrilobular necrosis with infiltration of inflammatory cells (X 100). (c) periportal edema with infiltration of mononuclear cells and formation of fibrous connective tissue (X 200). d, e and f (after 12 weeks) showing (d) fatty change (X 200). (e) sever necrosis with macrophages infiltration associated with sinusoidal dilatation (X 100). (f) congestion in portal area with edema and infiltration of inflammatory cells and appearance of newly formed bile ducts (X 200). All pictures were stained with H & E.	
Fig (5)	(a and b) Liver of G5 rat (after 6 weeks) showing (a) showing focal area of necrosis associated with infiltration of inflammatory cells and kupffer cells activation, hepatocyte suffering from mild vacuolation and apoptosis (X 200). (b) mild congestion in portal area with infiltration of inflammatory cells (X 200). c and d liver of G5 (after 12 weeks) showing (c) necrosis of hepatocyte with macrophages infiltration associated with sinusoidal dilatation (X 100). (d) congestion of portal blood vessels with edema and formation of fine fibrous connective tissue (X 100). (e) Liver of G6 rat (after 6 weeks) showing infiltration of inflammatory cell in portal area and between hepatic cords (X 400). (f) Liver of G6 rat (after 12 weeks) showing	50

	vacuolar degeneration and apoptosis of sporadic hepatocytes (X 400). All	
	pictures were stained with H & E.	
Fig (6)	(a and b) Kidney of G4 rat (after 6 weeks) showing (a) vacuolar degeneration of renal tubules with infiltration of inflammatory cells (X 400). (b) focal interstitial nephritis and degeneration of renal tubules (X 200). c, d, e and f (after 12 weeks) showing, (c) degeneration and necrosis of renal tubules. Notice the eosinophilic substance inside the lumen of tubules (X 400). (d) interstitial nephritis and degeneration in renal tubules (X 200). (e) perivascular edema and thickening in basement membrane of renal glomeruli with dilatation of bowman's space (X 200). (f) atrophy of renal glomeruli (X 200). (g) focal area of fibrous connective tissue proliferation with vacuolar degeneration (X 400). All pictures were stained with H & E.	51
Fig (7)	(a and b) Kidney of G5 rat (after 6 weeks) showing (a) congested blood vessel with perivascular edema associated with inflammatory cells (X 100). (b) vacuolar degeneration. c and d (after 12 weeks) showing (X 200). (c) necrobiotic changes in renal tubules. Note the renal cast in the lumen of tubules (X 200). (d) interstitial nephritis and degeneration of renal tubules (X 100). (e) Kidney of G6 rat (after 6 weeks) showing congestion of renal blood vessels with infiltration of inflammatory cells (X 100). (f) Kidney of G6 rat (after 12 weeks) showing perivascular edema associated with infiltration of inflammatory cells, degeneration and necrobiotic changes in renal tubules and dilated Bowman's spaces (X 200). All pictures were stained with H & E.	52
Fig (8)	Immunostaining for iNOS. (a, b and c): Liver of rats G1, G2 and G3 showing negative expression of iNOS (X 400). (d): Liver of rat from G4 showing marked strong positive expression of iNOS (brown color) in hepatocytes (X 400). (e) Liver of rat from G5 showing moderate expression of iNOS (X 400). (f) Liver of rat from G6 showing weak expression of iNOS (X 400).	54
	Third Published Paper	<u> </u>
In Vit	ro Hepatoprotective Effect of Ginger (Extract and Nanoparticles) Agains	st
	Acetaminophen Induced Hepatotoxicity in Isolated Rat Hepatocytes	
Fig 1.	The graphical abstract of this study.	56

Fig 2.	(A&B) SEM images of GNPs. Images are representative of a minimum of	70
	3 independent samples, with $n > 500$ NPs assessed in total	
Fig 3.	Showing the mean values of viability in all groups at different point times	70
Fig 4.	Showing the mean values of ALT in all groups at different point times	71
Fig 5.	Showing the mean values of AST in all groups at different point times	71
Fig 6.	Showing the mean values of MDA in all groups at different point times	72
Fig 7.	Showing the mean values of CAT in all groups at different point times	72
Fig 8.	(A) Small group of hepatocytes from control group (G1) showing normal round cells with centric round nucleus and intact well-defined cell membrane (x1000). (B) Binucleated single hepatocyte from G1 showing eosinophilic granular cytoplasm (x1000). (C) Hepatocytes from ginger extract treated group (G2) showing normal structure (x1000). (D) Single hepatocyte from ginger nanoparticles treated group (G3) showing normal cytological structure (x1000). All slides stained with H&E.	73
Fig 9.	Hepatocytes of APAP treated group (G4) showing, (A) Hepatocellular cytoplasm appears foamy due to presence of multiple round clear vacuoles, nucleus appear pyknotic and move to peripheral side (x1000). (B) Two hepatocytes, one of them showing pyknotic nucleus (arrow) and the other one showing necrosis with complete lysis of nucleus (x1000). (C) Three hepatocytes (arrows) showing necrosis and complete lysis of nucleus (x1000). (D) nucleus undergo lysis associated with complete ballooning of hepatocytes (x1000). All slides stained with H&E.	74
Fig 10.	(A) Hepatocytes of GE+APAP treated group (G5) showing pyknotic nucleus with deep eosinophilic cytoplasm (x1000). (B) Single hepatocyte from G5 showing single vacuole with pyknotic nucleus and irregular cell membrane (x1000). (C) Hepatocytes from G5 showing vacuolar degeneration of cytoplasm (arrow), moreover the nucleus undergoes lysis (x400). (D) Two hepatocytes from G5, one of them showing necrosis (arrow), and the other one showing single vacuole with pyknotic nucleus (x1000). (E) Single hepatocyte from GNPs+APAP treated group (G6) showing normal eosinophilic granular cytoplasm with central nucleus (x1000). (F) Single hepatocyte from GNPs+APAP treated group (G6) showing minor vacuoles in cytoplasm (x1000). All slides stained with H&E.	74

Appendix		
Fig (1)	Liver of rat from APAP- treated group (group4, 6 weeks) showing vacuolar	103
	degeneration in hepatic cytoplasm and congestion of blood sinusoids (H&E X 400).	
Fig (2)	Liver of rat from APAP- treated group (group4, 6 weeks) showing vacuolar	103
	degeneration of hepatocyte, massive infiltration of inflammatory cells and	
	congestion of blood vessels (H&E X 400).	
Fig (3)	Liver of rat from APAP- treated group (group4, 6 weeks) showing focal	104
	area of necrosis with infiltration of inflammatory cells, necrobiotic changes	
	in sporadic cells (arrows) and vacuolization in hepatic cytoplasm (H&E X 400).	
Fig (4)	Liver of rat from APAP- treated group (group4, 6 weeks) showing	104
	hepatocellular necrosis and vacuolization in cytoplasm with macrophages	
	infiltration associated with sinusoidal dilatation (H&E X 400).	
Fig (5)	Liver of rat from APAP- treated group (group4, 6 weeks) showing	105
	centrilobular necrosis replaced by inflammatory cells in addition to vacuolar	
F: (6)	degeneration of some hepatocytes (H&E X 400).	107
Fig (6)	Liver of rat from APAP- treated group (group4, 6 weeks) showing congestion in portal area with infiltration of inflammatory cells (H&E X	105
	200).	
Fig (7)	Liver of rat from APAP- treated group (group4, 12 weeks) showing fatty	106
	degeneration and congestion of blood sinusoids (H&E X 400).	
Fig (8)	Liver of rat from APAP- treated group (group4, 12 weeks) showing sever	106
	necrosis with infiltration of inflammatory cells and sinusoidal dilatation	
F' (0)	(H&E X 100).	107
Fig (9)	Liver of rat from APAP- treated group (group4, 12 weeks) showing sever congestion in portal area with edema and infiltration of inflammatory cells	107
	and appearance of newly formed bile ducts (H&E X 200).	
Fig (10)		108
	congestion of blood vessels with vacuolation of hepatocytes (H&E X 200).	
Fig (11)	Liver of rat from GE and APAP treated group (group 5, 6 weeks) showing	108
	vacuolation of hepatocytes with infiltration of inflammatory cells, sporadic	
	hepatocytes suffering from necrobiotic changes (arrows) (H&E X 400).	
Fig (12)	Liver of rat from GE and APAP treated group (group 5, 6 weeks) showing	109
	mild congestion in portal area with infiltration of inflammatory cells and	
	vacuolar degeneration in hepatocytes (H&E X 400).	

Fig (13)	Liver of rat from GE and APAP treated group (group 5, 12 weeks) showing congestion of portal blood vessels, perivascular edema associated with infiltration of inflammatory cells and formation of fine fibrous connective tissue (H&E X 200).	109
Fig (14)	Liver of rat from GNPs + APAP treated group (group 6, 6 weeks) showing congestion of blood vessels (H&E X 200).	110
Fig (15)	Liver of rat from GE and APAP treated group (group 6, 6 weeks) showing congestion of portal blood vessels (H&E X 200).	110
Fig (16)	Liver of rat from GE and APAP treated group (group 6, 6 weeks) showing infiltration of mononuclear inflammatory cells between hepatic cords and dilatation of blood sinusoids (H&E X 400).	111
Fig (17)	Liver of rat from GE and APAP treated group (group 6, 6 weeks) showing infiltration of inflammatory cell in portal area and between hepatic cords (H&E X 400).	111
Fig (18)	Liver of rat from GE and APAP treated group (group 6, 12 weeks) showing vacuolar degeneration of hepatocytes and congestion of blood sinusoids (H&E X 400).	112
Fig (19)	Liver of rat from GE and APAP treated group (group 6, 12 weeks) showing vacuolar degeneration and apoptosis of sporadic hepatocytes (H&E X 400).	112
Fig (20)	Kidney of rat from APAP treated group (group 4, 6 weeks) showing congestion of peritubular blood vessels and glomerular tuft with vacuolation of renal tubules epithelium (H&E X 400).	113
Fig (21)	Kidney of rat from APAP treated group (group 4, 6 weeks) showing periglomerular mononuclear cell infiltration and vacuolation of renal tubules (H&E X 100).	113
Fig (22)	Kidney of rat from APAP treated group (group 4, 6weeks) showing some renal tubules suffering from vacuolar degeneration and necrosis (arrow) while the other showing cystic dilatation (H&E X 400).	114
Fig (23)	Kidney of rat from APAP treated group (group 4, 6 weeks) showing focal Interstitial nephritis and degenerative changes of renal tubules (H&E X 100).	114
Fig (24)	Kidney of rat from APAP treated group (group 4, 12 weeks) showing hemorrhage and perivascular edema with inflammatory cells infiltration (H&E X 100).	115